With many thanks to past and present consultants, registrars, nurses and allied health teams who have contributed substantially to this handbook and to the care of babies in the NICU.

Updated 22nd July 2020
Any versions prior to this may not contain accurate information and should be discarded
GLN 0001 (235777)
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**HAND HYGIENE**

- Hand hygiene is the cornerstone of Infection Control programmes and is frequently referred to as the **single most important measure in reducing the transmission of healthcare associated infections.**
- Hand hygiene is a general term that applies to hand washing or use of the alcohol based handrub.
- Hand washing removes blood, body fluids, dirt and transient micro-organisms from the hand using water and either plain or antimicrobial liquid soap (chlorhexidine/green soap).
- Alcohol based hand rubs rapidly destroy microorganisms on the hands and can be used at any time where otherwise hand washing would be carried out but they **must not be used if the hands are visibly soiled or following a procedure where this is likely.**
- The latest approach to hand hygiene is based on the World Health Organisation’s ‘5 moments’ approach:
  - Before patient contact
  - Before a procedure
  - After a procedure or body fluid risk exposure
  - After patient contact
  - After contact with a patient’s surroundings
- For hand hygiene of any type to be effective, the following must apply:
  - The hands and wrists should be **free of jewellery** (with the exception of wedding bands) eg: rings, bracelets, watches.
  - Nails must be short and clean
  - Any cuts/lesions must be covered
RESUSCITATION AND ADMISSION GUIDELINES

Labour Ward Attendance

Policy  A member of the paediatric team will attend high risk deliveries or if problems can be anticipated.

Purpose  To provide optimal and timely resuscitation which can be predicted from characteristics of the pregnancy, labour, or interventions required to effect delivery.

Scope  This policy applies to all medical and midwifery staff including Access Agreement Holders conducting deliveries at Christchurch Women’s Hospital.

- The paediatric team members will either be the Paediatric Registrar or CNS(ANP) on delivery suite duty and contactable on Pager 5019 in the first instance. The Paediatrician on call will be contacted in the event of unavailability due to workload or seriousness of the need.
- The Neonatal Associate Clinical Nurse Manager (ACNM – Pager 5088) is often able to attend deliveries if the RMO / CNS (ANP) is unavailable. They will also accompany the RMO / CNS (ANP) if resuscitation is likely (see charts below). They should be contacted preferably prior to delivery, where admission to the neonatal unit is expected.
- It has been estimated that 2-5% of low risk deliveries may require basic resuscitation at birth but only 0.1% require advanced resuscitation skills such as intubation, cardiac massage and medications. This translates into 120–300 babies requiring basic resuscitation and 6 advanced resuscitation at Christchurch Women’s Hospital. The deliveries identified as high risk or with anticipated problems may include:
  - premature labour < 35 weeks gestation
  - vaginal breech or other malpresentations
  - instrumental deliveries - forceps, ventouse
  - cord prolapse
  - elective caesareans ie those < 39 weeks, infant of a diabetic, maternal complications
  - fetal distress, IUGR
  - multiple births
  - significant fetal anomaly - check for information in the FAAC folder in Neonatal Reception (especially babies with possible surgical problems)
  - meconium liquor
  - maternal chorioamnionitis (any gestation)
  - maternal illness (diabetes, PET, polyhydramnios, oligohydramnios)
- The Neonatal Consultant should be called before delivery when there is a reasonable chance of the baby needing advanced resuscitation. For example:
  - In ALL category 1 caesarean sections (this will be done by Delivery suite Ward Clerk)
  - In other categories of caesarean sections where there are additional factors that increase the chance of advanced resuscitation being required (eg congenital cardiorespiratory anomalies)
  - In vaginal or instrumental deliveries where there is a significant chance of advanced resuscitation being required (eg breech with IUGR)
- Equipment for resuscitation must be checked and ready prior to delivery when anticipated problems exist.
- Other cases which require discussion with the paediatric team prior to delivery:
  - Rhesus iso-immunisation
  - etal anomalies – mild / moderate eg, renal pelvis dilatation > 10mm
  - Possible intra-uterine infection eg CMV, toxoplasmosis, Syphilis serology positive
  - Gestation 35- 36 weeks – attendance at delivery not required unless additional factor.
  - Suspected IUGR or SGA small for gestational age, where BWT < 2.5kg
- Examine all babies whose deliveries you are requested to attend and are actively involved in and document the findings in the 0-2 hour section of the delivery sheet. If an abnormality is present, show it to and discuss it with the parents. Don’t be afraid to tell them frankly if you are not sure of the cause or what to do. If you are uncertain or if a major anomaly is present, consult a senior colleague. Never conceal anomalies from parents.
- Do not do the “24 hour check” at the time of resuscitation as transition will not have occurred and congenital heart conditions will be missed.
- When transporting from the labour ward use the transport incubator. All ventilated infants must be accompanied by medical staff. Tell the parents what you are doing and why. Encourage the father or (with mother’s permission) other support person to accompany you to the unit.
Communication for Neonatal Attendance at Deliveries

- The standard communication is for a Midwife or Obstetric Registrar to phone the Neonatal Registrar or CNS (ANP) with the details of the case and its location. ISBAR should be used to communicate who is calling, what the situation is, the details of the case, location and expectation of timing.
- A ‘777’ call is an emergency call made through the switchboard when neonatal presence is urgently required. This requires the call to be answered and switchboard will pass on the message. The ‘777’ Neonatal team is the RMO / CNS (ANP) and ACNM.
- The Neonatal Consultant will only be automatically called by the Delivery Suite Ward Clerk for ALL Category 1 Caesarean Sections and will be updated by Delivery Suite Coordinator.
- If the Neonatal Consultant is required urgently or non-urgently at any other time they will have to be phoned directly.

Caesarean Section Categorisation

Caesarean sections are categorised according to the Table below. It is no longer acceptable for the phrase ‘emergency’ or ‘crash’ to be used. The Categorisation must be stated in communications.

**Category 1 – threat to the life of the woman or fetus**

**Category 2 – maternal or fetal compromise requiring rapid delivery**

**Category 3 – maternal or fetal clinical situation requiring early delivery**

**Category 4 – delivery at a time to suit the woman and maternity service**

The table below outlines examples of when neonatal attendance is expected at Category 1, 2, 3, 4 sections.

<table>
<thead>
<tr>
<th>Caesarean Classification</th>
<th>Neonatal CNS(ANP)</th>
<th>ACNM</th>
<th>Neonatal Consultant</th>
</tr>
</thead>
<tbody>
<tr>
<td>Any General Anaesthetic</td>
<td>Yes</td>
<td>Yes</td>
<td>Yes</td>
</tr>
</tbody>
</table>

If a credentialled Senior Registrar or CNS/ANP is available and the GA is for indications such as inability to site a spinal or maternal anxiety in the setting of no maternal or fetal distress then they are to discuss the case with the SMO who may not need to attend the delivery.

<table>
<thead>
<tr>
<th>Category 1</th>
<th>Yes</th>
<th>Yes</th>
<th>Yes</th>
</tr>
</thead>
</table>

e.g.: maternal arrest, cord prolapse, uterine scar dehiscence, fetal bradycardia < 100/min for ≥ 5mins, fetal scalp pH < 7.0 and/or lactate ≥ 5.8, placenta praevia and/or major haemorrhage + maternal/fetal compromise

<table>
<thead>
<tr>
<th>Category 2</th>
<th>Yes</th>
<th>Yes</th>
<th>Not routinely but have low threshold for calling</th>
</tr>
</thead>
</table>

e.g.: CTG abnormality (ie: late decelerations) +/- with scalp pH 7.01 – 7.20 and /or lactate 4.8-5.7, breech presentation in active labour unsuitable for vaginal delivery, meconium with fetal distress

<table>
<thead>
<tr>
<th>Category 3</th>
<th>Yes – see below</th>
<th>Not routinely</th>
<th>No</th>
</tr>
</thead>
</table>

- Yes – booked for elective section and present in labour with no risk factors
- No booked for elective section and present in labour with no risk factors

eg: Failed induction of labour presuming indication for induction still exists, pre-eclampsia at term unsuitable for vaginal delivery, suspected IUGR unsuitable for vaginal delivery with normal CTG, delay in progress in labour with no evidence of maternal / fetal compromise, abnormal Dopplers or maternal PET necessitating preterm delivery, APH

<table>
<thead>
<tr>
<th>Category 4</th>
<th>Yes – see below</th>
<th>Not routinely</th>
<th>No</th>
</tr>
</thead>
</table>

- Yes – if no risk factors, singleton, ≥39 weeks
- No if no risk factors, singleton, ≥39 weeks

eg: Congenital abnormality, maternal diabetes, oligohydramnios, polyhydramnios, maternal BMI>30, prolonged rupture of membranes, placenta anterior/accreta/praevia, maternal drug use, breech presentation
# Vaginal Delivery Categorisation

<table>
<thead>
<tr>
<th>Clinical Scenario</th>
<th>Neonatal CNS(ANP) / RMO</th>
<th>ACNM</th>
<th>Neonatal Consultant</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Preterm Singleton Deliveries</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt; 28 weeks</td>
<td>Yes</td>
<td>Yes</td>
<td>Yes</td>
</tr>
<tr>
<td>28 – 34⁺⁶ weeks</td>
<td>Yes</td>
<td>Yes</td>
<td>Neonatal ACNM/RMO/CNS(ANP) to consider calling</td>
</tr>
<tr>
<td>35-37 weeks</td>
<td>Yes</td>
<td>No</td>
<td>No</td>
</tr>
<tr>
<td><strong>Multiple Births</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt; 30 weeks</td>
<td>Yes</td>
<td>Yes</td>
<td>Yes</td>
</tr>
<tr>
<td>30-36 weeks</td>
<td>Yes</td>
<td>Yes</td>
<td>Neonatal ACNM/RMO/CNS(ANP) to consider calling</td>
</tr>
<tr>
<td><strong>Antenatal Diagnoses</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fetal abnormality likely to affect cardiorespiratory condition at birth eg: diaphragmatic hernia or neck mass</td>
<td>Yes</td>
<td>Yes</td>
<td>Yes</td>
</tr>
<tr>
<td>Vaginal breech</td>
<td>Yes</td>
<td>Yes</td>
<td></td>
</tr>
<tr>
<td>Fetal growth restriction &lt; 3% with estimated weight &lt; 2.5 kg</td>
<td>Yes</td>
<td>Not routinely unless CTG abnormal</td>
<td>No</td>
</tr>
<tr>
<td><strong>Perinatal Complication</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cord Prolapse</td>
<td>Yes</td>
<td>Yes</td>
<td>Yes</td>
</tr>
<tr>
<td>Pathological CTG eg: sinusoidal trace, sustained bradycardia &lt; 100 for ≥ 5 mins</td>
<td>Yes</td>
<td>Yes</td>
<td>Yes</td>
</tr>
<tr>
<td>Fetal Scalp pH &lt; 7.0 and/or lactate ≥ 5.8</td>
<td>Yes</td>
<td>Yes</td>
<td>Neonatal ACNM/RMO/CNS(ANP) to consider calling</td>
</tr>
<tr>
<td>Meconium liquor</td>
<td>Yes</td>
<td>Not routinely unless CTG abnormal</td>
<td>No</td>
</tr>
<tr>
<td>Maternal Chorioamnionitis</td>
<td>Yes</td>
<td>No</td>
<td>No</td>
</tr>
<tr>
<td><strong>Instrumental Delivery</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ventouse / Kiwi Cup</td>
<td>Yes</td>
<td>No</td>
<td>No</td>
</tr>
<tr>
<td>Mid cavity forceps</td>
<td>Yes</td>
<td>No</td>
<td>No</td>
</tr>
<tr>
<td>Lift out forceps</td>
<td>Yes</td>
<td>No</td>
<td>No</td>
</tr>
</tbody>
</table>
Preparation for Resuscitation

Notify the consultant on duty of all Level III admissions, plus any others who are significantly unwell, or about whom you have urgent questions.

Good stabilisation is important in the management of all infants and can make a critical difference to a sick or premature baby's subsequent course. However, comprehensive assessment and management must be undertaken, as far as possible, in a way that minimises stress to the baby and facilitates parent/infant contact.

High Risk Deliveries
- At the beginning of the shift, visit labour ward so that you can anticipate high risk deliveries.
- When attending a high risk delivery, ask the ACNM or another neonatal nurse to accompany you.
- Call the consultant if the baby is < 28 weeks, or if there is any other reason to expect that the baby will need a full resuscitation (eg. severe fetal distress, major congenital malformations, significant hydrops etc.) or if you have any other concerns.
- Take surfactant to the delivery of babies < 28 weeks
- Surfactant dosing: 100 mg/kg (1.25 mL/kg) in babies after a complete steroid course (need 3 mL vial if > 1.2kg) and 200 mg/kg (2.5 mL/kg) in babies with incomplete steroid courses (3mL vial needed for most babies).
- Remember to return it to the refrigerator if it is not used as it is VERY expensive.

High Risk Multiple Births
- Multiple births < 30 weeks require additional experienced nursing and medical attendance.
- During the day the Float Registrar/CNS-ANP will discuss the impending delivery with the neonatal consultant on service and the ACNM.
- Staff to attend will be identified so that each baby will have a Registrar/CNS-ANP and a neonatal nurse in addition to the consultant and ACNM.
- The Level 3 staff member will be suggested in the first instance as the second Registrar/CNS-ANP.
- The ACNM will identify nurses with intensive care experience utilising the clinical support nurse and level 3 allocated nurse who will caring for the baby after admission.
- After handover and at the weekend the Retrieval person will be called in to attend these deliveries along with the neonatal consultant
- In the event that they are unavailable attempts will be made to identify a backup by the neonatal consultant.

Preparation
- Maternal history for abnormalities of family history, maternal history, pregnancy, labour and delivery.
- Oxygen supply, Suction
- Laerdal bag and mask 500 mls is on every resuscitaire – 250 ml ones are available for small babies. (works properly, correct size mask, cushion not deflated)
- Laryngoscope (correct blade, good light, clean).
- ET tubes (correct size, open and insert stylet if strong likelihood of intubation)
- Overhead warmer on, 2 warm towels available to dry the baby.
- UVC’s are available in both theatre 26 and 27 resuscitaires and also in the storeroom beside the Delivery Suite handover room for birthing rooms deliveries. UVC’s are not stocked in every room due to supply issues, expiry dates and infrequent use.
- Intraosseous is available and is stored in NICU in the resus trolley
- Draw up resuscitation drugs (1:10,000 adrenaline, normal saline, see later section) and prepare umbilical catheter and call for consultant help if there is a strong likelihood they will be needed (eg. persistent fetal bradycardia before emergency delivery).
- Babies < 30 weeks gestation or estimated to be < 1500gm should be placed in a polyethylene plastic bag at delivery to maintain their temperature. The bag should not be removed for auscultation of the heart or cardiac compressions.
- If using radiant heat source and admission temperature is < 37°C leave in bag, cut a hole in the bag and insert the umbilical lines through this, and then remove bag.
- If using radiant heat source and admission temperature is ≥ 37°C remove bag.
- If using humidified incubator remove bag on admission regardless of admission temperature
Intubation
- Baby less than 28 weeks unless the baby is very active and has minimal or no respiratory distress
- Persistent bradycardia (unresponsive to bag and mask ventilation).
- Inadequate or ineffective respiratory effort after 2 minutes.
- Exposure to thick or particulate meconium, or heavily blood stained amniotic fluid and the baby requires immediate ventilation. Oropharyngeal suctioning at the perineum is no longer in our protocol. If the baby is not active suction by direct visualisation of the laryngopharynx and suction with 10 gauge catheter, meconium aspirator or yanker.
- Respiratory distress in the presence of antenatal or postnatal diagnosis of anomalies (such as congenital diaphragmatic hernia, intestinal atresia, abdominal wall defect) in which bag and mask ventilation is generally contraindicated.

Equipment
- Laerdal bag or Neopuff
- Oxygen, Stethoscope, Pedi-cap
- Appropriate sized face mask
- Suction equipment – size 10 at least
- Cardiac monitor and pulse oximeter for neonatal unit intubations
- Laryngoscope and appropriate blade, with a bright light
- Neofit fixation device
- Introducer or Magill’s forceps

Technique
- Prepare and check equipment
- Note markings on ET tube and how far to insert through cords (trachea length is 2.5 – 5.0cm from 600g – 5.0kg)
- Position baby and have head slightly extended
- Ensure adequate preoxygenation, which may be by bag and mask ventilation
- If an elective procedure give premedication with morphine, atropine, suxamethonium as per drug protocols
- Place laryngoscope by holding it in the left hand with thumb and first two fingers
- Stabilise head with right hand
- Open the infant’s mouth with right index finger
- Introduce blade from right side of mouth, advance to just beyond base of tongue
- Lift blade and observe the landmarks but avoid tilting the laryngoscope
- The epiglottis and cords should come into view
- Cricoid pressure by an assistant may help
- If the laryngoscope is in the oesophagus, withdraw until epiglottis flips into view
- Insert tube through cords into trachea
- An introducer may be necessary to keep the correct curvature on the tube
- Hold tube in place firmly and ventilate
- Check tube position:
  - Observe chest movement
  - Attach pedi-cap and observe for colour change (purple to yellow if in the airway with cardiac output
  - ET tube may mist up
  - Auscultate to assess tube position
  - Observe response of heart rate and saturation
  - Note length of ET tube at lips/nares
- Attach Neofit to secure the tube in place
- CXR to check tube position

Complications
- Hypoxaemia. If it takes too long to intubate or incorrect tube placement. Do not persist with intubation attempts. If heart rate or saturation fall, stop and bag/mask the baby until they recover
- Bradycardia. Either a vagal response or related to hypoxaemia.
- Trauma. Related to laryngoscope or tube being advanced. Can perforate the oesophagus or trachea, or damage vocal cords.
**ET Tube Size** | **Indication** | **Comments**
---|---|---
2.0 | Cannot insert 2.5 ET tube. | Lumen too small to suction down tube, use as a last resort.
2.5 | < 1.5 kg | In general, use a tube that is not a tight fit, to reduce the risk of trauma and later subglottic stenosis, however change to a bigger tube if there is excessive air leak.
3.0 | 1.5 - 2.5 kg | 
3.5 | 2.5 - 4.0 kg | 
4.0 | 4.0 kg | 

**Insertion distance**: because the trachea is short in babies, there is little leeway between a tube that is too short (increased air leak, risk of accidental extubation) and a tube that is too long (bronchial intubation and/or airway trauma). Therefore, it is important to get used to judging correct insertion distance. It is rare to need to insert a tube more than 1 to 1.5 cm below the vocal cords (the exceptions being such situations as trachea-oesophageal fistula or other subglottic anomaly, and the occasional baby in whom elective ventilation of one lung is indicated). The black segment at the tip of the tube is 1 cm long, so that should just disappear below the cords.

The following is a rough guide to tube insertion distance, but the exact distance will need to be individualised because of variation in jaw size, so always check the breath sounds immediately, and request a radiograph promptly. Weight (over 0.5kg) + 6 in cm is also a good guide.

<table>
<thead>
<tr>
<th>Gestation Age</th>
<th>Weight (kg)</th>
<th>ET Tip to Lip (cm) (Oral)</th>
<th>ET Tip to Nostril (cm) (Nasal)</th>
<th>ET size (mm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>24-25 weeks</td>
<td>0.75</td>
<td>6.5</td>
<td>7.0</td>
<td>2.5</td>
</tr>
<tr>
<td>26-28 weeks</td>
<td>1.0</td>
<td>7.0</td>
<td>8.0</td>
<td>2.5</td>
</tr>
<tr>
<td>29-32 weeks</td>
<td>1.5</td>
<td>7.5</td>
<td>8.5</td>
<td>3.0</td>
</tr>
<tr>
<td>33-34 weeks</td>
<td>2.0</td>
<td>8.0</td>
<td>9.0</td>
<td>3.0</td>
</tr>
<tr>
<td>35-36 weeks</td>
<td>2.5</td>
<td>8.5</td>
<td>9.5</td>
<td>3.0</td>
</tr>
<tr>
<td>37-39 weeks</td>
<td>3.0</td>
<td>9.0</td>
<td>10.0</td>
<td>3.5</td>
</tr>
<tr>
<td>40-42 weeks</td>
<td>3.5</td>
<td>9.5</td>
<td>11.0</td>
<td>3.5</td>
</tr>
</tbody>
</table>

**Confirmation of Tube Position**
- Chest movement with assisted breaths
- Breath sounds audible in axilla bilaterally with assisted breaths
- ET tube mists with breaths
- Pedi-cap on the end of the ET tube changes colour from purple to yellow but not if cardiac output is low
- Improvement in heart rate and saturations
- CXR – it is important that a double check of the ET tube placement is undertaken by a Consultant and Registrar or CNS(ANP). This should be done individually. If in doubt a radiology registrar is available on site between 1700 and 0800 and a radiologist is at the CWH reporting desk between 0800 and 1700.

**Neopuff**

The neopuff resuscitator can be used as a ventilation device or be used to deliver CPAP to a baby once an infant has established respirations. It can allow delivery of CPAP from birth rather than waiting until admission to NICU.

- **Gas source**
  - desired flow rate is 8-10 l/min.
- **Maximum pressure relief**
  - allows maximum PIP to be preset. To set, occlude the PEEP cap, turn PIP control clockwise.
  - adjust the maximum pressure control knob to desired pressure eg. 35mmHg.
- **Inspiratory pressure control**
  - allows you to ventilate at a preset PIP by occluding the PEEP cap for the desired inspiratory time.
- **Gas outlet**
  - gas exits the neopuff through attached tubing to a T piece that has the PEEP cap. This can connect to a mask or ETT.
- **T-piece PEEP Cap**
  - the cap can be turned to deliver the prescribed PEEP.
- **To deliver PEEP**
  - place mask over the face with PEEP cap uncovered.
- **To ventilate**
  - attach T-piece PEEP cap to ET or mask. Occlude PEEP cap to deliver PIP at desired rate and inspiratory time.
- **To deliver facial O₂**
  - loosely place mask near face so gas escapes around the edges
Meconium Aspiration

- Meconium stained amniotic fluid (MSAF) comes in various consistencies – thin, moderate and thick.
- Thin occurs in 50-60% of MSAF deliveries and carries a 2-3% risk of needing some respiratory assistance, with 1% needing ventilation.
- Thick occurs in 15% and 25-30% require significant resuscitation and 10% on going respiratory support.

Risk Factors for Meconium Aspiration Syndrome

- Primiparity
- Pacific Island ethnicity
- Male gender
- Low cord pH
- Abnormal CTG (if severe)
- Thick meconium.

8-12% of deliveries are complicated by meconium stained liquor.
In a study by Wiswell 2000, 7% developed respiratory distress of which 3% had MAS and 4% other disorders.

Management of Meconium Stained in the Liquor

- Babies born through meconium stained fluid do not need the mouth and nose suctioned while the head is on the perineum.
- If the baby is flat then the obstetric/midwifery team would stimulate the baby.
- There is no requirement to routinely suction below the vocal cords in an infant with meconium aspiration who is apnoeic and flat. In experienced hands airway suction can be done swiftly without causing prolonged hypoxia but if the resuscitator is not experienced at intubation then they should proceed immediately to bag/mask ventilation and normal resuscitation guidelines (NLS Guidelines 2016).
- If the baby shows signs that the nasal passages or airway are blocked and this is impeding the resuscitation then suction of the nose and mouth and below the cords should be performed at that stage.
- If the baby is vigorous then usual cares should be given.

Cord Gases and Placental Pathology

- Ask for a cord pH if a baby requires active resuscitation with bag and mask, fetal distress on CTG, meconium stained liquor, APGAR < 6 at 5 minutes.
- A segment of cord can be clamped at each end and the blood for this drawn up to 20 minutes later.
- It is the joint responsibility of Obstetrics and Neonates to remember to request a cord gas in emergency situations.
- Ask for the placenta to be sent to pathology for:
  - all cases of suspected intrauterine infection or chorioamnionitis,
  - prolonged rupture of membranes,
  - placental anomalies,
  - all babies in whom a significant resuscitation occurs,
  - when perinatal death occurs and
  - any baby with significant IUGR.

Asking for some cord blood to be saved in a green top tube for later serology or other studies can also be useful.

Drugs for Resuscitation

- Adrenaline is the first choice drug in a newborn resuscitation after establishing effective ventilation.
  - External cardiac massage (ECM) for 1 minute is required to distribute it to the heart.
  - Repeat doses every 3 minutes.
  - Give by ETT, if intubated, until an umbilical venous line is inserted.
- Sodium bicarbonate 8.4% requires dilution with water in equal parts to give 4.2%.
  - Use in prolonged resuscitation (>15 minutes) after effective ventilation/ECM is established.
- Note that naloxone was removed from the resus drug inventory in 2018. However, if considered do not give to the baby if the mother is opioid dependant eg: on methadone.
# Neonatal Resuscitation Drug Dosing

<table>
<thead>
<tr>
<th>Weight</th>
<th>0.5 kg</th>
<th>1 kg</th>
<th>1.5 kg</th>
<th>2 kg</th>
<th>2.5 kg</th>
<th>3 kg</th>
<th>3.5 kg</th>
<th>4 kg</th>
<th>4.5 kg</th>
<th>5 kg</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>ETT Adrenaline 1:10,000</strong></td>
<td>1.0 mL/kg</td>
<td>0.5 mL</td>
<td>1 mL</td>
<td>1.5 mL</td>
<td>2 mL</td>
<td>2.5 mL</td>
<td>3 mL</td>
<td>3.5 mL</td>
<td>4 mL</td>
<td>4.5 mL</td>
</tr>
<tr>
<td><strong>IV Adrenaline 1:10,000</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Dose 1 via UVC (then dose every 3 mins)</td>
<td>0.1 mL/kg</td>
<td>0.05 mL</td>
<td>0.1 mL</td>
<td>0.15 mL</td>
<td>0.2 mL</td>
<td>0.25 mL</td>
<td>0.3 mL</td>
<td>0.35 mL</td>
<td>0.4 mL</td>
<td>0.45 mL</td>
</tr>
<tr>
<td>Further doses via UVC</td>
<td>0.3 mL/kg</td>
<td>0.15 mL</td>
<td>0.3 mL</td>
<td>0.45 mL</td>
<td>0.6 mL</td>
<td>0.75 mL</td>
<td>0.9 mL</td>
<td>1.05 mL</td>
<td>1.2 mL</td>
<td>1.35 mL</td>
</tr>
<tr>
<td><strong>Volume IV over 5-10 min</strong></td>
<td>10 mL/kg</td>
<td>5 mL</td>
<td>10 mL</td>
<td>15 mL</td>
<td>20 mL</td>
<td>25 mL</td>
<td>30 mL</td>
<td>35 mL</td>
<td>40 mL</td>
<td>45 mL</td>
</tr>
<tr>
<td>0.9% saline, whole blood, 4% albumin</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Glucose 10% iv bolus</strong></td>
<td>2 mL/kg</td>
<td>1 mL</td>
<td>2 mL</td>
<td>3 mL</td>
<td>4 mL</td>
<td>5 mL</td>
<td>6 mL</td>
<td>7 mL</td>
<td>8 mL</td>
<td>9 mL</td>
</tr>
<tr>
<td><strong>Sodium Bicarbonate 4.2% slow IV push</strong></td>
<td>2 mL/kg</td>
<td>1 mL</td>
<td>2 mL</td>
<td>3 mL</td>
<td>4 mL</td>
<td>5 mL</td>
<td>6 mL</td>
<td>7 mL</td>
<td>8 mL</td>
<td>9 mL</td>
</tr>
<tr>
<td>Dilute 8.4% with equal volume of water</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>If prolonged resus but effective airway and CPR</td>
<td></td>
<td></td>
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</tr>
</tbody>
</table>
Term (≥ 37 weeks) Newborn Resuscitation Algorithm

CALL for HELP at any time

Birth
- Breathing or crying
- Good muscle tone

YES
- Dry
- Warm – skin/skin
- Observe colour and breathing

NO
- Dry and provide warmth
- Assess breathing, heart rate, colour and tone

Meeting criteria for PPV – no regular respiratory effort, gasping or HR < 100

YES
- Check head position
- Give positive pressure ventilation in air: 5 inflation breaths then ventilation breaths for 30 seconds

NO
- Assess chest movement and heart rate response
- Attach oximeter to right hand or wrist and then turn on

Continuously reassess – at least every 60 seconds

- HR > 100 bpm
- Breathing regularly

If respiratory distress give CPAP in air

Increase oxygen by 10-20% every 30 seconds to meet target saturations

- HR 60 bpm
- Apnoeic or irregular respiration

Continue IPPV and increase oxygen by 10-20% every 30 seconds to meet target saturations

If CPR inadequate, check seal and position, increase PIP, consider intubation

If HR < 60 bpm start chest compressions at 3:1 with lung inflation – oxygen to 100%

If no improvement intubate, give adrenaline via ETT, site UVC, UVC adrenaline, consider volume expansion

Reduction target SpO2

1 min 60-70%
2 min 65-85%
3 min 70-90%
4 min 75-90%
5 min 80-90%
10 min 85-90%
**Preterm (< 37 weeks) Newborn Resuscitation Algorithm**

**CALL for HELP at any time**

1. Birth
   - If < 30 weeks or 1500 g place in plastic bag feet first under radiant heater (set to max), do not cover bag with towels
   - Otherwise dry, remove wet towel and warm as for term infant

2. Assess breathing, heart rate, colour and tone

**Meeting criteria for PPV – inadequate respiratory effort, apnoea, gasping or HR < 100**

- Check head position
- Give positive pressure ventilation in 30% oxygen: 5 inflation breaths and then ventilation breaths for 30 seconds

3. Assess breathing, heart rate, colour and tone

4. Assess chest movement and heart rate response
   - Attach oximeter to right hand or wrist and then turn on

5. If infant < 28 weeks consider intubation to give surfactant

**Continuously reassess – at least every 60 seconds**

- HR > 100 bpm
- Breathing regularly

If respiratory distress or cyanosis give CPAP in 30% oxygen

Alter oxygen by 10-20% every 30 seconds to meet target saturations

- HR > 80 but < 100 bpm
- Apnoeic or irregular respiration

Continue IPPV and alter oxygen by 10-20% every 30 seconds to meet target saturations. Consider intubation

If HR < 60 bpm
- Centrally blue or white
- Apnoeic or gasping

If chest movement is inadequate, check seal and position, increase PIP, consider intubation

If HR < 60 bpm
- Start chest compressions at 3:1 with lung inflation – oxygen to 100%

If no improvement intubate, give adrenaline via ETT, site UVC, UVC adrenaline, consider volume expansion

**TARGET SpO2**
- 1 min: 60-70%
- 2 min: 65-85%
- 3 min: 70-90%
- 4 min: 75-90%
- 5 min: 80-90%
- 10 min: 85-90%

**Reduce oxygen by 10-20% every 30 seconds to meet target saturation**

*Authorized by Clinical Director Neonatal Services*
*Ref: 237134 July 2016*
Admission Guidelines

Level 3
- Respiratory support required
- Concern that infant is moderately unwell

Level 2
- Weight < 2.3kg
- No respiratory support required
- If there is a concern that the baby is unwell then they should not be admitted to Room 6
- For infants needing oxygen, antibiotics, iv fluids, thermoregulation, phototherapy, apnea monitoring, tube feeding
- Infants at risk of neonatal abstinence syndrome
- MIPS mother’s stay is usually continuous until discharge unless the baby is preterm or not orally feeding.
- One of the intentions is to maintain mother-infant contact, mothers may be admitted to the parent facility once discharged from postnatal. If beds are short then priority is given to mothers establishing feeding or within 48 hours of discharge.

Admission Routine

- Weight, length and head circumference
- If the baby is in a polyethylene bag, and intubated then take off 10g for the actual birthweight
  - Polyethylene bag = 5g
  - ETT 2.5 = 3g
  - Neofit = 3g
  - No need to take off the umbilical clamp weight as it is standard for that to be included
  - If using a Neohelp bag with the hood take off 30g if in a <1000g bag, or,
  - 40g if in the 1000-2500g bag

- Plot growth parameters on the centile charts
- Physical examination.
- Temperature and blood sugar

Unwell babies may also require:
- Blood Pressure
- Blood Gases
- Blood Cultures
- Full Blood Count with differential
- CRP – at least 6 hours after onset of illness/birth
- Bag urine/BPU/Catheter urine whenever there is deemed to be significant risk of sepsis for GBS Ag
- Group and Coombs
- Group and hold (on admission for infants < 30 weeks and others likely to need transfusion at any stage)
- Lumbar puncture (this can be deferred in the presence of severe or unstable respiratory distress or in situations where the risk of sepsis is deemed very low -- if in doubt discuss with consultant)
- Intravenous line and commencement of 10% dextrose
- Antibiotic therapy as indicated

Very small and/or seriously ill babies, including those who will need continuous monitoring of blood pressure and frequent sampling for blood gases, biochemistry etc. should also have:
- Umbilical or peripheral arterial line
- Umbilical venous catheter
- Treatment of hypotension (saline, packed red cells, dopamine or dobutamine infusion, second line treatment includes albumin or hydrocortisone)
- CXR/AXR to check position of UAC, UVC and ET tube
- Again ..... remember to explain to the parents what is happening.
Vitamin K

Haemorrhagic disease affects one in 2-400 babies who are not given vitamin K prophylaxis. Recommendations are:

- All infants should have vitamin K prophylaxis (2mg/0.2ml)
- Vitamin K given either as a single IM injection or as repeated oral doses is safe and effective in preventing haemorrhagic disease of the newborn in well newborns.
- Parents should be free to choose, either an injection or oral doses for their child.
- Oral vitamin K is given as 2mg at birth with the first feed.
- For breastfed infants repeat doses (2mg) should be given at 5 days (at the time of the Guthrie test) and at 6 weeks (with the first immunisation).
- IM vitamin K is given as a single injection of 1mg (0.1ml) at birth (< 1500 g 0.5 mg =0.05ml).
- IM vitamin K is the strongly recommended route for high risk infants (prematurity, birth asphyxia, traumatic deliveries, known hepatic disease, or any illness that will delay feeding, eg most NICU admissions.
- Maternal phenytoin, primidone, methsuximide or phenobarbitone therapy is an indication for the mother to be given vitamin K (10 mg IM vitamin K) 24 hours before delivery then the baby should have 1 mg IM vitamin K at birth then again 24 hours later.
- Usually maternal warfarin therapy is stopped well before delivery because it crosses the placenta and can cause severe neonatal haemorrhagic disease. Occasionally, it cannot be stopped for maternal reasons. In these circumstances, the baby will need immediate and usually repeated doses of vitamin K, as well as measurement of PT and PTT and fresh frozen plasma on standby.

Admissions to the Neonatal Unit in the first month after birth (or corrected age)

This document is to assist the clinical teams to consider whether an ill infant who presents aged less than one month (corrected age) needs admission to the neonatal unit.

Good communication between GP/Midwife, Paediatric, Paediatric Surgery and Neonatal staff from both the referring and receiving hospitals is essential to ensure the baby is cared for in the appropriate place. Consultant to consultant discussion facilitates appropriate and timely decisions.

The most common pathway for presentation is via GP/LMC/Ambulance/ED to Paediatrics / CAA. Less frequently, the neonatal team or the paediatric surgeons are contacted directly.

For the specific indications outlined below, neonatal (re)admission can be arranged. Generally, this is for:

1. Infants up to one month of age born at term,
2. Infants less than one month corrected age if born preterm,
   AND
3. Infants with "neonatal" conditions as outlined below that require intensive care.

There may be a place for stabilising an infant in NICU prior to retrieval to PICU Starship, or if a short stay is predicted until they are transferred back to PHDU.

Involvement of the ACNM (neonatal co-ordinator pager 5088) is needed early, particularly if the neonatal transport team is required.

Admission to the neonatal unit is dependent on available cot space; sufficient staff and specific requirements (for example, some need isolation).

A neonate who ideally should be admitted to the NICU but is not able to be accommodated there, should be reviewed by the Neonatologist at the time of admission. The neonatologist can provide advice to the admitting paediatrician and surgeon re ongoing management and will review the patient on a daily basis in consultation with the paediatric team (and paediatric surgeons, as required).
Conditions and Severity Which May Need Admission to NICU

Jaundiced babies
- Babies with an SBR above the threshold for an exchange transfusion as NICU has the skills and equipment to perform exchange transfusion, if needed
- Babies with an SBR >400umol/L after day 3 requiring intensive phototherapy

Surgical babies
- Presentations may include Hirschsprung disease, malrotation with volvulus, bowel obstruction causing ischaemia, and some patients with strangulated inguinal hernias or pyloric stenosis.
- Ex-very preterm infants with major co-morbidity, such as those with chronic lung disease, growth failure < 3rd %.
- The need for post-operative respiratory support.
- The need for TPN post-operatively (or at any time)

Neonates (<1mth corrected) needing surgery must have a surgeon and a paediatrician or neonatologist involved in their care.
- The surgeons will inform the neonatal consultant on service as soon as they accept a referral from another DHB who meet the criteria for NICU Admission Above or need Neonatal transport
  - to ensure there is a NICU bed available, the appropriate mode of transport and a transport team available.
- The surgeons will inform the Paediatrician of the day as soon as they accept a referral from another DHB for infants not needing NICU.
- Discussion between the teams will decide on the appropriate mode of transfer of the baby (if from another centre) and the most appropriate place for the neonate to be admitted to.
- Where transport is need the neonatal ACNM pager 5088 should be contacted ASAP.
- Transport of a neonate who may be NBM for >4hrs requires iv access and a neonatal transport team

Respiratory support
- It is more appropriate for neonates with suspected infections and respiratory symptoms such as RSV and pertussis to be admitted to Paediatrics / HDU to prevent the spread of infectious diseases through the neonatal nursery.
- However, if there is a need for escalating respiratory support such as prolonged CPAP or the possibility of short-term ventilation and the isolation room (Room 2) is available then an admission is possible to prevent a transport to Starship
- Neonates with severe seizures due to trauma, meningitis or encephalitis may also require respiratory support in NICU
- Transfer of neonates from paediatrics to the neonatal unit and vice versa requires consultant to consultant discussion.

Cardiac
- Neonates with cyanotic congenital heart disease are best cared for in NICU due to the monitoring needed, possible requirement of prostaglandin infusions and the potential need for intubation and transport to Starship
- Diagnosis by echo is required
- A neonatal echo roster is available by contacting neonatal reception on phone 85885. Alternatively the Cardiology technicians will perform the echo.

Transport of babies to weight 5.5kg.
- The neonatal service has a registrar / CNS advanced -rostered 24/7.
- A transport nurse is available from the neonatal unit staff
- The retrieval team takes 20-30 minutes to assemble
- Transportation is by ambulance for Christchurch units (Burwood, St Georges, Rangiora, Lincoln, Darfield) and Ashburton.
- For Timaru transport is either by ambulance – low risk, or helicopter – high risk, where a shorter time to get to CHCH is important due to clinical condition.
- West Coast – usually fixed wing, occasionally helicopter.
- Weather factors determine the use of fixed wing and at times this requires utilising teams from Dunedin, Wgtn
- For medical and surgical transfers > 1 month the transport service is under-resourced. If the neonatal team is unable to assist calling in Paediatric registrars or Paediatricians is dependent on goodwill and availability.
Small Baby Protocol – Guidelines for the < 28 Week Neonate

- One of the most challenging aspects of perinatal medicine is the management of the delivery of an extremely premature infant.
- Care of the mother, her fetus and the baby will always need to be individualised.
- The objective of this protocol is to optimise the management of the extremely preterm infant born at < 28 weeks gestation.

### Number of babies < 28 weeks admitted to CWH NICU over past 10 years

<table>
<thead>
<tr>
<th>Time Period</th>
<th>Number Admitted Mean and Range</th>
<th>Survival to Discharge Mean</th>
</tr>
</thead>
<tbody>
<tr>
<td>2006-2015</td>
<td>25 (22-31)</td>
<td>85%</td>
</tr>
<tr>
<td>2015-2019</td>
<td>28 (23-34)</td>
<td>85%</td>
</tr>
</tbody>
</table>

**Antenatal Care**

Parental hopes and expectations need to be explored with honesty and compassion in a realistic manner, drawing upon available local and national population data.

Communication and agreed plans must be documented in full and signed legibly.

These plans may need to be revised frequently based on the best assessment of gestational age, in addition to information regarding the wellbeing of the fetus and parental wishes.

**Steroids**

- The benefits of antenatal administration of corticosteroids to fetuses at risk of preterm delivery vastly outweigh the potential risks. Benefits include not only a reduction in the risk of RDS but also a substantial reduction in mortality and IVH.
- Treatment consists of two doses of 11.4 mg of betamethasone given intramuscularly 24 hours apart
- No additional benefit has been demonstrated by accelerating the course and giving the second steroid dose at 12 hours instead of at 24 hours
- The effects of steroids last for approximately 7 days after the last dose. Repeated courses are not routinely given. If 10 days have lapsed and a woman remains at risk of giving birth within 48 hours, a single dose of steroids should be considered.
- Steroids should be discussed with the neonatal consultant prior to administration if < 24 weeks gestation.

**Antibiotics**

- Preterm rupture of membranes and not in labour – oral erythromycin 10 days.
- As premature infants are at risk of E Coli and Group B strep sepsis, antibiotics are given to all women who are in established preterm labour.
- When in labour – IV amoxicillin and gentamicin
- If signs of maternal sepsis – IV amoxicillin, gentamicin and metronidazole

**Other**

- Magnesium sulphate should be given for neuroprotection in babies <30 weeks as standard management. This is given as a loading dose over 20 minutes and then a maintenance infusion.
- Offer enrollment in relevant research trials if appropriate.

**Resuscitation Considerations**

Preterm labour often progresses rapidly and in these circumstances there may be insufficient time to hold a detailed discussion with the parents before the baby is born. A decision about resuscitation may need to be made based on the most recent management plan, if any, and the available clinical information.

These guidelines apply to singleton pregnancies and have been adapted from the British Association of Perinatal Medicine¹
< 23° weeks
- Babies very rarely survive at this gestation and in favour of the best interests of the baby and standard neonatal practice; it is acceptable for resuscitation not to be commenced

23° to 24+6 weeks
- At these gestations babies are at the limits of viability.
- Each case needs to be individualised, accounting for parents wishes and the clinical situation.
- In 2019 a New Zealand consensus statement on the care of the mother and baby/ies at periviable gestations which covers antenatal advice and counselling and intrapartum care advice.
- As part of this consensus there is a Parent Information and Decision Aid tool to help share decision making and discussions with the family.
- There is also material to help health professionals with counselling. Periviability information for health professionals.
- A decision not to start resuscitation is an appropriate approach if the parents have expressed this wish after discussion with the neonatal team.
- Clinical information to be considered when discussing resuscitation include the gender of the baby, steroid coverage, presence of sepsis and any known congenital abnormalities.
- If resuscitation is started with lung inflation, the response of the heart rate will be critical in deciding whether to continue or to stop resuscitation.
- It is usually not appropriate to provide cardiac compressions or give adrenaline at these gestations.

≥ 25 weeks
- It is usually considered appropriate to resuscitate babies at this gestation unless there any underlying congenital factors.

Multiple pregnancies
- Fetal maturity is decreased and it may not be appropriate to resuscitate multiple births at < 25 weeks gestation.

Neonatal Resuscitation Preparation
In addition to routine equipment / procedures for the management of the high risk infant, resuscitation and stabilisation of < 28 weeks gestation infant includes:
- Calling the consultant. If possible 2 consultants should be present at births of multiples <26 weeks.
- Ask the ACNM or senior nurse to accompany you.
- ACNM to prepare the humidification system for Neopuff or ventilation.
- Surfactant –100mg/kg (1.25mL/kg) in babies after a complete steroid course (3mL vial needed if >1.2kg) and 200mg/kg (2.5mL/kg) in babies with incomplete steroid courses (3mL vial needed for most babies). Return it to the refrigerator if it is not used.
- Size 5F feeding tube for administration of surfactant.
- Blade or sterile scissors.
- Paper tape measure.
- Plastic bag on resuscitaire to pre-warm, pre-cut right hand bottom corner, if Neohelp then no cut is needed.
- Transport incubator.
- Laryngoscope blade 00 / 0.
- 2.5 ET Tube with stylet inserted if required.
- Delegate tasks so everyone knows their responsibility at delivery including documentation.

Obstetric Resuscitation Preparation
- Place sterile cord clamp on the cord 6cm from base to allow placement of the baby into plastic bag.
- Deferred cord clamping for at least 60 seconds if appropriate.
Care at Birth

As part of the National consensus of the care of babies at the limits of viability there is a NZ practice recommendation for the bundle of neonatal care at 23-24-weeks that can also be used as a reference tool.

Thermal Management

Optimal thermal management of preterm infants is a problematic but important aspect of care.

Low birth weight infants are vulnerable to hypothermia (≤36.4°C) since they have impaired ability to prevent heat loss and decreased heat production capability.

Delivery suites and operating theatres are generally cold environments (<22°C) and can cause thermal stress for the newborn which is associated with increased mortality and morbidity. A warm environment for a neonate would be 24-26°C.

After delivery

- Infant is transferred to the resuscitaire and placed in a polyethylene plastic bag or Neohelp bag under radiant heater (set to maximum)
- Do not cover bag with towels as it impedes radiant heat
- Only dry face and head
- Cover head with a hat
- Assess breathing, heart rate, colour, tone and perform resuscitation as per neonatal guidelines (see algorithm below)
- Insert pulse oximeter through the hole in the bag and apply to right hand and then turn it on to obtain pre-ductal saturations and heart rate
- Do not lower the bag to auscultate heart rate

Airway

Ventilation

- Aim for a PEEP of around 5cm H20
- Use adequate PIP to ensure adequate chest wall movement
- Aim to ventilate at about a rate of 50-60/min
- Use humidified set-up

Intubation

- Consider intubation if the infant is not very active, has not had a complete steroid course and has apnoea, increased work of breathing or oxygen requirement >30%
- Once intubated, most very small infants respond rapidly to IPPV

<table>
<thead>
<tr>
<th>Gestation</th>
<th>Oral ET Tip to Lip (cm)</th>
<th>Nasal ET Tip to Nostril (cm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>24-25 weeks</td>
<td>6.5</td>
<td>7.0</td>
</tr>
<tr>
<td>26-28 weeks</td>
<td>7.0</td>
<td>8.0</td>
</tr>
</tbody>
</table>

Following intubation confirm ETT position:

- Chest movement with assisted breaths
- Breath sounds – audible in axilla bilaterally with assisted breaths
- Change in Pedi-cap colour from purple to yellow except when cardiac output is poor
- Improvement in heart rate and saturations
- Secure ETT prior to giving surfactant with a Neofit (if using a 2.5mm ETT a strip of brown tape around the tube and neofit is required to ensure the tube does not slip)

<table>
<thead>
<tr>
<th>Target SpO2</th>
<th>2 min</th>
<th>3 min</th>
<th>5 min</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>65-70%</td>
<td>70-75%</td>
<td>80-85%</td>
</tr>
</tbody>
</table>
Oxygen saturations
- Commence IPPV in 30% oxygen
- Reduce/increase oxygen by 10–20% every 30-60 seconds to meet target saturation
- If CPR required increase oxygen to 100%

Pre-Term (< 37 weeks) Newborn Resuscitation Algorithm
- See algorithm above

Drugs and Administration

Curosurf
- Give in delivery room immediately after delivery via ETT following discussion with consultant
- Dose 1.25–2.5ml/kg (100–200mg/kg). See drug protocol for full details
- Draw up the volume required in a syringe and place on the end of nasogastric tube cut to the length of the ETT (any longer and the surfactant will only go into the right lung)
- During administration check chest movement and be prepared to increase the PIP/oxygen
- Following administration check chest movement and be prepared to reduce the PIP/oxygen

UVC Insertion
- Insert UVC as cleanly as possible (this cannot be a strictly sterile procedure in an emergency situation)
- Insert to 3-5cm and tape to the abdomen

Adrenaline
- Adrenaline is the first choice drug in a newborn resuscitation after establishing effective ventilation
- In order to distribute it to the heart cardiac massage is required for 1 minute.
- ETT Dose: 1:10,000 1ml/kg
- UVC Dose: 1:10,000 0.1ml/kg initially then up to 0.3ml/kg
- Doses can be given every 3 minutes – intravenous is the preferred route

Volume expansion
- Normal saline - readily available
- Whole blood - urgently requested from Blood Bank
- Dose: 10ml/kg - give over 5-10 mins

Sodium Bicarbonate 4.2%
- This is rarely required
- Dilute 8.4% with equal volume of water to make a 4.2% solution
- Dose is 2ml/kg of 4.2% solution given over 2mins
- Administer in prolong resuscitation (>15minutes) after effective ventilation/cardiac massage

Glucose
- 10% Glucose, dose is 2ml/kg
- Consider if resuscitation has been prolonged and only after documented hypoglycaemia

Vitamin K
- IM, dose for those <1500g is 0.5mg = 0.05ml

Cord Gases and Placental Pathology
- Request a cord pH and lactate (preferred test if both cannot be done)
- Send the placenta for pathology

Transfer to NICU when infant is stabilised
- Notify NICU, update parents
- Ensure ETT is secure, leave infant in polyethylene bag and nest in warm towels in incubator
- Check exhaled tidal volumes on the Crossvent ventilator to ensure that the baby is not being over ventilated after receiving surfactant
- Place the name band supplied in delivery suite and when in NICU replace with a posy name band
Care in the Golden Hour

Sick newborns do not tolerate handling or hypothermia. Minimal handling and maintenance of normal temperature are paramount.

The aim should be to undertake all nursing and medical procedures as quickly and with as little disturbance to the infant as possible, usually within 1 hr of admission (golden hour) 4

Thermoregulation / Humidification

High transepidermal water losses can be reduced by increasing the relative humidity of the infant’s environment

- On admission weigh infant in the polyethylene bag and admit into Giraffe incubator (preferred)
- If the baby is in a Neohelp bag then take off 30g for the birthweight if in the <1000g bag or 40g if in the 1000-2500g bag
- If the baby is in a Polyethylene bag then take off 10g for the birthweight
  - The weight of the ETT and Neofit are negligible and can be ignored if ventilated
  - No need to take off the umbilical cord or clamp weight as it is standard for that to be included
  - Do not take the baby out of the bag and weigh it to get the birthweight as it will be inaccurate
- Weigh infant on Giraffe incubator scales and use these scales for subsequent daily / alternate day weights
- Humidification settings should commence at 80%
- It takes 15 minutes from turning on Giraffe humidification to achieve 80% and takes 1 hour on other incubators so make sure it is turned on so that on admission humidification is underway
- If using radiant heat source and admission temperature is <37°C leave in bag until procedures such as line insertion is completed and then remove bag.
- If using radiant heat source and admission temperature is ≥ 37°C remove bag.
- If using humidified incubator remove bag on admission regardless of admission temperature.
- Place limb band ECG electrodes on infant’s ankles or wrists
- Place a skin temperature probe to avoid extremes in temperature
- Once umbilical lines are inserted and secured apply petroleum jelly to torso (above elbows and knees) Avoid application to face or scalp.

Ventilation (See Neonatal Handbook for full details)

Common starting values are:

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>SIMV or PTV</td>
<td>Inspiratory time 0.34sec</td>
</tr>
<tr>
<td>PIP 16-20cm</td>
<td>Rate 50-60 /min</td>
</tr>
<tr>
<td>PEEP 5cm</td>
<td>Trigger 0.2</td>
</tr>
</tbody>
</table>

There are no magic set of ventilator settings that will be appropriate for all babies

- When an infant is ventilated it is important to constantly monitor the infant’s response including; tidal volumes, chest movement, saturations, blood gases
- Lung compliance can change rapidly following surfactant so need to observe above parameters closely and do a blood gas early to avoid over ventilation
- The second dose of surfactant is determined by CXR and degree of ventilation support required at 12 hours. Consultant’s decision.

Acceptable arterial blood gases:

<table>
<thead>
<tr>
<th>pH</th>
<th>PaCO2</th>
<th>PaO2</th>
</tr>
</thead>
<tbody>
<tr>
<td>7.25 – 7.35</td>
<td>40 -50 mmHg</td>
<td>50- 70 mmHg</td>
</tr>
</tbody>
</table>

Target Oxygen saturations

<table>
<thead>
<tr>
<th>Sats Targets on Oxygen</th>
<th>Alarm Limits on Oxygen</th>
<th>Sats Targets in Air</th>
<th>Alarm Limits in Air</th>
</tr>
</thead>
<tbody>
<tr>
<td>90-92%</td>
<td>88- 94%</td>
<td>90-100%</td>
<td>88 -100%</td>
</tr>
</tbody>
</table>
UAC / UVC line insertion  (See Neonatal Handbook for full details)

- Ensure a second helper is available
- Wear gown, hat, mask and gloves
- Sterile technique
- Use low strength chlorhexidine (0.1%, blue solution) to prevent burning to skin
- Ensure skin is allowed to dry before commencing the procedure
- Place cord tie around stump and cut stump below the cord clamp
- Watch for bleeding and tighten cord tie if bleeding occurs
- 3.5g double lumen for UVC and 2.5 or 3.5g single lumen for UAC
- Once lines are inserted suture in and place supporting flag before x-raying
- Bridging of umbilical lines is not necessary as long as there are well placed flags around the suture material and catheter
- AP and a lateral Xray are required
- Document the type of line that has been placed and how far in
- Document the tip position on Xray and if the line is in a satisfactory position
- Re-Xray if line is manipulated after initial Xray and document tip position
- Use sticker for procedure note

UVC

- Many ways of estimating length – including suggestion below, measure charts, formulas.
- Estimate length by measuring nipple to umbilicus (cm) plus the additional length of the umbilical cord, or see chart below
- Position in the IVC but not in the heart (usually just below the diaphragm is a good guide), any line in the liver should be discussed whether it should be withdrawn to be outside the liver or whether it has passed through the ductus venous and is satisfactory (see procedure section later in the Handbook)

UAC

- Estimate length by measuring shoulder tip to umbilicus or ear to umbilicus (cm) plus the additional length of the umbilical cord, or see chart below
- Position at T6 – T9 or L3 – L4

<table>
<thead>
<tr>
<th>Birth weight (kg)</th>
<th>UAC length (cm)</th>
<th>UVC length (cm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.5</td>
<td>11.0</td>
<td>6.5</td>
</tr>
<tr>
<td>1</td>
<td>12.0</td>
<td>7.0</td>
</tr>
<tr>
<td>1.5</td>
<td>13.5</td>
<td>8.0</td>
</tr>
</tbody>
</table>

Blood Tests  (Take bloods off UAC once inserted)

- ABG / BSL
- FBC/differential
- Blood culture, CRP (after at least 6 hours)
- Group and DAT (write on request form extreme preterm infant – so blood bank will prepare quad pack units of blood)
- Consider coagulation screen if severe chorioamnionitis / sepsis

Record amount of blood withdrawn as accumulative total may be taken into account along with the Hb when deciding on when to transfuse in the first 72 hours.

Fluids  (See Neonatal Handbook for full details)

- TPN Starter bag 90mL/kg/day and Lipid 1g/kg/day
- UAC infusion 0.45% saline plus heparin 1unit/mL
- Record fluid intake and output
- If hypoglycaemic, BSL< 2.0 mmol/L consider 2ml/kg 10% dextrose and repeat BSL in an hour
Drugs
- Amoxicillin and gentamicin/cefotaxime as per neonatal drug protocols
- Take gentamicin levels after the first dose
- Oral nystatin 0.5ml 8 hourly for fungal prophylaxis
- Sedation with morphine should be individualised depending on the babies comfort and likelihood of extubation
- If morphine is required give a loading dose (may not be required if intubation drugs were given recently) of 50 mcg/kg over 30 minutes first before starting an infusion of 10 mcg/kg/hr. May need to be increased to 20mcg/kg/hr if infant very unsettled.
- The need for inotropes will be dictated by the blood pressure and cardiac output assessed with an echocardiogram plus urine output, lactate and perfusion.

Blood Pressure and Cardiac Output
- Central measurement of blood pressure via the UAC is the ideal as manual recordings can be inaccurate.
- Manual recordings at admission will not be needed if a UAC is going to be placed
- In reality we should aim for a MAP that provides adequate blood perfusion to all vital organs and cardiac output also needs to be taken into consideration
- Aiming for a mean arterial pressure (MAP) the same as the gestation is an easy rule and equates to the 10th% for gestation but is only a starting point. The consultant will suggest BP limits.
- Also monitor and document urine output, perfusion, colour, acidosis, lactate
- All ventilated infants < 28 weeks should have an echo in the first 12 hours (earlier if necessary) to assess cardiac contractility, output and PDA

Care in Week One
Keep the family up to date with any changes in clinical condition and when any important tests have been done i.e. head ultrasound or echo. Document when parents updated by medical team.

Ventilation
- Aim to wean ventilation as able.
- Send ETT aspirate for Mycoplasma/Ureaplasma
- In the days prior to extubation and to sometimes facilitate extubation to CPAP give loading dose of Caffeine Citrate 20mg/kg IV followed by maintenance 7.5mg/kg/dose once daily.
- Avoid collapse of alveoli by establishing PEEP via nasal mask or prongs before extubating

Bloods
- NEON at 24hrs of age and then 24 hourly for first few days
- Blood gases between formal NEON can provide 12 hourly electrolyte trends and minimises blood sampling
- Discuss frequency of blood tests with Consultant
- Monitor BSL as hyperglycaemia occurs frequently with the potential of causing an osmotic diuresis with associated weight loss and dehydration.
- Sepsis should also be considered if BSL elevated.

Fluids
- Calculate fluid balance 24 hourly
- Weighing is an accurate way to monitor fluid balance and daily weights may be requested if baby is in Giraffe incubator
- Preterm infants have limited but some urine on D1 with increasing amounts from D2
- High Sodium TPN may be needed from day 5-7 (rapid fall in sodium or levels <135mmol/L)
- In most extreme premature infants after the first few days there is an ongoing loss of sodium due to their inability to reabsorb sodium
- Upon reviewing sodium results need to consider weight and effects of any current medication eg: indomethacin, diuretics.
  - If hypernatremic:
    - Change any infusions made with 0.9% saline to 0.45% saline or dextrose.
    - If need to increase mls/kg/day – increase total TPN volume or commence dextrose sideline
  - If hyperglycaemic:
    - Review glucose intake using glucose calculator in TPN folder (aim for <10mg/kg/min)
    - Review infusions and change any in dextrose to 0.45% or 0.9% saline if able
    - A 7.5% glucose TPN bag is available if sugar control is problematic despite insulin
• **Lipids:**
  - Increase lipids as per neonatal handbook guidelines
  - Decrease lipids to 2g/kg/day when on 80ml/kg/day enteral feeds
  - Stop lipids when on 100ml/kg/day enteral feeds
• If sufficient EBM aim to give trophic feeds 0.5ml - 1ml 4 hourly from D1
• Start probiotics the day after birth (a patient information sheet is available)
• Feeds to increase as per the feeding guideline
• Addition of HMF to feeds needs to be discussed with Consultant and information provided to parents (a parent information sheet is available)
• Start HMF when the baby achieves 100ml/kg/day enteral feeds if no contraindications (see HMF guideline)
• Referral to Infant Feeding specialist with a view to be seen in the weekly clinic

**Drugs**
• Consider hydrocortisone for prevention of bronchopulmonary dysplasia in babies <28 weeks.
  o Emerging evidence that hydrocortisone reduces BPD, death and neurodevelopmental disability at 2 years of age.
  o The effect may be greatest in those <26 wks, those with no/ incomplete antenatal steroids
  o Refer to the drug profile for further information on patient selection and dosing
• If on antibiotics review at 24 hours with the blood culture result and serial CRP and FBC measurements
• Discuss commencing insulin infusion if 2 consecutive BSL>10mmol/ L
• Hyperglycaemia is likely to be related to insulin resistance and poor production but infection should be considered as well 9
• Calculate the glucose load in mg/kg/min to ensure it is not >10mg/kg/min using the Glucose calculator
• Use insulin dosing by the insulin computer
• Caffeine maintenance dose to start at 10mg/kg/day
• Start oral Vitamin A, Vitamin D and Micelle E when lipid stops as per drug protocols
• Start Folic acid if < 1500 gm at birth and are to receive unfortified breastmilk long term
• Continue on nystatin until no longer receiving Level 3 care

**Imaging**
• Further echos will be needed to follow the PDA and assess response to inotropes if required
• Head ultrasound scan – day 1-2 and repeat on Day 7-10

**Longline** *(See Neonatal Handbook for full details)*
• Insert longline if full enteral feeds not likely to be achieved by day 7 as UVC will then need to be removed
• Premicath can be used in infants < 1000gm or in those with difficult veins
• The larger size catheter is preferable in bigger infants as it blocks less often and can tolerate more volume
• Double lumen longlines may be necessary for infants with multiple infusions
• Document which catheter is inserted, how far in and if the position is satisfactory on X-Ray
• If line position is altered re-X-Ray prior to use and re-document
• Use sticker for procedure note
• On confirmation of correct placement on X-Ray remove UVC.

**Appropriate tip position:**
• Upper extremity lines should end in a subclavian vein or SVC (not atrium or ventricle due to risk of myocardial damage)
• Lower extremity lines should preferably end in an iliac vein or IVC
• Lines can migrate inwards so take this into consideration when deciding on whether the line tip will be satisfactory if this occurs

**Humidification**
• 80% humidification should continue until day 7
• Day 8 onwards reduce by 5% daily until at 50% - when humidification can be discontinued
• Apply petroleum jelly every 12 hours up to a maximum of 14 days when stratum corneum is matured 10 11 12
RESPIRATORY SUPPORT

Antenatal Steroids

(Maternity Guideline: Antenatal Corticosteroid Therapy (GLM0065))

- The benefits of antenatal steroids to fetuses at risk of preterm delivery outweigh any potential risks.
- Antenatal steroids decrease neonatal mortality, respiratory distress syndrome, IVH, NEC and early onset sepsis.
- Treatment consists of two doses of 11.4 mg of betamethasone given intramuscularly 24 hours apart.
- Optimal benefit begins 24 hours after completion of the course and lasts 7-10 days.
- Even if delivery seems imminent steroids should still be given as timing of delivery is unpredictable and benefits are still seen after incomplete steroid courses.
- No additional benefit has been demonstrated by accelerating the course and giving the second steroid dose at 12 hours instead of at 24 hours.

Indications for Antenatal Steroids

- **22+5 to 24+6 weeks** – to be given if active intervention at birth is the shared decision after joint consultation with the Obstetric team, Neonatal team and parents.
- **25+0 to 34+6 weeks** – give steroids if at risk of preterm birth within 7 days.
- **< 39 weeks** – steroids to be given prior to elective caesarean section.
- **< 38 weeks** – steroids to be given prior to elective caesarean section in diabetic mothers.

Indications for Repeat Antenatal Steroids

- Give a single repeat dose of betamethasone if:
  - the gestation is <35 weeks
  - at risk of preterm birth within 7 days
  - it is more than 7-10 days from the completion of the last steroid course.
- Up to 3 single repeat doses can be given. If a woman remains at risk of preterm birth after 3 repeat doses a joint discussion should occur between Obstetrics, Neonatal and the parents to weigh up the risk and benefits of further doses.

CPAP

- **Flow Driver** - preferred method for small babies
  - double nasal prongs, wide bore tubing and flow driver
  - adjust flow to achieve target pressure range (5-6cm)
- **Bubble CPAP** - preferred method for babies ≥32 weeks
  - bi-nasal with midline prongs and a F&P bubble device.
  - select the oxygen concentration required and set the flow meter at 6-8 L/min
- **Single Prong** - rarely used
  - single nasal prong attached to a ventilator (shortened ETT inserted approximately 2 cm, or about 1/2 the nostril to ear distance).
  - commence with 5 cm water and gas flow of 4-6 L/min
- To assess the adequacy of CPAP, watch for / assess:
  - Bubbly secretions at the mouth.
  - Transmitted hum of gas flow in the chest.
  - Clinical response including improved gas exchange and decrease in respiratory effort.
- If the baby is not responding, consider reasons for failure of CPAP:
  - Inadequate respiratory effort, e.g. secondary to maternal sedation or apnoea of prematurity (consider caffeine).
  - Respiratory failure that is severe or worsening.
  - Inability to achieve an adequate seal (e.g. due to crying or mouth opening).
  - Excessive abdominal distension (this can be because the compliance of the respiratory system has deteriorated so that it is worse than that of the abdomen!).
  - Development of a pneumothorax or other air leak.
  - Unsuitable disease process – e.g. anomaly such as tracheo-oesophageal fistula or diaphragmatic hernia.
Stopping CPAP

- There are many ways to wean CPAP but from May 2014 we have based our CPAP Weaning Guideline (Ref.2400577) on a multicentre randomised trial published in Archives of Dis in Childhood Fetal and Neonatal Edition July 2012
- Babies ≥ 34 weeks gestation usually come off CPAP on the first trial and clinical judgement should be used to decide when they are ready to come off CPAP and the guideline does not need to be used
- The guidelines are to be used for babies <34 weeks gestation
- The guidelines will suit the majority of babies but not all and babies with severe chronic lung disease may have an individualised plan
- Clinical judgement should always be used and CPAP restarted if a baby is clinically unwell but does not fully meet the criteria to restart
- If unable to remain off CPAP after 2 attempts and is ≥32 weeks corrected then consider High Flow
- High Flow may also be considered to be a better option in babies born at <28 weeks from 32 weeks corrected if they will need long term respiratory support.
- Consider increasing caffeine prior to weaning if events have been a concern

Stable Criteria

All criteria need to be present for >24 hours to be stable to trial off CPAP:

1. Corrected GA >28 weeks
2. PEEP 4-6 cmH20
3. Oxygen ≤ 25%
4. No significant recession
5. Tolerating time off with cares (ie: for 15min)
6. Minimal self correcting events with no clusters (≥ 3 events an hour)
7. No intercurrent illness being treated, immunisations or eye check due
8. Sats are <90% less than 10% of the time
9. Respiratory rate <60/min

Review criteria daily after stopping CPAP to ensure stability and at any time there is a clinical change.

Remain in Level 3 for 48 hours after stopping CPAP (if able) as 80% of babies who become unstable after trialling off CPAP do so within 48 hours (CWH NICU Audit 2017)

Unstable Criteria

If ≥ 2 criteria are present then restart CPAP:

1. Increased work of breathing and respiratory rate >75/min
2. Oxygen >25% on nasal prong oxygen (see algorithm below to calculate)
3. 3 or more apnoea/desat/brady in an hour
4. pH < 7.2
5. PaCO2 > 65mmHg or a rise in PaCO2 >10mmHg

If only 1 criteria present restart CPAP:

6. Increase work of breathing and respiratory rate >90/min
7. Major apnoea/brady needing IPPV

Once back on CPAP to remain on CPAP for 48 hours before reassessing if they are stable enough to trial off again. If events are the main cause of instability then review the caffeine dose.
Low Flow Oxygen Calculations

<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>$\text{FiO}_2 = 25%$ On the following Low Flow Rates</th>
<th>$\text{FiO}_2 = 30%$ On the following Low Flow Rates</th>
</tr>
</thead>
<tbody>
<tr>
<td>1kg</td>
<td>0.02 L/min</td>
<td>0.04 L/min</td>
</tr>
<tr>
<td>1.5kg</td>
<td>0.03 L/min</td>
<td>0.06 L/min</td>
</tr>
<tr>
<td>2kg</td>
<td>0.04 L/min</td>
<td>0.07 L/min</td>
</tr>
<tr>
<td>2.5kg</td>
<td>0.05 L/min</td>
<td>0.08 L/min</td>
</tr>
<tr>
<td>3kg</td>
<td>0.06 L/min</td>
<td>0.1 L/min</td>
</tr>
<tr>
<td>3.5kg</td>
<td>0.08 L/min</td>
<td>0.12 L/min</td>
</tr>
<tr>
<td>4kg</td>
<td>0.1 L/min</td>
<td>0.15 L/min</td>
</tr>
<tr>
<td>4.5kg</td>
<td>0.1 L/min</td>
<td>0.18 L/min</td>
</tr>
<tr>
<td>5kg</td>
<td>0.1 L/min</td>
<td>0.2 L/min</td>
</tr>
</tbody>
</table>

Adapted from ADHB Guidelines – Oxygen: Low Flow Calculator

Heated Humidified High Flow Nasal Cannula Oxygen

HHFNC is defined as oxygen through nasal cannula that is heated and humidified at flows above 1L/minute. It is increasingly being used as another form of respiratory support in addition to CPAP.

Indication
• Infants with corrected gestational age ≥ 32, after 2 attempts to wean off CPAP, or, as an alternative to CPAP in babies that they are likely to need ongoing respiratory support for weeks (usually <28wks at birth)
• Directly after extubation if clinically appropriate
• As an alternative to CPAP if still requiring respiratory support and FiO2 requirement <40%
• It is not for use currently for infants as the initial respiratory support irrespective of gestational age

Potential Benefits
• Provides a warmed and humidified flow of air and/or oxygen mixture (via a blender) to the infant where FiO2 can be monitored
• There is some degree of end distending pressure, however, debate remains as to how much
• May be better tolerated by some infants becoming unsettled on CPAP
• Reduced gastric distension than on CPAP
• Less pneumothorax
• Less nasal trauma
• Sucking feeds are more easily attempted

Equipment
• Fisher and Paykel Optiflow nasal cannula connected to our humidification circuits
• Size to be determined by nares - aim for most of nare to be filled without pressure
• Nare MUST NOT be tightly filled as the pressure generated will be excessive
• Mouth should be closed for optimal effect but active closure with a chin strap is not required

Management
• Start at 6L/min (maximum rate is 8L/min)
  • Pressure delivered will increase as flow increases and with decreasing infant weight
  • High Flow can be stopped from 2-4L/min flow
  • Flows <1L/min will have fluid “rainout” so should not be used
• Maximum flow rates vary: Small prongs – max rate 9L/min
  Medium prongs – max rate 10L/min
  Large prongs – max rate 23L/min
• Flow must be connected to heated humidification system.
• Infants should be placed back on CPAP if at any stage if:
  − the FiO2 increases by more than 10%
  − there are frequent apnoeas
  − there is substantial increase in work of breathing

**Complications**
• Potential for asynchrony in breathing which may result in the infants becoming more tired over long periods; therefore, a good assessment of work of breathing is required
• Potential for nasal erosion remains although less than with nasal CPAP
• There is concern about the unknown distending pressure and varied results gained in research studies, therefore ensure that prongs do not seal the nares and reduce flow as able.
• Potential problems with “rainout” resulting in lavage and apnoea; therefore, nurses need to be aware of clearing “rainout” regularly and ensuring that only heated tubing is used

**Weaning Humidified High Flow**
• The guideline for [weaning humidified high flow](Ref.2400578) has been based on the CPAP weaning guidelines, however, there are no published data on the best weaning methods
• As for the CPAP weaning guidelines they will fit most babies but not all and babies with severe chronic lung disease may have an individualise plan fine tuned with the information from sleep studies
• Some babies may need higher caffeine doses or treatment for reflux before weaning is successful
• Weaning of flow to commence under direction from Consultant
  − Wean oxygen first then wean flow
  − Generally flow will reviewed 12-24 hourly and reduced by 1L/min as tolerated until 2-4L/min and then place on low flow oxygen if required

**Stable Criteria**
All criteria need to be fulfilled for > 24 hours to be stable to wean High Flow by 1 L/min:

1. Oxygen ≤ 30%
2. No significant recession
3. Events are self correcting or are minor and no clusters (≥3 events an hour)
4. No intercurrent illness being treated, immunisations or eye check due
5. Sats are < 90% less than 10% of the time
6. Respiratory rate < 80/min

**Unstable Criteria**
If any criteria are fulfilled then increase the flow rate by 1L/min and consider CPAP if deteriorating:
Review criteria 12-24 hourly to screen for ongoing instability

1. Increased work of breathing and average respiratory rate >90/min
2. Rising oxygen requirement ≥5%
3. 3 or more events of any type in an hour
4. pH < 7.2
5. PaCO₂ rise of >10mmHg

**Indications for Intubation and Ventilation**
• Intubation at delivery – see Preparation for Resus (above)
• Intubation later on – similar indications as well as respiratory failure evidenced by deteriorating blood gases
• Intubation should be performed with morphine (occasionally fentanyl), atropine and suxamethonium prior (except if at delivery or as an emergency intubation)
• Atropine should not be used in babies prone to supraventricular tachycardia
• Suxamethonium should not be used in babies with major airway anomalies (in case you can’t intubate) or a family history of acetyl cholinesterase deficiency or malignant hyperthermia.
Videolaryngoscope – CMAC

**Indications**
- Elective intubations on NICU – note that the technique for intubation with the videolaryngoscope is different from without and both techniques need to be learnt as a videolaryngoscope is not universally available (hence why we are not suggesting its use in delivery suite for teaching)
- Baby >1000g (due to the size of the laryngoscope blade)
- For MIST therapy

**Supervision**
- Under supervision by SMO, Fellow or CNS/NP with credentialing.

**CMAC Guide**
- The videolaryngoscope is stored in pod 1 of room one (surgical space).
- An easy access guide with photos is available and attached to the CMAC trolley
- Available blades (2 of each):  
  - Size 0  Weight 1000-2500g
  - Size 1  Weight > 2500g

1. Plug monitor power cord in and turn on at the wall
2. Plug blade (laryngoscope handle) on to grey cord which is already attached to the monitor
3. Turn on monitor (top right button)
4. Screen will load (takes 15 sec)
5. Now ready to go – should not need to reset white balance each time but if colour is “off” can be changed under settings (wrench button – bottom right of the screen)
6. Taking photos/videos – either by pushing button on the base of blade or on the monitor.
7. After use – remove the blade from grey cord (cord stays attached to monitor)
8. Insert plug to the base of the blade for protection and put on the tray already with the equipment and take the blade in the tray to the sluice room to be sent to TSSU for cleaning by the ward aides.

**Supraglottic Airways in NICU**

- Supraglottic airways (SGAs) are a group of airway devices that can be inserted into the pharynx to allow ventilation, oxygenation, and administration of anaesthetic gases, without the need for endotracheal intubation.
- For anaesthesia, these devices are used for primary airway management, for rescue ventilation when facemask ventilation is difficult, and by an anaesthetist as a conduit for endotracheal intubation.
- We anticipate that in our *neonatal setting* they may be used when endotracheal tube insertion has not been possible but an airway is required.

**Device**  
iGel supraglottic airway
- Size 1, for patients 2-5 kg

**Storage location**  
Stored in Theatre 26 & 27 and in clean store room in labour ward.
In NICU – in resus trolleys room 1

**Placement**  
- i-Gel should be lubricated then inserted with supraglottic airway opening facing tongue surface.
- Gently advance to behind the tongue and then check chest wall movement
- If no chest wall movement consider adjusting to optimise ventilation and chest movement (moving up or down)
Protocol
- Resus should follow basic and advanced NLS algorithms
- Airway support is initially by bag/mask ventilation or T-piece (Neopuff)
- If further airways support is needed
  - 2 attempts may be made to intubate
  - If attempts unsuccessful (and baby still needs intubation) move to supraglottic airway
  - If unsuccessful then move back to Neopuff
- Remember that the LMA position may move during use – optimally the mask should cover from the epiglottis to below cords as in the above photo. If chest wall movement is not seen small adjustment of the LMA (in or out) will be required.
- Remember that if further airway support is required to call the neonatal SMO. There is always anaesthetic support available within the hospital if required (in main theatre and in obstetrics).

SLE Ventilator
- The SLE ventilator can provide pressure limited / time cycled ventilation, volume targeted ventilation and high frequency ventilation
- Ventilation has become much more complicated with the development of more versatile ventilators The basic principles remain the same, but if in doubt, discuss all issues of ventilator management with a consultant or more senior registrar until you have a good grasp of all the ventilator modalities we use.
- The SLE can provide 5 main forms of ventilation and a good review of the SLE ventilator can be found in the NICU G:drive under Ventilators and is a good presentation to understand how the SLE ventilator works

Modes of Ventilation
CMV – Continuous Mandatory Ventilation
- All breaths are initiated by the ventilator at the rate that is set
- The ventilator has no capability to synchronise with the baby’s breaths
- This mode can be used in babies with no respiratory effort ie: heavily sedated or muscle relaxed
- This mode should be changed when the baby starts to have respiratory effort as it is much better for the baby to have synchronised ventilation

SIMV – Synchronised Intermittent Mandatory Ventilation
- The ventilator breaths are synchronised with the baby’s breathing efforts
- If the baby does not breathe the ventilator will still produce breaths at the rate that has been pre-set
- The trigger needs to be set (usually at 0.2)
- The trigger senses when there is airflow from the baby. When it reaches the threshold, a breath is triggered
- If the baby breathes at a rate above the set rate then these breaths do not received pressure support
- When weaning from this mode the PIP is brought down first and then the rate is weaned to about 20-30/min before extubation

PSV – Pressure Support Ventilation
- In this mode the ventilator breaths are synchronised with the baby’s breathing efforts
- In contrast to SIMV, if the baby breaths above the dialled rate, these breaths do receive pressure support
- If set up as PSV only then the amount of pressure support is 100% for all breaths
- If set up as SIMV + PSV ON then the set rate of breaths receive 100% support but any breaths above this can have the amount of support manipulated from 5 -100%
- When the set % is reached the pressure support will terminate, this is called the termination sensitivity
- It is good to think of the breathing rate as a back-up rate rather than a set rate and it is there in case the baby’s breathing efforts are intermittent
- This can be the optimal mode of ventilation in a selected group of infants (e.g. those with musculo-skeletal problems affecting the chest)
- This mode is good to use if the baby needs more ventilatory support than SIMV can provide
- Weaning from this mode is accomplished by decreasing the PIP. Weaning the rate does not decrease the amount of ventilation provided as all breaths are augmented.
- Weaning can also be accomplished by weaning the amount of pressure support which in effect will wean them to SIMV once pressure support is removed
PTV – Patient Triggered Ventilation
- This mode is also synchronised
- The trigger is set and the baby triggers ventilation breaths
- The rate that is set is a back up rate only as all breaths are supported
- The difference from PTV and PSV is that the baby’s breaths are all supported 100% to the set PIP, whereas in PSV the amount of pressure support can be altered
- Weaning from this mode is achieved by weaning the PIP, weaning the rate will have no effect on the amount of ventilatory support provided

TTV – Targeted Tidal Volume
- Volume ventilation can be used in conjunction with all of the above methods of ventilating
- This mode can be very useful, especially in babies with rapidly changing pulmonary mechanics as it is continually adjusting for the infants respiratory effort and lung compliance
- It cannot be used if there is a large leak around the ET tube of >25%
- It changes from the conventional method of setting pressures to be delivered to setting tidal volumes to be delivered with the PIP and PEEP being set as alarm limits only
- Tidal volumes should range from 4-6ml/kg. The ventilator automatically adjusts the PIP to achieve this
- The inspiratory time will change with each breath and consider it as a back up setting only
- The ventilator assesses the expired tidal volume of each breath and adjusts the pressure for the subsequent breath to achieve a pre-determined target tidal volume.
- If the infant takes a large spontaneous breath that exceeds the target volume then the expiratory valve will open and no PIP is given.
- SIMV + TTV is useful in those infants with little lung disease that require minimal ventilation and as the last step in weaning to extubation.
- PSV + TTV should be used in infants with good respiratory drive and with little to moderate lung disease that is improving. It is a useful mode in infants with chronic lung disease
- PTV + TTV can be used in any infant with moderate to severe lung disease provided there is an adequate back up rate to provide adequate ventilation if the infant has poor respiratory drive.

Ventilator Settings
- Generally start with CMV or SIMV using pressure limited, time cycled, continuous flow mode
- For consistency, the ventilators are usually set up with settings of:

<table>
<thead>
<tr>
<th>Setting</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>PIP</td>
<td>18-20 cm</td>
</tr>
<tr>
<td>PEEP</td>
<td>5 cm</td>
</tr>
<tr>
<td>Gas Flow</td>
<td>8L/min</td>
</tr>
<tr>
<td>Rate</td>
<td>60 breaths/min</td>
</tr>
<tr>
<td>Inspiratory Time</td>
<td>0.34 sec</td>
</tr>
<tr>
<td>Oxygen</td>
<td>100%</td>
</tr>
<tr>
<td>Trigger</td>
<td>0.2</td>
</tr>
</tbody>
</table>

There are no magic set of ventilator settings that will be appropriate for all babies, and we are all responsible for carefully assessing each baby to arrive at the best individualised ventilation strategy.

In particular, keep in mind that:
- Babies who do not have very severe respiratory distress may be markedly hypocarbic on the above settings.
- There are many papers in the neonatal literature that suggest that hypocarbia/respiratory alkalosis is associated with worse pulmonary and cerebral outcomes than mild respiratory acidosis, probably due to excessive trauma to the lungs and impairment of cerebral blood flow.
- When a baby is connected to a ventilator you must constantly monitor the baby’s response including; tidal volumes, chest movement, saturation, blood gases
- After surfactant the lung compliance can change rapidly and you need to be vigilant of this and monitor the tidal volumes and wean ventilation appropriately and do blood gases early to prevent over ventilation
- Babies with severe lung disease/atelectasis and large babies will commonly require a significantly higher inspiratory pressure.
- Larger babies may need a longer inspiratory time
- In small sick babies early in the course of RDS, high rates can be useful to achieve synchrony.
Blood Gases

Acceptable blood gas ranges for most ventilated babies are:

- **pH**: 7.25-7.35
- **PaCO₂**: 40-50 mmHg for permissive hypercarbia in babies with severe lung disease as long as the pH is maintained > 7.25
  50-70 mmHg
- **PaO₂ (arterial)**: 45-70 mmHg < 1000 grams
  50-80 mmHg > 1000 grams
  see below for target oxygen saturations which are the predominant monitoring modality
- **HCO₃**: 20-30 mmol/L levels will rise overtime in babies with chronic lung disease to balance the acidity of higher CO₂ levels. Lower levels may be due to excess loss from premature kidneys or severe acidosis from hypoxia or sepsis, or a metabolic condition
- **Lactate**: < 3 mmol/L persisting higher levels need investigation for the cause such as sepsis, poor cardiac output or perfusion, metabolic conditions

Principles of Ventilation

Oxygenation

This is most affected by the mean airway pressure (MAP) and FiO₂

- **MAP** = \( K \times (\text{PIP} - \text{PEEP}) \times \frac{\text{RR} \times \text{Ti}}{60} + \text{PEEP} \)

- From the equation you can see that adjusting the PIP, PEEP and Ti can alter the MAP
- Increasing the PIP in steps of 1-2 will improve oxygenation and recruit alveoli but if the PIP is too high this will come with the additional problems of causing volutrauma to the lungs
- As you reach total lung capacity any increase in PIP will not provide an increase in lung volume and so is “unnecessary” pressure that could cause damage
- In some cases a slightly higher FiO₂ may need to be accepted to keep the PIP down
- Increasing the PEEP (usually from 5 to 6 or 7) can be helpful in a baby who is hard to oxygenate
- Increasing the FiO₂ is the easiest way to improve oxygenation but the side effects of oxygen in preterm babies must not be ignored
- Remember that there may be extrapulmonary shunting that us affecting oxygenation eg: PDA, CHD
- The inspiratory time is set at 0.34 sec but may need to be altered
- The Ti is chosen when considering the baby’s respiratory time constant which is the product of pulmonary resistance and compliance
- The Ti will be short (0.3 sec) in babies with normal resistance and low compliance (eg: early in RDS)
- The Ti will be longer (0.34-0.4 sec) in babies where the compliance improves (eg: after surfactant)
- The Ti may be much longer (0.5sec) in babies with high airways resistance (eg: MAS, BPD)

Ventilation

- The elimination of CO₂ is proportional to the minute ventilation (200-300ml/kg/min)
- Minute ventilation is improved by decreasing dead space, increasing rate, increasing PIP, or adjusting PEEP (PEEP may need to go up or down depending on the effect on alveolar ventilation).
- Increasing the rate up or down to increase the CO₂ is the most often used way to manage the CO₂
- Changes in the rate are usually changed by steps of 5-10 breaths/minute depending on the blood gases
- Increasing the PIP will also help clear CO₂

Weaning

- There is no magic number to reach before extubating a baby and it will depend on their clinical condition
- A rough guide would be to wean a preterm baby to a PIP of around 14-16 and a term baby to around 16-18 and a rate of 20-30/min before extubating
- Infants <30 weeks should be loaded with caffeine before extubation
- In SIMV weaning is achieved by minimising the PIP and then dropping the rate
- In PTV and PSV weaning is achieved by dropping the PIP
- In TTV weaning is achieved by dropping the tidal volume

### High Frequency Ventilation

- This has become a very important method of ventilation for babies with severe respiratory failure
- Ventilation occurs with a high continuous distending pressure, supraphysiologic respiratory rates, small tidal volumes and achieves gas exchange by oscillatory flow
- Discuss with a consultant as it is not often used and is different from conventional ventilation
- There are a number of different ventilators that can provide HFOV but the ventilators we use at CWH are the SLE or the Sensomedics (only delivers HFOV)
- HFOV tends to be used as a rescue treatment for babies who have failed on conventional ventilation
- However, there is no conclusive evidence that HFOV versus conventional ventilation improves outcomes whether it is used electively or as a rescue method of ventilation

### Open Lung Volume Concept

- Using an open lung or high volume strategy irrespective of the method of ventilation will be lung protective and better outcomes will be achieved
- Ventilation in this manner improves gas exchange, reduces ventilation induced lung injury, preserves surfactant function and improves lung mechanics
- Open lung volumes can be achieved with recruitment methods in babies with certain respiratory diseases

### Principles of HFOV

- Mean Airway Pressure (MAP) – sustained inflation and recruitment of lung volume by applying a continuous distending pressure. Usually start 2-4cmH2O above the conventional MAP. Found as Mean on the SLE screen.
- Amplitude – the size of the chest excursion or “wobble” from the baseline distending lung volume. The amplitude needs to be adjusted so that the chest is visibly shaking. Starting points are 1.5x the PIP on conventional ventilation or 2x the MAP. Found as delta P on the SLE screen
- Frequency – the speed in Hz of the “wobbles” of the chest, usually between 6-12 Hz. Found on the SLE screen as HFOV rate

#### Ventilation (ie: C02)

- This is manipulated by changes in the amplitude and frequency mainly
- It is best to set the frequency for the disease process and leave it alone and then make changes with the amplitude first
- Increasing the amplitude will cause a fall in C02 and decreasing it causes a rise in C02
- Make adjustments in increments of 1-2 units
- Aim for a tidal volume of 2ml/kg
- Decreasing the frequency will cause a fall in C02 (opposite to what occurs with conventional ventilation)

#### Oxygenation (ie: O2)

- This is manipulated with the FiO2 and MAP
- Increase the MAP in steps of 1-2 to improve oxygenation and decrease in the same way when weaning
- If not improving, consider the presence of a pneumothorax or overinflation and check with a CXR
- Increasing the MAP will also cause a fall in the CO2

![Lung Hysteresis Curve](image)

- **% TLC**
- **Pressure mmHg**
- **Underinflated**, collapsed lung with low tidal volumes and lung compliance and a high oxygen requirement with poor CO₂ clearance
- **Improved inflation** and subsequently improved tidal volumes, compliance, oxygenation and ventilation
- **Overdistended** lungs with deteriorating tidal volumes, lung compliance and a rise in oxygen requirements. Depending on the severity of the overinflation the systemic circulation and blood pressure can be compromised
- **Optimal inflation** with lungs that have passed through total lung capacity and recruited alveoli which remain open when the MAP is reduced ie: maintaining the same lung volume at a lower pressure. Optimal lung compliance, oxygenation and ventilation are seen at this point in the hysteresis curve

**Recruitment Manouevres**
- The lung volume is increased through total lung capacity (ie: from point 1 to 4 in the above graph)
- Not all disease processes are compatible with recruitment methods and sometimes are contraindicated
- These manoeuvres should only be done by experienced staff who understand the principles behind them

**Diseases to recruit**
- HMD
- Pneumothorax (drained)
- Pneumonia
- PPHN with parenchymal lung disease

**Diseases to avoid recruitment**
- PPHN with normal lungs
- Pulmonary hypoplasia
- PIE
- Congenital diaphragmatic hernia

**Incremental Inflation (see graph below)**
- Best method of recruitment with improved lung volumes, better oxygenation, no increased risk of overdistension or air leak and more lung units are recruited compared to other methods
- Start at the MAP already in use or start at 8-12cmH₂O
- Hold O₂ and frequency the same
- Increase MAP by 2cmH₂O increments to about 20-25cmH₂O
- Endpoints to indicate that you are reaching TLC are that the sats stop improving or actually fall (if you have gone too far), the FiO₂ drops and the tidal volumes improve then decline as you reach TLC

**Incremental Deflation**
- Decrease the MAP by increments of 2 until just above the starting MAP until there are signs of de-recruitment - rising FiO₂ and falling sats, rising CO₂ and falling tidal volumes. This is the closing pressure.

**Re-Recruitment**
- Increase the MAP to the peak pressure previously achieved for 2-5 minutes
- Re-recruit lung then drop the MAP back to 2cmH₂O above the closing pressure
- Now the lung will maintain good inflation but at a lower mean airway pressure
Reassessment
- Need to repeatedly reassess MAP and lung volumes as the disease processes are dynamic
- May need to recruit 2-3 times a day
- After suctioning with loss of lung volume a shorter version of recruitment would need to be used eg: single dynamic inflation of increasing the MAP by 2-4 for a few minutes to re-recruit alveoli

Supportive care
- Correct tube size and well strapped
- Minimal leak around the ET tube
- Sedation ± muscle relaxation
- Transcutaneous monitoring
- Correct volume depletion if present
- Correct acidosis
- Correct cardiac function and hypotension
- If PPHN is present – normalise Hb, Ca, Mg levels and manage with NO

Suggested Settings for Disease Processes
- Preterm HMD frequency 12Hz (range 12-15) The lower the frequency the lower the shear forces and potential for lung injury
- Term lung disease frequency 10Hz (range 8-12)
- Pulmonary Hypoplasia frequency 10Hz (range 10-12)
- Meconium Aspiration frequency 8Hz (range 6-10)
- Established PIE or CLD frequency 6Hz (range 6-8).
- Pulmonary hypoplasia
  - Structurally immature lungs and a low pressure strategy is best
  - MAP around 12cmH2O and Frequency 10-12Hz
  - Hold the same lung volume and wait for time dependent recruitment
  - Avoid aggressive recruitment
  - CXR is often unhelpful to assess lung volume
- Congenital Diaphragmatic Hernia
  - Hypoplastic lung with abnormal pulmonary vascular tree
  - Use a low pressure strategy without lung recruitment
- Pulmonary Intersitital Emphysema
  - Low pressure strategy aiming for the lowest MAP possible to maintain oxygenation
  - MAP around 10-14cmH2O and frequency 6Hz
  - No recruitment manoeuvres, but, accept high FiO2
  - Position good lung up

Monitoring on HFOV
- The blood pressure and a chest radiograph must be checked soon after starting HFOV to assess the effect of the MAP on cardiovascular function.
- Check lung volumes (aiming for 9-10 posterior ribs) and cardiac contour (a narrow cardiac contour may imply too high a mean airway pressure or need for blood volume expansion).
- Reduction of the MAP and/or administration of fluid to expand intravascular volume may be necessary.
- Repeat CXR’s will be needed 8-12 hourly to assess lung volume
- In cases with pulmonary hypoplasia the CXR is an inaccurate method of assessing lung volume

Pneumothorax

Clinical Signs
- Respiratory distress
- Increase in oxygen requirement – can be rapid or insidious
- Reduced air entry on one side
- Asymmetric chest movement
- Shocked baby if the pneumothorax is under tension
Causes
- Spontaneous – at delivery
- Iatrogenic – bag-mask resuscitation or artificial ventilation
- Secondary to lung pathology – Respiratory distress syndrome
  Meconium aspiration syndrome
  Pulmonary hypoplasia

Transillumination
- If a pneumothorax is suspected clinically, use the cold light to transilluminate the chest
- Place the light in the axilla on both sides and below the xiphisternum to rule out pneumopericardium
- If the hemithorax transilluminates unequivocally, then a CXR is not required before definitive management.
  False positives occur if there is tissue oedema, lung emphysema or an enlarged stomach
  False negatives occur in bigger babies when the light cannot penetrate as well

Management (see procedures section)
- Confirm the presence of a pneumothorax by transillumination or CXR prior to a formal chest drain
- CXR’s can be hard to interpret at times as in babies the air leak rises anteriorly under the sternum so a lateral XRay as well as an AP are needed for diagnosis
- Needle aspiration may need to be done urgently without transilluminating or getting a CXR first
- Not every pneumothorax needs to be drained
- Small to moderate pneumothoraces, especially in larger babies who are not on assisted ventilation will often resolve without treatment.
- Even in smaller babies and those on ventilators, one-time needle aspiration will sometimes suffice, especially if the baby can be maintained on relatively low mean airway pressure.
- If there is underlying disease such as meconium aspiration or surfactant deficiency then ventilation, surfactant administration (if indicated) and chest drain insertion is usually needed.
- If the baby needs to be transported the air leak needs to be drained before travelling at altitude as undrained air expands at altitude
- Always re-Xray after aspiration or inserting a chest drain

Sedation for Ventilated Babies
- Many babies who commence mechanical ventilation will not require continuous sedation especially if they look like they will wean quickly and can be managed with CPAP. For those who do (all babies on high frequency and those needing high pressures), we generally begin with 10 mcg/kg/hour morphine. A loading dose of 50 to 100 mcg/kg given over 30 minutes should be considered (discuss with the consultant) to ensure adequate sedation is achieved in a short time frame, but caution is required if the baby is hypotensive. Infrequently, a baby will need a muscle relaxant (always discuss with consultant).
- Use of sedatives should be kept as low as possible by the optimal use of synchronised modes of ventilation, high frequency ventilation and volume ventilation in appropriate infants. Generally, sedatives should not be used to control the agitation of air hunger and hypercarbia due to less than optimal control of the airway or use of respiratory support.
- Careful positioning of the baby, attention to reduction of noise and light levels, and minimal handling
- However, when gas exchange cannot be improved by other means, or the baby seems uncomfortable or in pain (hypertensive and tachycardic) sedation and analgesia are appropriate to relieve suffering and reduce tissue O2 demands and CO2 production to a level that can be matched by cardiorespiratory function.
Artificial Surfactant

- Curosurf is a pig surfactant, administered via ETT at dose of 1.25ml/kg.
- Surfactant should be drawn up using a sterile syringe and needle, using the technique described on the package insert. Always discuss plans to use surfactant with a consultant. A usual course is two doses 12 hours apart, although the second dose is not always necessary and occasionally a third dose is indicated. If the baby is < 1 kg, the rest of the vial should be labelled with the baby's name, time and date and returned to the refrigerator. It can then be used for the second treatment. Alternatively, twins, or other babies born in rapid succession can be treated from the same vial if their weights are appropriate and careful sterile technique is used. Opened vials older than 24 hours should be discarded.
- We use surfactant in the delivery room for some extremely low birth weight babies (< 1kg, < 28 weeks). For most of these births, there will be a consultant present who can help decide whether to give the surfactant. When given immediately after delivery, the surfactant can usually be given safely and effectively as a single bolus, although most give it in 2 aliquots.
- For most bigger babies, we tend to treat with surfactant in the NICU, as soon as it is obvious that the baby has Respiratory Distress Syndrome that will require intubation and mechanical ventilation. When given after admission to the NICU, it is usual to split the administration into two doses. We do not routinely reposition the baby between fractions of the dose and the aliquots can be given in quick succession (total dose over a minute or two) if the baby is tolerating the process well and saturations have returned to baseline between aliquots.
- Watch for spillage into the ventilator circuit or around the ET tube in the trachea. Examine the chest movement carefully and check tidal volumes and blood gases soon after administration as pulmonary mechanics can improve within a few breaths, necessitating adjustment of ventilator settings.
- On auscultation, the baby's chest may resemble a washing machine for up to hours after the dose. This is not an indication to suction the airway, as you may merely remove the administered surfactant. Occasionally artificial surfactant treatment will mobilise mucous plugs and cause them to block the airway. Suspected airway blockage is a good reason to override the protocol of not suctioning the baby for at least 6 hours after surfactant administration.
- The 2nd dose is determined by the XRay features of HMD and the degree of ventilation support needed. An audit of our use of surfactant from 2002-2004 showed that babies who had received complete antenatal steroids who received their first dose < 90 minutes of age ie: prophylactically in birthing suite were less likely to need a second dose. Many babies are extubated between 6-12 hours of age on to CPAP. If this is the case a second dose of surfactant is not usually required.

Minimally Invasive Surfactant Therapy (MIST)

- Infants in the delivery suite who have respiratory distress requiring surfactant should be intubated and have surfactant administered via the endotracheal tube.
- Infants in NICU who fit the following criteria may receive surfactant without sedation whilst spontaneously breathing and then continue on CPAP/BiPaP

Indication for MIST

- To be performed in NICU only within first 24 hours of life (preferably <6hrs age)
- ≥ 28 weeks gestation
- History and/or CXR consistent with a diagnosis of Respiratory Distress Syndrome
- Oxygen requirement >30%
- Regular respiratory effort without apnoea
- Surfactant is felt to be needed but not an ongoing period of ventilation

Process

- Refer to video of the procedure – Neonatal Intranet/Other Guidelines/MISTVideo
- Administer sucrose prior to the procedure
- Atropine prior to the procedure may also be considered
- Draw up the surfactant with and extra 0.5mL of air. Attach to an iv connector. The dead space for the connector and catheter is 0.2mL
- Baby should remain on CPAP throughout the procedure if at all possible. If the interface on the nose is in the way of the laryngoscope push it out of the way whilst visualising the cords then replace when administering surfactant
Remove the needle from the 13cm long 16G angiocath and mark circumferentially with the wax pen at 2cm from the tip to show how far in to insert the tube past the vocal cords – both are stored in the bottom drawer of the Respiratory Support trolley in Room 1.

May also mark 16G angiocath at the estimated length at the lips for the size of the baby

Fashion an anterior curvature of the catheter to aid intubation

If a less experienced practitioner is inserting the catheter then use the videolaryngoscope (when available) and/or attach a Pedi-cap to the blue adaptor from a 3.5 ETT (discard the tube) and when the catheter is placed throught the cords attach the ETT adaptor and Pedi-cap to the MIST catheter to detect CO₂. This is to ensure the tube is placed correctly prior to surfactant administration

Visualise the vocal cords with the laryngoscope and insert the angiocath to the marked distance. Magills can be used to help if preferred.

Remove the laryngoscope and hold the baby’s mouth shut with your hand to secure the angiocath position

Place Pedi-cap to the catheter as described above to check placement

Instill the surfactant as a continuous slow bolus over at least 60 seconds

Remove the angiocath once the surfactant has been given and remain on CPAP

Be prepared to give positive pressure ventilation if clinically indicated ie: apnoeic, chest not moving, bradycardia

There may be some refluxing of surfactant into the oropharynx and this is to be expected

There should be no more than 2 attempts by the Reg/CNS/NNP to intubate and 1 attempt by the SMO before the procedure is stopped in favour of premedication and formal intubation

**Oxygen Saturation Targets – NICU Inpatients**

**Note that the sats targets and limits below are for babies in the NICU.** Babies on the postnatal ward should continue to target sats of ≥ 95% in air with neonatal review if sats are <95%.

Saturation targets should be set for all babies and in particular babies that are ventilated and on oxygen and should be documented in the care plan and updated with any changes.

These limits were reached by consensus by the Newborn Clinical Network: Practice recommendation for oxygen saturation targets for newborns cared for in neonatal units, NZ (October 2015)

An individualised approach may be taken for babies if it is deemed more appropriate. For example: a term baby with cyanotic heart disease may have lower saturation targets than in the table below.

Variation to these targets may be decided by the SMO on service and should be documented and updated with any changes.

If a decision is made on ward round to keep at set level to keep saturations more stable this needs to be recorded on MDCP.

Oxygen sats will fluctuate but the aim is to keep the sats as stable as possible within the target range.

There is a trade off between too much oxygen delivery and retinopathy of prematurity and CLD and lower oxygen delivery with less retinopathy but increased disability and mortality.

<table>
<thead>
<tr>
<th>Corrected Gestation</th>
<th>Saturations Targets on Oxygen</th>
<th>Alarm Limits on Oxygen</th>
<th>Saturation Targets in Air</th>
<th>Alarm Limits in Air</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt;36 weeks</td>
<td>90-94%</td>
<td>89-95%</td>
<td>90-100%</td>
<td>89-100%</td>
</tr>
<tr>
<td>Start oxygen if sats are &lt;90% in room air</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>≥36 weeks</td>
<td>93-97% ≥95% if PPHN</td>
<td>92-98%</td>
<td>93-100%</td>
<td>92-100%</td>
</tr>
<tr>
<td>Start oxygen if sats are &lt;93% in room air</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Caffeine

Starting Caffeine

< 32 wks – start on admission for all babies
≥ 32 wks – start if apnoea/desaturation/bradycardia events occur, usually in the first week, frequent events predominantly occur < 34 weeks corrected gestation but can occur at any gestation.

**Maintenance dose:** 10mg/kg if <28 weeks
7.5mg/kg if ≥28 weeks

Dose once daily charted for 1000 in the morning
Dose twice daily charted for 1000 and 2200 if the dose is ≥15mg/kg/day and the baby continues to be unstable on once daily dosing

- **Periodic breathing** is a normal phenomenon in the preterm infant and can be seen clinically but also inferred if the histogram shows a RR <30/min. However, caffeine does not need to be started for this indication alone, there needs to be clinical events as well that are either significant or frequent.
- It is an appropriate option to simply monitor a baby with infrequent or minor events alone without starting caffeine

Stopping Caffeine

If < 28 weeks at birth – review at 36⁺⁰ weeks
If ≥ 28 weeks at birth – review at 35⁺⁰ weeks

- Longer duration of caffeine is often needed if there were problematic apnoeas and these babies have usually been on ≥ 12.5mg/kg/day caffeine +/- doxapram.
- The respiratory status assessment for when to stop caffeine includes the baby’s gestation, current clinical state, Level 2 chart of events and the O₂ saturations and respiratory rate histograms and these should not be looked at in isolation.
- If a baby is becoming ready for discharge but remains on caffeine consider going home on caffeine rather than stopping caffeine in hospital and delaying discharge for a further 5-7 days.

Criteria to Stop

- No apnoeas (1) or no other events (2,3) requiring stimulation (7) for 5 days
- No or infrequent self-correcting desaturation/bradycardia events (2,3,5) for 3 days
  - maximum of one event in any hour – ie: no suggestion of clustering.
  - < 3 events per day
  - observe the respiratory pattern on the monitor looking for periodic breathing
- Saturation histogram –< 5% time with saturations <90% (note: weaning from CPAP or Hiflow criteria <10% time with sats < 90%)
- Respiratory rate histogram - a rate < 30 is suggestive of more frequent short pauses in breathing and a surrogate measure for periodic breathing

Assessment after Stopping Caffeine

- A review of the 12 hour saturation histogram is recommended every 2-3 days and on day 7 after stopping caffeine – aiming for <5% time saturations are <90%. If in doubt a sleep study will be needed after 7 days to determine stopping monitoring.
- If desaturations are noted after stopping caffeine then assess the histogram and event record, Caffeine can be restarted without a sleep study.
- Ideally a baby should stay in NICU monitored for 7 days after stopping caffeine but if this is holding up discharge in a baby who is orally feeding then the baby could go home 5 days after stopping caffeine if there is a home monitor available to be used until 7 days after caffeine was stopped.
- If a baby **restarts caffeine** and is ready for discharge it is recommended they stay **at least** 2 days after restarting caffeine. If they have no events they can go home without monitoring but this should be individualised
- If there are clinical events after **restarting caffeine** the baby should stay at least 5 days and go home with monitoring.
Monitoring in the Unit

- Cardiorespiratory monitoring is required for all babies up to 35 weeks (irrespective of caffeine requirements) and must continue whilst on caffeine and for at least 7 days after stopping.
- Refer to OPS Monitoring-of-infants-in-NICU-PPN83 for normal parameters for HR, RR, Temp, BP and further details on monitoring and observations.

Considerations for Length of Monitoring in NICU

- Birth gestation
- How difficult was the apnoea of prematurity
  - caffeine needed beyond 38 weeks
  - was doxapram needed
  - was CPAP/High Flow needed beyond 36 weeks
- Chronic lung disease diagnosis – oxygen or respiratory support at 36 weeks. Especially if still needing more than 25% FiO2 on the shift test.
- Upper airway obstruction abnormalities eg: Pierre-Robin
  - These babies require cardiorespiratory monitoring until review for home.

Home Monitoring

- The decision for monitoring at home will be made by the baby’s Consultant and should be discussed during discharge planning. The need for home monitoring would usually be indicated from results of a sleep study.
- Babies born < 32 weeks with a reassuring Level 4 saturation study by 38 weeks with DSI 4s of 10% < 20 do not require monitoring after discharge.
- Graesby apnoea monitors have been replaced by Bitmos monitors that monitor HR and saturations.
- Parents/caregivers will need to be taught CPR and the patient issued with a Blue Card if going home on monitoring.

Bitmos Monitor

- < 32 weeks going home on caffeine, will continue caffeine to 44 weeks corrected GA, then if the Level 4 saturation study is normal will stop caffeine and the monitor will stop 1 week later if there have been no alarms. Outreach will review.
- Short term use in babies who have stopped caffeine less than 7 days before discharge or have restarted caffeine close to discharge and have ongoing events.
- Chronic lung disease (oxygen or respiratory support at 36 weeks GA) babies on home oxygen at discharge. Monitor to continue until off oxygen / clinic review.
- Individual cases in consultation with BD / NA

Home Cardiorespiratory Monitor

- Obstructive apnoeas eg: Pierre Robin, CHARGE syndrome, airway anomalies
- When the Massimo or polysomnograph indicates obstructive events
- Home oxygen in selected cases

Discharge on Caffeine / Oxygen when a monitor is not required

- Term 37+ week infant needing low flow oxygen for meconium aspiration / pneumonia. These babies will need sleep studies to wean from oxygen.
- 32 weeks or more on caffeine or oxygen for discharge, no stridor, events resolved clinically. They will stop caffeine if there have been no concerns or events 4 weeks after discharge. They will be seen by outreach who can do saturation checks at a visit but do not need further studies. This is based on an audit of cases in 2018. They do not need to be seen in clinic after stopping caffeine unless NORN request.
Follow up summary:

<table>
<thead>
<tr>
<th>Group</th>
<th>Caffeine at discharge</th>
<th>Monitor at DX</th>
<th>FU sleep study</th>
<th>When to stop caffeine</th>
<th>Clinic FU</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt; 32 weeks</td>
<td>Yes</td>
<td>Yes</td>
<td>Yes</td>
<td>After 44 weeks PCA, and after a home sleep study</td>
<td>Yes Primary consultant POPD</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Discretion can be used in babies 30-31 wks</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt; 32 weeks</td>
<td>No</td>
<td>No</td>
<td>No</td>
<td></td>
<td>Yes Primary consultant POPD</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Based on histogram</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>&gt; 32 weeks But stridor / event concerns</td>
<td>Yes</td>
<td>Yes</td>
<td>Yes 4 weeks post discharge prior to stopping caffeine</td>
<td>Decision made after reviewing home sleep study</td>
<td>Yes Primary consultant POPD</td>
</tr>
<tr>
<td>&gt; 32 weeks No stridor / events</td>
<td>Yes</td>
<td>No</td>
<td>No</td>
<td>4 weeks after discharge</td>
<td>If requested by NORN at CWH clinic</td>
</tr>
</tbody>
</table>

Level 4 Saturation Studies

- Neonatal saturation studies are pulse oximetry studies with a 2 second averaging time that are performed in the neonatal unit.
- They are not a full cardiorespiratory sleep study as they only measure heart rate and saturations via a saturation monitor. There may be rare occasions when a full polysomnography study is required (including respiratory monitoring, airflow monitoring). The full sleep studies are usually performed in Christchurch Hospital or Wellington and so are logistically difficult for a NICU baby. Consider a full study in high risk patients such as with upper airway obstruction or neuromuscular disorders.

Timing

- Studies are usually performed after 36 weeks corrected gestational age.
- Studies can be performed earlier when respiratory support and oxygen are ceased in babies < 32 weeks but with the extended use of cardiorespiratory monitoring and histogram checks this should not be necessary as babies < 32 weeks stay monitored until 36 weeks. Many infants will only require a single study and repeat studies should usually be after 2 weeks.
- Repeat studies are requested in infants who are having oxygen titrated or where there are clinical concerns.

Indications

- Babies <28 weeks at birth should have a Level 4 saturation study
  - Day 7-10 after caffeine is stopped
  - by 38 weeks if still on caffeine / respiratory support /oxygen to assist with timing of weaning
- If desaturations (needing stimulation, or frequent > 1/hour) occur after caffeine is stopped and recommencement of caffeine is being considered and there is time for a study before changing management
- Babies unable to be weaned off oxygen and preparing for discharge
- Babies with upper airway symptoms/congenital malformations/hypopotonia – stridor, retrognathia, cleft palate, Pierre Robin, CHARGE, laryngomalacia
- Babies with desaturations during feeding (possible aspiration events or severe GOR) not improving with time and > 39 weeks
- Did not pass a car seat trial
Equipment
- Masimo radical 7 saturation monitor (that has been set up for sleep study software)
- Level 4 saturation study recording sheet

Process
- The Discharge facilitators or ACNM can source the Masimo monitor
- Record for at least 6 hours, preferably 24 hours. Minimum standard is 12 hours
- Observation chart to be filled in hrly by staff of all handling events/ sleep position to help in data interpretation
- Observations include – sleep position, quiet or active sleep, feeding and by which route, any problems with the trace, reasons for poor pick up
- Once the study is completed the Massimo monitor should be removed and downloaded (on the computer in room 6)

Targets
- Targets will differ depending on the corrected gestational age of infant.
- At times infants will require different saturation targets for specific conditions e.g. may be lower for congenital heart disease or higher for infants who have pulmonary hypertension.

Studies performed between 32-35+6 weeks CGA
a. The agreed target saturations are Mean saturations > 93%, and 5-10% of the time with saturations < 90% (British Guidelines 2009, Australasian Guidelines).

Studies performed at 36 weeks or greater CGA
b. The agreed target saturations are Mean saturations > 93%, and < 5% of the time with saturations < 90% (British and Australasian Guidelines 2009, Starship clinical guidelines for domiciliary oxygen accessed 9/8/16).
   i. The Thoracic Society of Australia and New Zealand position statement on oxygen therapy in infants with chronic neonatal lung disease has been updated in 2018.
   ii. Prior to discharge on home oxygen (GA < 28 weeks) an echo should be performed to assess for pulmonary hypertension

- Low flow oxygen to be reduced in increments on oxygen regulators - 0.2 / 0.12 / 0.08/ 0.05 / 0.03 / 0.02 L/min
- Babies with chronic lung disease need to be ≤0.5L/min via nasal cannulae to be considered ready for discharge

Reporting
All inpatient Level 4 saturation studies are to be reported by Bronwyn Dixon (if unavailable then Nicola Austin will report and if she is unavailable then the SMO on service)
- A copy of the reported study should be put in the notes and imported onto Health Connect South into the clinical investigation section by the NICU secretary/ward clerk

The reporting format should include:
- Quality of recording
- Assessment of mean saturations and time <90% saturations (are targets met)
- Assessment for features such as evidence of reflux or feed incoordination, periodic breathing
- Desaturation index (4%) – located on page 2 of study.
- Any changes in treatment that are required – such as how long caffeine treatment is required for and what monitoring is required and the plan for when study needs to be repeated.

References
Car Seat Trials

There are no gold standard to refer to regarding assessing the safety of a preterm infant in a car seat, however, in this NICU we will continue to undertake car seat trials prior to discharge as routine practice.

The car seat trial needs to be done in conjunction with family education around safe positioning and use of the car seat.

Criteria
- <36 weeks gestation or < 2500g
- ≥ 36 weeks gestation if on home oxygen or caffeine, hypotonic, respiratory or airway abnormalities, complex or cyanotic heart disease, abnormal neurology

Process
1. Car seat to be brought in prior to rooming to ensure the car seat trial can be undertaken and the baby deemed safe prior to preparing for rooming in and discharge
2. Attach a Car Seat Trial Massimo monitor on the baby’s foot (averaged to 2 seconds)
3. Monitor supine in the cot for 30 minutes and preferably when in a deep sleep. If sats are below 93% commence a sleep study overnight.
4. If the supine cot trace is acceptable then place baby in the car seat and monitor for at least a further 30 min

Car Seat Positioning
- Proper positioning of the infant in the seat is important to minimise the risk of respiratory compromise
- Rear facing car seat – 3 or 5 point harness
- Semi-reclined position – 45 degrees
- Buttocks and back flat against the back of the seat
- Head is upright and airway not compromised – padding can be added for lateral support of the head and neck but any head support should not be pushing the head forward and therefore compromising the airway
- Harness straps at or below the shoulder level – generally at the lowest position
- Secure harness around baby - no more than 1 finger should fit between harness and collarbone
- Harness straps should be snug
- Harness retainer clip at level of armpits or midpoint of chest

Failure Criteria
- Bradycardia <80/min for >20 secs
- Oxygen saturations <90% for >60 secs or regular fleeting desats <90%
- Respiratory distress/obstructive episodes

If the baby fails the car seat trial place supine in the cot and commence a sleep study with the Massimo monitor currently on the baby and inform the ACNM, Registrar/CNS/NNP and SMO.

Management options
- If the baby stabilises in the cot then a sleep study overnight is all that is required initially
- Oxygen can provide immediate treatment for desaturations but may eradicate periodic breathing and if put on straight away may normalise the trace limiting the information available to assess
- Caffeine may be recommended after review of the sleep study

Car Seat Education
- Staff to show the family how to secure the baby safely in the car seat
- Rear facing and secured on the back seat (tether, strap and bolt if indicated)
- Recommend an adult sit beside the baby in the back seat if possible
- Advise not to leave the baby unsupervised in the car seat at any time
- Baby not to be left to sleep in the car seat – cot or bassinet is the safest place to sleep
- If travelling for >1 hour then stop the journey for at least 10 min every hour and get the baby out of the car seat to stretch and feed if required. This should be continued until the baby has good head control.
• Once you reach your destination take the baby out of the car seat
• Minimise the time the baby is in the car seat
• If the baby is on an apnoea monitor or cardiopulmonary monitor at home this should also be used when in the car seat
**FLUIDS**

### Total Fluids

<table>
<thead>
<tr>
<th>Day</th>
<th>Normal Birth Weight for Gestation and &gt;1500 g</th>
<th>IUGR or &lt;1500g</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>65</td>
<td>90 - Consider checking electrolytes 12 hourly and adjust</td>
</tr>
<tr>
<td>2-3</td>
<td>90</td>
<td>90 - 120</td>
</tr>
<tr>
<td>3-5</td>
<td>120</td>
<td>120 -150</td>
</tr>
<tr>
<td>5-7</td>
<td>150</td>
<td>150 - 180</td>
</tr>
<tr>
<td>&gt;7</td>
<td>150 - 180</td>
<td>150 - 180</td>
</tr>
</tbody>
</table>

- Just as with ventilator settings, there is no magic fluid prescription that will work for all babies.
- The baby's weight and serum sodium tend to be the best guides to hydration status in the first few days. A high serum sodium in the first few days is much more likely to be due to dehydration than to an elevated total body sodium (a rise in serum sodium of 10 mmol indicates 10% dehydration), and conversely, hyponatremia is commonly caused by overhydration. Although many babies in intensive care are too sick to weigh often, weight measurements can be very useful on occasion. In using weight as a guide, allow for a weight loss of 2-4% per day for the first few days. Measured fluid intake, urine output, blood pressure and clinical appearance are also important cues in estimating fluid requirements. Urinary electrolytes have a role in some babies but are useless in babies who are on dopamine or diuretics. Measurement of losses via other routes such as ostomies and gastric aspirates can be very useful in selected babies. Two caveats are that overhead warmers and phototherapy lights can increase insensible losses, whereas (due to enhanced diffusion of water vapour) high frequency ventilation can result in net free water uptake.
- We usually base decisions about fluid management on results from samples analysed in the main Canterbury Health Laboratories. The blood gas machine measures electrolytes can provide a useful guide to the electrolytes and to follow trends but the values are not always accurate. Keep in mind that the unit machine measures whole blood electrolytes, so the normal ranges are about 3-5% lower, (e.g. about 132 - 142 for Na). Also, suspected erroneous results are usually too low, rather than too high. **Thus, elevated results from the unit machine should be taken seriously.**
- In general, both premature and term babies are born with a reserve of extracellular fluid and sodium equivalent to 5-7% of birth weight (roughly 150-300 ml in a term baby) that they can lose after birth without a rise in aldosterone levels or any other indication of dehydration. Creatinine clearance is low immediately after birth and increases with advancing post-natal age. This is the reason why the prescribed fluid intake is initially low and increases.
- Typical insensible water loss at term is about 20-60 ml/kg/day and can be far greater in very premature babies, who have a much less adequate cutaneous barrier to water loss, and in infants under radiant warmers and/or phototherapy lights. Thus, all these infants need very careful attention to their fluid balance.
- We have a protocol for the use of petrolatum (Vaseline®) on <1000 gram babies to reduce transepidermal water loss. However, an extremely low birth weight baby < 28 weeks should be transferred to a humidified incubators once lines are inserted. Plastic sheeting with humidification, is also used to reduce fluid losses for babies who are on radiant warmers.
- Small babies, particularly those with arterial lines in and those on multiple medications can end up receiving a large amount of their projected daily fluid and sodium requirements as flushes. The nurses record these for intensive care babies and the total volume received in the last 24 hours should be noted. If it is significant, it should be recorded in the notes and discussed in the planning of the next day's fluids. The amount that is 'significant' may vary with the size of the baby, the severity of illness and the baby's particular problems, but if the flush volume exceeds 15% of the projected daily fluid intake, it is worth mentioning.

### Intravenous Fluids

**Dextrose**

- Usually 10% dextrose
- May need another dextrose concentration if hyperglycemic (7.5%) or hypoglycaemic (12.5-15%)
Sodium
- Commence sodium on the first or second day unless serum sodium is >140, discuss with the consultant on the morning or afternoon round.
- Subsequently, aim to keep the serum Na in the normal range (135-145 mmol/L). Remember that the baby may already be receiving a substantial amount of sodium via arterial lines and flushes so this intake may need to be calculated and subtracted from the amount given intravenously. Babies who have been fully orally fed and switch to IV therapy should have some sodium added from the beginning.

Potassium
- Usually, due to low renal potassium excretion, no or low levels of potassium are needed for the first one to two days. After that, provided the serum potassium is <5.5 mmol/L, start K to keep the serum K 3.5-5 mmol/L.

Calcium and Phosphate
- It is very difficult to ensure optimum calcium and phosphate without enteral feeding. The best we can usually ensure is that the ionized calcium from the blood gas analysis is in the normal range (>0.8 mmol/L) and that the serum P04 is in the normal range.
- The reason to use ionized calcium as the guide is that it is the ‘functional’ calcium and is also the regulator of calcium balance. The ratio of ionized to total calcium is higher in the presence of acidosis and hypoproteinemia, both of which are common in sick and premature babies.
- Aim for 1.5 mmol/kg/day calcium intake and a ratio of total Ca:P04 intake of 1.3:1. However, because calcium is a potent sclerosing agent, especially in very small veins, and can cause severe tissue necrosis if the IV infiltrates, it is impossible to give this much through a peripheral IV.
- Ca gluconate in a peripheral IV line should not exceed 15 ml of 10% solution in 500ml
- Ca gluconate in a central line should not exceed 50 mls of 10% solution in 500 ml and for ease of charting 30mls in 500ml bag has been chosen to be the standard amount in babies without specific Ca issues
- In an emergency (e.g. cardiac arrhythmia, decreased cardiac output thought to be secondary to hypocalcemia, during exchange transfusion), a bolus of 0.5 ml/kg 10% calcium gluconate, diluted 1:5 can be given over 15-20 min via a syringe pump or as a slow push. You can use a good peripheral IV for this if necessary, but the infusion site must be watched closely.
- Never infuse calcium-containing solutions into peripheral arterial lines, and only into umbilical arterial lines after specific discussion with a consultant, and weighing the alternatives.

Prescribing Additives

Day 1
- If unable to receive full enteral feeds and TPN is not required then start 10% dextrose at 65ml/kg/day

Day 2 onwards
- Most babies will require electrolyte additives by 24-48 hours
- Change fluids to the 500ml premix fluid bag containing 10% dextrose, 15mmol NaCl, 10mmol KCl
- Fluid rate will usually be 90ml/kg/day on day 2 and volumes graded up as discussed on the ward round
- The more volume the baby gets the more additives they will receive in the same way as TPN is given
- The bag can be hung for 48 hours before being changed unless Ca is added to the bag when it should be changed 24 hourly
- Ca should be replaced in babies that are proven to be hypocalcaemic (ionised Ca <0.8, formal < about 1.8)
- Usually babies on fluids with additives will be receiving some milk enterally which should prevent hypocalcaemia. If they are not receiving milk over a few days then TPN may be a better fluid option.
- If Ca gluconate is being given peripherally – add 3.3mmol (15mls) per 500ml bag (no need to remove 15ml first)
- If Ca gluconate is being given centrally – add 6.6mmol (30mls) per 500ml bag (no need to remove 30ml first)
- Below are the amounts of electrolytes that will be provided at the fluids rates stated.
Individual Fluids with Additives
- If a baby has complex electrolyte requirements and an individual TPN bag is unable to be obtained or is not felt to be required then individual fluids with additives can be prescribed as per the instructions below.
- The Electrolyte Calculator on the G:Drive in the Drugs Folder can be used to calculate doses and volumes as well.
- Standard solutions used for preparation of IV fluids:

<table>
<thead>
<tr>
<th>Stock Solution</th>
<th>Typical Dose</th>
</tr>
</thead>
<tbody>
<tr>
<td>NaCl 23%</td>
<td>4mmol/ml</td>
</tr>
<tr>
<td>KCl 15%</td>
<td>1mmol/ml</td>
</tr>
<tr>
<td>Ca Gluconate 10%</td>
<td>0.22mmol/ml</td>
</tr>
</tbody>
</table>

Start at the lower end and increase if there is a significant deficit

<table>
<thead>
<tr>
<th>Step</th>
<th>Example for 1.2 kg baby</th>
</tr>
</thead>
<tbody>
<tr>
<td>a)</td>
<td>Decide the total fluids for the day.</td>
</tr>
<tr>
<td></td>
<td>150 ml/kg = 150 x 1.2 = 180 ml</td>
</tr>
<tr>
<td>b)</td>
<td>Subtract non-nutritional fluid, such as arterial line and other infusions</td>
</tr>
<tr>
<td></td>
<td>UAC at 1 ml/h = 24 ml/day</td>
</tr>
<tr>
<td></td>
<td>=&gt; IV fluid is (180-24) = 156 ml/day</td>
</tr>
<tr>
<td></td>
<td>≡ 6.5 ml/hr</td>
</tr>
<tr>
<td>c)</td>
<td>Decide planned electrolyte intake.</td>
</tr>
<tr>
<td></td>
<td>3 mmol/kg Na</td>
</tr>
<tr>
<td></td>
<td>2 mmol/kg K</td>
</tr>
<tr>
<td></td>
<td>1.5 mmol/kg Ca</td>
</tr>
<tr>
<td>d)</td>
<td>Multiply by the ratio of 500/IV fluid volume (from step b) because the nurses make up 500 ml bags of fluid.</td>
</tr>
<tr>
<td></td>
<td>NaCl: 3mmol x 1.2kg x 500ml bag /156ml =11.5 mmol</td>
</tr>
<tr>
<td></td>
<td>KCl 2mmol x 1.2kg x 500ml bag /156ml =7.7 mmol</td>
</tr>
<tr>
<td></td>
<td>CaGluc1.5mmol x 1.2 x 500ml bag/156ml=5.8 mmol</td>
</tr>
<tr>
<td>e)</td>
<td>Convert the mmol dose to the volume of stock solution to be added to the 500 ml bag (see above).</td>
</tr>
<tr>
<td></td>
<td>NaCl 11.5mmol/4mmol = 2.8 ml</td>
</tr>
<tr>
<td></td>
<td>KCl 7.7mmol/1mmol = 7.7 ml</td>
</tr>
<tr>
<td></td>
<td>CaGluc 5.8mmol/0.22mmol = 26.4 ml</td>
</tr>
<tr>
<td>f)</td>
<td>Check that the calcium concentration is appropriate to the type of line being use</td>
</tr>
<tr>
<td></td>
<td>Peripherally max amount = 15 mls in 500ml bag</td>
</tr>
<tr>
<td></td>
<td>Centrally max amount = 50mls in 500ml bag.</td>
</tr>
<tr>
<td>g)</td>
<td>Add 0.1 unit heparin per ml if infusing through anything other than a peripheral IV.</td>
</tr>
<tr>
<td></td>
<td>0.1 unit/ml of heparin in a 500ml bag</td>
</tr>
<tr>
<td>h)</td>
<td>Check that the total volume of additives doesn’t exceed 50 ml (i.e. 10% of the volume in a 500 ml bag). If it does, write the prescription with instructions to remove a volume equivalent to the total volume of additives first (to avoid major dilution errors in the electrolytes).</td>
</tr>
</tbody>
</table>
Thus, the fluid prescription for the baby in the example (if CVL) would be:

10% dextrose 500 ml. Add:
2.8 ml 23% NaCl (4mmol/ml) (3 mmol/kg/day)
7.7 ml 15% KCl (1mmol/ml) (2 mmol/kg/day)
26.4 ml 10% Ca Gluconate (0.22mmol/ml), or (1.5 mmol/kg/day)
0.1 unit/ml heparin (0.05ml of 1000 units/ml solution) Run at 6.5 ml/hour

If you write out this detail, your prescription can be readily checked by the nurses.

### Calculating Dextrose Concentrations

- **To make 500ml bags of:**
  - 7.5% dextrose
    - Add 250ml 5% and 250ml 10% dextrose
  - 12.5% dextrose
    - Add 470ml 10% and 30ml 50% dextrose
  - 15% dextrose
    - Add 440ml 10% and 60ml 50% dextrose
  - 20% dextrose
    - Add 380ml 10% and 120ml 50% dextrose

- **To make a 10% premix iv fluid bag up to 12.5% dextrose**
  Remove 30mls from the 500ml premix bag and replace with 30mls of 50% dextrose
  Concentration of additives will now be:


### Hyponatraemia (Na+ < 135mmol/L)

Hyponatraemia can be caused by:
- Increased water retention eg: renal failure, SIADH after an asphyxial insult, early days of RDS
- Increased Na losses eg: leaky preterm kidneys, 3rd spacing after surgery, diuretics

**Mild Hyponatraemia (Na ≥ 130mmol/L)**
- Start or increase the oral supplements if on half enteral feeds
- Increase the amount of Na additives to the 10% dextrose maintenance fluids
- Increase the TPN rate to provide more Na per day
- Change to High Na TPN
If none of the above are appropriate – for example in a baby on Standard Na TPN at 165ml/kg/day who is NBM because of aspirates and can’t have oral supplements and a High Na TPN bag is unavailable – a Na sideline needs to be started

**Sodium Sideline**
- Calculate the Na deficit using the formula: \((\text{Target Na} – \text{Current Na}) \times 0.6 \times \text{weight (kg)} = \text{mmol deficit}\)
- Target Na is usually 135mmol/L
- Add the deficit to a 10% dextrose sideline and infuse over 24 hours
- The maximum concentration the infusion should be is **0.8 mmol/mL**

  eg: 1.5kg baby, Na 131mmol/L  
  \((135 - 131) \times 0.6 \times 1.5 = 3.6\text{mmol Na deficit}\)
  Take 3.6mmol Na (0.9ml of 4mmol/ml NaCl) and make up to a total of 12 mL with 11.1ml 10% dextrose, infuse at 0.5ml/hr for 24 hours.
  Concentration of the infusions is 3.6mmol ÷ 12mL = 0.4 mmol/mL

**Severe Hyponatraemia (Na < 130 mmol/L)**
If the Na is below 130mmol/L it is best to firstly correct the deficit with a Na correction and then increase the daily maintenance provided by the methods in “mild hyponatraemia” above or order an individual TPN bag with an increased concentration of Na.

If the baby is well and is on mainly enteral feeds then the daily sodium supplements could be increased and the sodium corrected over 24 hours. Using this method would depend on how low the sodium is, the rate of fall of the sodium level and how well the baby is.

**Sodium Correction**
- Calculate the Na deficit using the formula: \((\text{Target Na} – \text{Current Na}) \times 0.6 \times \text{weight (kg)} = \text{mmol deficit}\)
- Make up a **0.8 mmol/mL** solution in 10% dextrose as per the sodium chloride correction infusion sheet
- Replace the deficit usually over 6 hours but the larger the deficit the slower the replacement should be
- Infuse the fluid into the same line as the TPN as this will further dilute the infusion as it enters the vein

**Hypokalaemia**
\((K^+ < 3.5 \text{ mmol/L})\)
Hypokalaemia can be caused by:
- Insufficient maintenance eg: K⁺ additives not introduced into maintenance fluids on day 2, on TPN but due to multiple other infusions the amount of K received is minimal
- Increased K losses eg: vomiting, 3rd spacing after surgery, diuretics, renal disorders, CAH
- Other eg: insulin, alkalosis, drugs such as fluconazole, amphotericin,

**Mild Hypokalaemia (K⁺ ≥ 2mmol/L )**
- Start oral supplements at 1-2mmol/kg/day (potassium chloride or potassium dihydrogen phosphate) hourly if on half enteral feeds, or
- Increase the rate of the premix 10% dextrose bag to provide more K, or
- Increase the K additives to an individually prescribed 10% dextrose fluid bag
- Increase the TPN rate to provide more K, or
- Prescribe an individual TPN bag with increased K in it, or
- Prescribe a potassium sideline to run over 24 hours (leave as a last line option as concentrated K infusions are not without risk and they take up a lot of the daily fluid volume)
Potassium Sideline. Take extreme care in calculating
- Replace 1-2 mmol/kg KCl over 24 hours by infusing a 40mmol/L concentration solution over 24 hours (see drug protocols for guidance on making up the 40mmol/L concentration solution)
  eg: 1.3 kg baby needing 2 mmol/kg/day additional KCl to be infused

  Calculate the volume of 40mmol/L solution needed to replace 2 mmol/kg over 24 hours

  \[
  \text{Dose in mmol KCL} \times \frac{1000}{40} = \text{mls of total infusion volume}
  \]

  \[
  2 \times 1.3 \times 1000 = 65 \text{mls}
  \]

  Draw up 65mls of the 40mmol/L solution and infuse at a rate of 2.7ml/hr for 24 hours
- Infusion can be made up to a 60mmol/L concentration if a central line is used and there is a need to fluid restrict.

Symptomatic or Severe Hypokalaemia (K⁺ ≤ 2.0 mmol/L)
- Acute treatment is needed to correct the deficit and a potassium correction is needed
- Only use in the most severe cases as high concentration potassium infusions are not without risk
- Signs of symptomatic hypokalaemia that are seen in neonates are ECG changes (ST segment depression, low-voltage T waves, U wave.). Other signs include neuromuscular weakness, ileus, urinary retention
- Use the potassium chloride infusion sheet to calculate the potassium infusion (do not use KH₂PO₄)
- The replacement is calculated to replace 0.6 mmol/kg over 4 hours using a 40mmol/L solution if via a peripheral line and a 60mmol/L solution if there is a central line and a need to restrict the volume.
- The rate at which the infusion can be infused should not exceed 0.2 mmol/kg/hr

Hypernatremia
- Defined as a serum sodium level > 146 mmol/L
- ELBW babies need 6-12 hourly electrolytes on ABG or formal blood tests in the first few days as they are at a high risk of developing hypernatremia

Risk Factors
- Insensible water loss
- Iatrogenic (Na bicarbonate, Na in fluids or TPN, arterial line infusion, oral supplements)
- Gastrointestinal loss of fluid eg: 3rd space loss in NEC, post surgery, high bile stained NG loss
- Glycosuria causing an osmotic diuresis
- Diabetes insipidus (central or nephrogenic)
- Sample technique (eg. excessive squeezing from a heelprick)

Diagnosis
- High serum Na on a formal lab test or an upward trend on ward gas machine results (these may be very different and if in doubt repeat as a formal lab specimen)
- Weight loss and decreased urine output suggest dehydration.
- Dilute urine with normal or increased urine output suggest glycosuria or diabetes insipidus
- Na in 1st 48 hours almost always reflects free water deficit.
- Increased Na in 1st 48 hours does not equal increased total body Na

Treatment
- In the first few days of life increase total daily fluid (water) intake
- Increase fluids by 30ml/kg/day increments (eg. 90 to 120ml/kg/day) either as an increase in the maintenance fluids or TPN or as an additional sideline of 10% dextrose.
- If the Na rises despite 180 mls/kg/day intravenously, a continuous infusion of water nasogastrically at 40 mls/kg/day can be used. Eg. 0.6kg infant: 0.6 x 40 = 24 mls / 24 hrs run at 1 ml / hr NG. Stop when Na falls to 150mmol/l.
- If there is evidence of fluid overload (oedema, weight gain, rising Na) then restrict Na and water intake.
- After the first 48 hours of age, hypernatremia is still likely to be due to a free water deficit, so increase maintenance volume but be careful with added electrolytes.
- If TPN prescribed with more Na⁺ than required reduce volume and add a sideline of dextrose.
Hypernatraemic Dehydration

- Some babies are readmitted to NICU from the community for management of moderate-severe hypernatraemic dehydration. By far the commonest reason is unidentified inadequate intake of milk.

- There may be risk factors able to be identified in the mother and baby:
  Mother: eg: primip, prolonged and difficult labour, antepartum or postpartum haemorrhage, maternal illness, delayed onset of lactation
  Baby: twin, preterm, IUGR, congenital abnormalities, mouth or jaw structural abnormalities.

Refer to the Clinical Practice Recommendation for management of moderate-severe hypernatraemic dehydration

Hyperkalaemia

- Defined as a serum potassium level > 6mmol/L
- Hyperkalaemia is a life threatening condition and when identified, needs to be managed promptly and aggressively.
- Hyperkalaemia most commonly occurs around 24-48 hours of age in the unwell, preterm infant < 28 weeks
- Always discuss treatment plan with the consultant before commencing

Risk factors

- Extreme prematurity
- Haemodynamic instability with low systemic blood flow
- Hypoxic ischaemic encephalopathy
- Acute renal failure
- Hypothermia
- Blood transfusion with old blood
- Sepsis
- Metabolic acidosis

Signs of Hyperkalaemia

- ECG changes with peaked T waves
- Widened QRS complexes
- Arrhythmias

Those babies identified as having risk factors for hyperkalaemia should have regular K levels checked in the first 24-48 hours ie: 6-12 hourly, from a free flowing sample to remove the effect of haemolysis on the potassium level.

Mild Hyperkalaemia: 6 - 7mmol/L

- Stop any potassium containing infusions or medications
- Stop any potassium sparing diuretics eg: spirinolactone
- Correct any hypocalcaemia that may be present (aim to keep ionised Ca >1.2mmol/L)
- Correct any metabolic acidosis with sodium bicarbonate and by treating the underlying cause

Moderate Hyperkalaemia: 7- 8 mmol/L

- As above
- Insulin/dextrose infusion (see drug protocol sheets and below)
- Salbutamol infusion (see drug protocol sheets and below)

Severe Hyperkalaemia: >8 mmol/L or any raised level with arrhythmias

- 10% calcium gluconate infusion immediately to stabilise the myocardium
- Management as above
Insulin/Dextrose Infusion
- Effective in treating hyperkalaemia.
- Rigorous monitoring of the blood sugars (as well as the K levels) is essential as the complications of the infusion can include hyperglycaemia and hypoglycaemia.
- A central line is required as the dextrose concentration of 25-50% is too high to go peripherally.

Salbutamol Infusion
- An infusion of salbutamol is also effective in temporarily lowering the potassium (for 1-2 hours) and may be easier to prepare quickly in an emergency situation rather than an insulin/dextrose infusion.
- However, the use of salbutamol for non-oliguric hyperkalaemia in the extreme preterm infant has not been as extensively studied and as such many units would use salbutamol as a second-line treatment for hyperkalaemia not responding to insulin/dextrose infusion.
- Salbutamol infusion does not lower the potassium level in babies with persistent metabolic acidosis
- Side effects of using salbutamol include, tachycardia, tremor, hyperglycaemia.

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- Stop any potassium containing infusions or medications
- Stop any potassium sparing diuretics eg: spironolactone
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NEONATAL NUTRITION GUIDELINES

Feeding Guidelines

<table>
<thead>
<tr>
<th>Babies born &lt; 32 weeks OR Birthweight &lt; 1800g * Refer to dietitian at any time that growth is inadequate*</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Transition to Oral Feeding</strong></td>
</tr>
<tr>
<td>Feed <strong>TPN ➔ Breast milk, PDM or standard milk mixture</strong></td>
</tr>
<tr>
<td>If &lt;32 weeks or &lt;1500g start Infloran probiotic on the day after birth (for prevention of NEC)</td>
</tr>
<tr>
<td><strong>Frequency</strong> Start feeds and grade up as per the chart below</td>
</tr>
<tr>
<td>Aiming for hrly feeds &lt;1250g and 2hrly feeds 1250-1800g</td>
</tr>
<tr>
<td>Include feeds when:</td>
</tr>
<tr>
<td>&lt;1000g:0.5ml/hr or 1ml 2hrly ≥1000g:1ml/hr</td>
</tr>
<tr>
<td><strong>Feed/Fluid Volume</strong> Day 1 90 ml/kg/day on TPN 65ml/kg/day if not TPN</td>
</tr>
<tr>
<td>Day 2-3 90-120 ml/kg/day</td>
</tr>
<tr>
<td>Day 3-5 120-150 ml/kg/day</td>
</tr>
<tr>
<td>Day 5-7 150-165 ml/kg/day</td>
</tr>
<tr>
<td>Day &gt;7 165 ml/kg/day</td>
</tr>
<tr>
<td>Can increase TPN to 165ml/kg/day when oral feeds &gt;40ml/kg/day if required on SMO/dietitian advice</td>
</tr>
<tr>
<td><strong>Supplements</strong> At 100ml/kg/day enteral feed stop lipid, and start HMF and vitamins:</td>
</tr>
<tr>
<td>Vitamin D - Two drops/day if ≤1000g, or, One drop/day if &gt;1000g</td>
</tr>
<tr>
<td>Vitamin A – Three drops/day if &lt;30 weeks, or Two drops/day if 30-34wks</td>
</tr>
<tr>
<td>Micelle E 30IU/day &lt;1250g or&lt;30wks</td>
</tr>
<tr>
<td>Folic Acid 50µg/day &lt;1500g if unfortified EBM long term</td>
</tr>
</tbody>
</table>
Babies born at ≥ 32 weeks AND Birthweight > 1800g
* Refer to dietitian at any time that growth is inadequate*

<table>
<thead>
<tr>
<th>Transition to Oral Feeding</th>
<th>Stable Growing</th>
<th>Preparation for Discharge</th>
<th>At Discharge</th>
</tr>
</thead>
<tbody>
<tr>
<td>Feed</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Breast milk, PDM or</td>
<td>Breast milk, PDM or</td>
<td>Breast milk, PDM or</td>
<td>Breast milk or Standard milk mixture</td>
</tr>
<tr>
<td>Standard milk mixture</td>
<td>Standard milk mixture</td>
<td>Standard milk mixture</td>
<td></td>
</tr>
<tr>
<td>Frequency</td>
<td>3 hourly unless risk of hypoglycaemia or other medical concerns</td>
<td>3 hourly feeds</td>
<td>7-8 feeds per day or responsive feeding</td>
</tr>
<tr>
<td>Feed Volume</td>
<td>Day 1 65ml/kg/day, but, 90ml/kg/day if on TPN Day 2 90ml/kg/day Day 3-4 120ml/kg/day Day 5-7 150ml/kg/day Day &gt;7 150-165 ml/kg /day Include feeds when 1ml/hr</td>
<td>180ml/kg/day or as tolerated</td>
<td>150-165 ml/kg/day as tolerated or responsive feeding</td>
</tr>
<tr>
<td>Supplements</td>
<td>Vitamin D one drop/day Vitamin A two drops/day if &lt;35 weeks</td>
<td>Start Fe at 4 weeks at 3mg/kg/day if no recent blood transfusion, not on FM85 fortifier or Preterm formula, MM At 36 weeks, if off respiratory support then stop Vitamin A Some on Preterm MM might need some Fe supplementation after dietician review</td>
<td>Vitamin D one drop/day to 1 year of age Iron 3mg/kg/day if breastfed or mixed feeds breastmilk and milk mixture to 1 year of age Stop Iron if on exclusive milk mixture Stop Vitamin A if still on it</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Weight of Infant</th>
<th>Initial Volume and Frequency of Feeds</th>
<th>Rate and Volume to Increase Feeds (once tolerating introduction of feeds in previous column)</th>
<th>Days to Full Feeds (150ml/kg/day)</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt;1000 g</td>
<td>0.5ml Q4H x 24-48 hours 1ml Q4H x24 hours 1ml Q2H x24 hours 1ml Q1H x 24 hours</td>
<td>When on hourly feeds increase by 0.5ml 24QH then 0.5ml12QH</td>
<td>10-12days</td>
</tr>
<tr>
<td>1000-1250g</td>
<td>1ml Q4H x 24-48 hours 1ml Q2H x 24 hours 1ml Q1H x 24 hours</td>
<td>When on hourly feeds increase by 1ml 24QH then 1ml12QH</td>
<td>8-10 days</td>
</tr>
<tr>
<td>1250-1799 g</td>
<td>2ml Q2H x 48 hours</td>
<td>Increase by 1ml Q8H Then 1-2ml Q6H</td>
<td>5-8 days</td>
</tr>
<tr>
<td>≥ 1800g</td>
<td>65ml/kg/day aiming for feeds Q3H</td>
<td>Titrate IV fluids with EBM/PDM</td>
<td>3-5 days</td>
</tr>
</tbody>
</table>
Transpyloric Feeds

Transpyloric feeding has several potential advantages over gastric feeding, including improved feeding tolerance and prevention of bronchopulmonary dysplasia, presumably due to reduced risk of aspiration (Wallenstein et al. 2019). However, definitive evidence is lacking (Watson & McGuire 2013), so it should be used with caution.

Indications

- Gastroesophageal reflux causing feeding intolerance or respiratory compromise
- Preterm infants with severe lung disease
- Older infants with severe bronchopulmonary dysplasia
- Note: placement in children post diaphragmatic hernia and other abdominal conditions is more difficult and may need interventional radiology assistance.

Procedure (From Clifford et al. 2017, Starship protocol has an alternative approach)

1. This is a joint nursing and medical effort
2. Ensure baby has not had a feed at least 1-2hrs prior to insertion to prevent vomiting and aid stomach motility.
3. Obtain 5 or 6 French feeding tube. Place in fridge for 20-30 minutes. Enteral polyurethane feeding tube such as a Corflo 6fg/91cm is preferred. Remove the stylet it is not to be used with insertion. Note: If using Standard PVC feeding tube for transpyloric placement it will need to be changed weekly.
4. Flush tube with sterile water.
5. Calculate the gastric distance and the advancement distance by measuring the distance from umbilicus to right iliac crest. **Final length of tube should = standard gastric length + distance from umbilicus to right iliac crest**
6. Swaddle baby and give oral sucrose (to provide comfort)
7. Place tube to usual gastric distance per standard protocol
8. Put baby right side down and elevate head of bed to 30-45 degree angle
9. Slowly inject 5-10ml air into the gastric tube as you advance the tube 1-2cm at a time to estimated postpyloric distance position. Listen for air entry over the pylorus (to right of the midline) with the final ml of air.
10. Secure tube well so dislodgement is minimised.
11. Leave the baby right side down for another hour and then obtain an AP abdominal xray
12. Any duodenal position is acceptable, examples shown below:

![Nasoduodenal tube tip in the first](image1)
![Nasoduodenal tube tip in the second, and](image2)
![Nasoduodenal tube tip in the third portion of the duodenum](image3)

Feeding

- **Continuous feeds only**
- Convert total daily enteral fluid volume to ml/hr for continuous feeds.

Contraindications

- History of NEC, perforation, or other intestinal pathology. Always discuss with consultant before initiating.
- **DO NOT USE FORMULA** for transpyloric feeds, use expressed breast milk or PDM only. Additives can be used in discussion with the dietician.
- In rare circumstances specific formulas can be used – this needs dietician and Paediatric Gastroenterologist input.
**Medications**

Most medications are adequately absorbed in the duodenum and can be given at the same dose.

- **Medications safe to give via transpyloric tube:**

  amoxicillin, vitamin C, vitamin E, caffeine, calcium, chlorothiazide, digoxin, doxapram, erythromycin, fluconazole, folic acid, furosemide, lactobacillus, levetiracetam, levothyroxine, metoprolol, omeprazole, phenobarbitone, propranolol, ranitidine, sildenafil, and ursodeoxycholic acid.

- **Medications that can be given by transpyloric tube but are not adequately absorbed if tube placement is in the jejunum:**

  calcium, digoxin, erythromycin, folic acid, vitamin E.

- **Medications that cannot be given by transpyloric tube:**

  aspirin and ferrous sulfate

Check with pharmacy regarding all other medications.

**References**

**Dietitian Referral Guidelines**

Any infant with:
- Poor weight gain
- Fluid restricted enteral feeds
- Intolerance to breast milk fortifier or preterm formula
- Extended TPN use
- Surgical GI resection +/- ostomy formation
- NEC
- Short Bowel syndrome
- Malabsorption
- Chronic lung disease where growth impairment is likely
- Congenital Heart disease
- Gastrointestinal anomaly
- Metabolic disorder
- Chylothorax
- Renal Failure
- Osteopenia requiring additional calcium and phosphate
- Possible Zinc deficiency
- Breastfed baby where there are concerns about maternal nutrition

### Calories

<table>
<thead>
<tr>
<th></th>
<th>kcal/ml (unless other units given)</th>
<th>g/100ml (unless other units given)</th>
</tr>
</thead>
<tbody>
<tr>
<td>EBM</td>
<td>0.66</td>
<td>1.27g protein</td>
</tr>
<tr>
<td>EBM with HMF (1 sachet/25ml)</td>
<td>0.83</td>
<td>2.9g protein</td>
</tr>
<tr>
<td>PDM</td>
<td>0.66</td>
<td>1.1g protein</td>
</tr>
<tr>
<td>PDM with HMF (1 sachet/25ml)</td>
<td>0.83</td>
<td>2.7g protein</td>
</tr>
<tr>
<td>Milk mixture (S26)</td>
<td>0.67</td>
<td>1.5g protein</td>
</tr>
<tr>
<td>Milk mixture (Karicare)</td>
<td>0.67</td>
<td>1.4g protein</td>
</tr>
<tr>
<td>Premature milk mixture (PreNAN)</td>
<td>0.80</td>
<td>2.9g protein</td>
</tr>
<tr>
<td>Pepti Junior</td>
<td>0.67</td>
<td>Whey hydrolysate</td>
</tr>
<tr>
<td>Elecare</td>
<td>0.68</td>
<td>Amino Acid formula</td>
</tr>
<tr>
<td>TPN Protein</td>
<td>4 kcal/g</td>
<td>0.9 g protein/g</td>
</tr>
<tr>
<td>Protifar</td>
<td>3.8 kcal/g</td>
<td>Start at 1g/100mls and discuss with dietitian</td>
</tr>
<tr>
<td>Duocal (lipid and carbohydrate)</td>
<td>4.7 kcal/g</td>
<td></td>
</tr>
<tr>
<td>Liquigen</td>
<td>4.5kcal/ml (9kcal/g)</td>
<td></td>
</tr>
<tr>
<td>Calogen</td>
<td>4.5kcal/ml (9kcal/g)</td>
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<tr>
<td>Polycal</td>
<td>3.8kcal/g</td>
<td>0.94g CHO/g</td>
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<tr>
<td>Lipid</td>
<td>9 kcal/g</td>
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</tr>
<tr>
<td>TPN Protein</td>
<td>4 kcal/g</td>
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</tr>
<tr>
<td>Liquigen</td>
<td>4.5kcal/ml (9kcal/g)</td>
<td></td>
</tr>
<tr>
<td>Calogen</td>
<td>4.5kcal/ml (9kcal/g)</td>
<td></td>
</tr>
<tr>
<td>Dextrose</td>
<td>3.4 kcal/g</td>
<td></td>
</tr>
<tr>
<td>5% Dextrose</td>
<td>0.19</td>
<td></td>
</tr>
<tr>
<td>10% Dextrose</td>
<td>0.38</td>
<td></td>
</tr>
<tr>
<td>12.5% Dextrose</td>
<td>0.48</td>
<td></td>
</tr>
<tr>
<td>15% Dextrose</td>
<td>0.57</td>
<td></td>
</tr>
</tbody>
</table>
HMF

- Breast milk does not supply enough calories or nutrients for a growing preterm baby
- Growth is important for longer term developmental outcome and early nutrition is important
- CWH audits have shown that as babies wean off TPN there is a drop in protein supply around days 9-15 as the breast milk volumes increase and TPN volumes decrease
  - At CWH babies reach full enteral feeds (150ml/kg/day) by day 11
  - Full HMF supplementation takes until day 13 to occur
  - Starting HMF earlier can reduce or prevent this drop off in protein delivery
- ESPGHAN guidelines recommend a protein intake of:
  - 3.5 to 4.0g/kg/day for 1-1.8kg babies
  - 4.0 to 4.5g/kg/day for <1kg babies

HMF

- HMF provides calories, protein, calcium, phosphate, vitamins and electrolytes
- The whey protein component is hydrolysed to break this down to smaller components
- 1 sachet per 25mL of EBM is a full dose
- Start when on 100ml/kg/day enteral feeds the same day lipid stops
- Parent information sheet to be given to parents prior to starting HMF
- Make sure any phosphate supplements are stopped when HMF is started as the phosphate levels predictably become too high

Comparison of EBM with and without HMF

<table>
<thead>
<tr>
<th>Protein (g/kg/day)</th>
<th>Sodium (mmol/kg/day)</th>
<th>Phosphate (mmol/kg/day)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>EBM</strong></td>
<td><strong>EBM + HMF</strong></td>
<td><strong>EBM</strong></td>
</tr>
<tr>
<td>150 mL/kg/d</td>
<td>1.9</td>
<td>4.3</td>
</tr>
<tr>
<td>165 mL/kg/d</td>
<td>2.1</td>
<td>4.8</td>
</tr>
<tr>
<td>180 mL/kg/d</td>
<td>2.3</td>
<td>5.2</td>
</tr>
</tbody>
</table>

Indications for HMF

- <32 weeks gestation
- <1800g at birth
- Poor weight gains in babies outside the above criteria ie: crossing centiles
- On the advice of the dietitian if indicated by abnormal ALP, phosphate or albumin levels

When to Start HMF

- Start when the baby is on enteral feeds of 100ml/kg/day
- This volume needs to be calculated and documented on the bottom of the TPN worksheet
- TPN/Lipid/HMF transition
  - 80ml/kg/day enteral feed: Wean lipid to 2g/kg/day
  - 100ml/kg/day enteral feed: Stop lipid and start HMF

Precautions/Contraindications to HMF

- Suspected or proven NEC
- Significant abdominal distension with discolouration
- Blood in stool
- Heavily bile-stained aspirates
- Vomiting – small spills not an indication to withhold HMF
- Aspirates > 50% feed volume since last aspirate
- Sepsis
- Indomethacin course – if stable on HMF do not stop but if not on HMF then withhold until indomethacin course is completed
When to Stop HMF
- On dietitian advice
- Gestation, weight, ALP, albumin, calcium and phosphate levels all need to be considered before stopping HMF and will be individualised
- Can reduce volume of feeds and continue HMF if required
- As a general rule most babies can stop HMF at about 36 weeks or 2.5k

HMF after NEC
- No clear evidence that HMF alone is causative in the pathophysiology of NEC
- After recovery from NEC reintroduction of HMF should be discussed as having NEC is not an absolute contraindication to restarting HMF
- If a baby does not restart HMF then consideration will have to be made regarding the appropriate supplementation which may include phosphate, protifar, Liquigen and folic acid

Food Supplements and Vitamins
Breast feeding is encouraged whenever possible. However, premature babies and growth restricted babies < 1800g and < 32 weeks gestation at birth are likely to need supplements to breast milk, which may include the following:

- **Probiotics**
  Infloran is the product available and contains lactobacillus and bifidobacterium species. It is to be started the day after birth for babies <32 weeks or <1500g as a preventative measure for NEC. Mix one capsule with 1mL of milk or water (if insufficient EBM) and give with a feed if in milk or between feeds if mixed in water. Stop at 36 weeks corrected age. Probiotics to continue past 36 weeks for individual patients and should continue if there is a transition from EBM/PDM to formula feeds.

- **HMF**
  HMF is a whey protein hydrolysate used to fortify breast milk. Commence with 1 sachet per 25mls of EBM when infants are on 100ml/kg/day enteral feeds. This will add 17.2kcal/100ml and 1.6g of protein /100ml. Each sachet weighs 1g. If additional protein is required see section on Protifar. Infants cannot receive HMF after discharge so if additional calories are still required at discharge consult the Dietitian.

- **Standard formula**
  This is called milk mixture and provides 67-69kcal /100ml. The NICU rotates 2 brands of ready to feed milk mixture, S26 and Karicare. Formula rotation is required to meet BFHI standards. Parents who are going home formula feeding need to receive advice on formula preparation and the type of formula to purchase depending on the baby’s requirements.

- **Low birthweight formula**
  This is called premature milk mixture (PreNan) and provides 80kcal /100ml. There is no preterm formula available on discharge.

- **Protifar**
  A casein protein concentrate that provides additional protein with low levels of calcium and phosphorus and minimal sodium and potassium. The dietitian will determine quantity. 1g protifar will provide 3.8 kcal/g and 0.9 g protein. May be required if a baby is intolerant to HMF, has had NEC or is very growth restricted.

- **Liquigen**
  A medium chain triglyceride, liquid emulsion used to provide additional calories. It does not require pancreatic lipase or bile acids for digestion and absorption. Start at 1 g/kg/day and increase stepwise to 3 g/kg/day. Chart in drug chart. The dose is divided 8 hourly. 1g Liquigen provides 9.0kcal/g. Check with dietitian to ensure the calorie to protein ratio is ideal.

- **Calogen**
  A lipid emulsion of long chain triglycerides used to provide additional calorie (contains no MCT). Start at 1 g/kg/day and increase stepwise to 2 g/kg/day. Chart in drug chart. The dose is divided 8 hourly or more frequently if required. 1g Calogen provides 9.0kcal/g. Check with dietitian to ensure the calorie to protein ratio is ideal.
• **Duocal**
  A carbohydrate/fat mixture, which is particularly useful in bigger babies who have increased energy requirements for various reasons. Discuss with the Dietitian. Start at 1g/100ml and increase up to 2g/100ml. 1g Duocal provides 4.7kcal/g. Special application is required for infants going home on Duocal which will be prescribed and arranged by the Dietitian.

• **Pepti Junior Gold or Neocate/Elecare are special formulas**
  Pepti Junior Gold is a semi elemental protein hydrolysate and Neocate/Elecare are a fully elemental formula. Discuss with the Dietitian regarding best formula to use. Premature infants on special formula may require additional calories, protein, electrolytes and minerals discuss with the Dietitian.

• **Specialised Formula for specific medical conditions:**
  These formulas will need to be ordered from Auckland and will generally be an overnight delivery. The Dietitian will arrange if needed.
  
  **Monogen** formula is used for chylothorax
  **Kindergen** is a specialised renal formula
  **Other specialised formulas** for metabolic conditions are available depending on condition

• **Food Thickener**
  A maize starch used to thicken infant formula for infants that spill. The standard dose is 1.5 g/100ml. This will provide 3.6kcal/g or 5.4kcal/1.5 g dose.

• **Electrolyte Supplements**
  When a change is made from TPN or IV to enteral nutrition and the baby has been needing electrolyte supplements in excess of those supplied in the breast milk or formula, oral electrolyte supplements will need to be prescribed as sodium chloride or potassium chloride. Alternatively if also requiring phosphate supplements can use KH₂PO₄ or NaH₂PO₄ (not more than 2mmol/kg/day of either). More common in babies on diuretics or has some other reason to have unusually high obligate electrolyte losses. Urine sodium level less than 20mmol/L indicates sodium depletion. In general, the IV preparations as described in a previous section can be used orally. Distribute the dose 6-8 hourly. The same principle applies if a baby is on oral electrolyte supplements, these will need to be charted IV if the baby cannot feed

• **Phosphate**
  Premature infants are at risk of metabolic bone disease and may require phosphate supplementation. A rising ALP with or without a low phosphate is usually an indication for supplementation which is usually discussed on the ward round. Phosphate is usually given as KH₂PO₄ with a starting dose of 1 mmol/kg/day. Alternatively if also requiring sodium supplements and serum chloride level is elevated can use NaH₂PO₄. Remember that HMF is an alternative source of phosphate and may be commenced first. Additionally if KH₂PO₄ was commenced before HMF was started then it should be stopped once HMF is tolerated and the phosphate level has risen to > 2.0 mmol/l. Never exceed 2mmol/kg/day PO₄

• **Vitamin D**
  o All babies < 37 weeks or <2500g should be prescribed Vitamin D Puria two drops once daily if ≤1000g and one drop once daily if >1000g.
  o Other babies who may need Vitamin D supplementation include those with fat malabsorption (includes babies with Cystic fibrosis and those deemed high risk as per Ministry of Health Guidelines) (see below)

    *High risk term baby (NZ Ministry of Health Guidelines)*
    Breastfed baby and:
    - with naturally dark skin
    - baby’s mother is vitamin D deficient
    - sibling of baby has had rickets or seizures resulting from low blood calcium levels

  Note: Babies who are breastfed over winter months in New Zealand may also be vitamin D deficient by late winter/spring.

  • At discharge all babies who have been commenced on Vitamin D Puria in NICU will be discharged on ONE drop per day. It is recommended to continue supplementation until 12 months of age. See drug profile for further details.

• **Vitamin A**
  Retinol (vitamin A) is an important co-factor in many biochemical processes and may be preventative in the development of chronic lung disease. Previously preterm babies have received Vitamin A from Vitadol C drops but that was discontinued in March 2020 and a standalone Vitamin A preparation made available.
Babies born <30 weeks to have 3 drops daily the day after lipid stops
Babies born 30-34+6 weeks to have two drops daily
Stop supplementation at 36 weeks if off respiratory support
Not to routinely be discharged home on Vitamin A

Levels of Vitamin A are no longer checked at day 21 after sequential audits (2016 and 2018) have shown good levels with the current dosing regimen. However, babies with conjugated hyperbilirubinaemia on supplements may benefit from levels being checked to ensure absorption is occurring.

- **Folic acid**
  All babies < 1500g who are not receiving fortified breastmilk long term should receive 50 µg daily. These will be babies who have contraindications to HMF, have failed HMF or parents decline HMF. Folic acid will be started after these criteria have been fulfilled (and not when they are still grading up on feeds in the first week of life) Individual babies may have this continued after discharge but it is usually stopped. It is also given to babies who have haemolytic anaemia or required an exchange transfusion.

- **Vitamin E**
  All babies <1250g or <30 weeks will begin Vitamin E 30IU (0.2ml) when tolerating enteral feeds and off lipid

Levels are no longer checked after sequential audits (2016 and 2018) showed no improvement in the levels after increasing the dose to 50IU daily. However, babies with conjugated hyperbilirubinaemia on supplements may benefit from levels being checked to ensure absorption is occurring. It is stopped at discharge for most babies but those with malabsorption conditions may be discharged home on Vitamin E and may need special authority forms to be completed

- **Iron**
  Babies <2500g or <37 weeks gestation should receive iron 3mg/kg/day from 4 weeks of age until 12 months of age if receiving breastmilk or formula. Exclusions to this are babies on Preterm formula who should get enough iron from the formula but individual cases may need iron supplementation after dietician review. Babies on FM85 fortifier should not receive iron until this has been discontinued as it has 2mg/kg/day iron in it already. Recommend withholding starting iron at 4 weeks if the baby has been transfused in the previous 2 weeks as the iron stores will be sufficient from the transfusions. Stop iron at discharge if the baby is going home fully formula fed. We usually adjust the dose for weight while the baby is in the NICU, but if the baby is well and there is no ongoing blood loss, it is appropriate to let the baby gradually outgrow the dose after discharge, so write discharge prescriptions for 3 months supply with a rounded mL volume for ease of administration. Maximum dose of 2ml as doses above this are not tolerated well. For babies who have not started iron at the time of discharge we have taken a pragmatic approach to prescribe the iron dose for the predicted weight at 4 weeks of age after reviewing the growth chart. If predicted to be <3kg then dose is 1mL daily, 3-4 kg dose is 1.5mL and >4kg dose is 2mL. This provides between 2-3mg/kg/day of elemental iron.

- **Zinc**
  Take a zinc level if there are concerns that the baby may have acrodermatitis enteropathica (suspect if there is bad thrush or nappy rash and a negative culture for candida) or peripheral oedema where other causes have been excluded. Babies on loop diuretics are also at risk. Zinc levels are no longer routinely checked on babies <1000g at birth (audit Jan 2019).

Zinc deficient babies (serum zinc <10mmol/l) should have an extra 1-2 mg/kg/day (0.4-0.8ml/kg/day) elemental zinc, as zinc chloride. Supplied by pharmacy as a solution of 5mg elemental zinc per 2mls. Since Zn competes with Fe for uptake in the intestine, it can be advisable to stop Fe supplements for a few days while Zn supplementation begins. Recommend the Fe at a different time of day.

**Parenteral Nutrition**

- The aim of parenteral nutrition is to provide protein, fat, carbohydrate, electrolytes, trace elements and vitamins to allow nutritional needs to be met and to optimise growth.
- The preferred form of nutrition, however, is enteral with breastmilk, or if unavailable, milk mixture.
- If enteral feeds will not be possible within 3 days parenteral nutrition may be required.
- Premature babies, tolerate starvation much more poorly than older children, and so parenteral nutrition has a very important role to play in neonatal intensive care but it does increase the risk of nosocomial infection, nutrient imbalances and potential toxicity (aluminium)
Indications for TPN:

- All babies ≤30 weeks or <1500g
- Babies at 31 and 32 weeks if unlikely to achieve full enteral feeds by day 5
  - review risk factors for delayed milk availability or the need to introduce feeds slowly, ie: growth restriction, abnormal antenatal Dopplers, maternal illness, birth condition, multiple birth
- Congenital gastrointestinal anomalies eg: gastroschisis, omphalocele, bowel atresia
- Necrotising enterocolitis (NEC)
- Risk of NEC
  - Infants with absent or reversed umbilical artery Dopplers
  - Perinatal asphyxia

IV Access for TPN

- TPN is preferably administered through a central line – this would be via a UVC or longline
- TPN may be administered through a peripheral iv line if:
  - A central line cannot be placed but TPN is still required
  - The risks of a non-ideally positioned central line outweighs the risks of extravasation via a peripheral iv line.
  - The baby is likely to establish full feeds within 7 days and does not need long term iv access
  - Only the Starter Bag and Peripheral Standard TPN Bag may be given peripherally
- If a baby has not achieved 1ml/hr NG feeds by day 5 then plan to insert a longline
- Longlines can be inserted 7 days a week with weekends not being a barrier to line insertion with current staffing
- The iv access being used needs to be reviewed daily to ensure it is still appropriate for the clinical situation

TPN Components

<table>
<thead>
<tr>
<th>Fluid</th>
<th>Guideline</th>
</tr>
</thead>
<tbody>
<tr>
<td>Day 1</td>
<td>90ml/kg/day to target protein requirements and avoid hypematraemic dehydration</td>
</tr>
<tr>
<td>Day 2-3</td>
<td>90-120 ml/kg/day</td>
</tr>
<tr>
<td>Day 3-5</td>
<td>120-150 ml/kg/day</td>
</tr>
<tr>
<td>Day 5-7</td>
<td>150ml/kg/day</td>
</tr>
</tbody>
</table>

- 165ml/kg/day as long as enteral feeds are ≥40ml/kg/day to ensure total protein intake is not too high (does not apply to Term TPN as the protein content is lower)

Protein

- Primene is the protein product that is currently in use
- Hang bag for up to 48 hours
- Targeting at least 2g/kg/day on day 1 and increase to 4g/kg/day in preterms and 3g/kg/day terms
- Intolerance to protein can occur due to extreme prematurity, renal and liver impairment and can be seen with high urea levels, hyperammonemia or hyperaminoacidemia. This is a particular concern as amino acids such as phenylalanine are neurotoxic, particularly in the developing brain

Dextrose

- Provided as 10% or 12.5% dextrose
- Most infants need a minimum of 4-6 mg/kg/min glucose
- Preterm babies may not tolerate >8mg/kg/min (10% dextrose at 130ml/kg/day = 9mg/kg/min)
- Term babies may not tolerate >13mg/kg/min (12.5% dextrose at 130ml/kg/day =11.5mg/kg/min)
- Use glucose calculator link in the TPN folder to easily calculate these rates
- If the baby is hyperglycaemic review all other infusions and see if it is practicable to decrease the dextrose intake from 12.5% to 10% or by changing any infusions to saline solutions
- If hyperglycaemia persists then start an insulin infusion (see hyperglycaemia section page 77)
- Amino acids are more potent stimulators of insulin release in premature infants than glucose, so the glucose tolerance may improve when TPN is started (conversely, lipid can impair glucose tolerance).

Heparin

- Prolongs the life of the catheter, decreases occlusion rates and catheter related sepsis
- 0.5 IU/ml heparin is added to TPN

Electrolytes

- Na, K, Ca, PO₄, Mg are provided
Trace Elements
- Zinc, Selenium, Iodine, Copper, Manganese, Chromium are supplied as 0.6ml/kg of AUSPEN trace elements when on fluids of 150ml/kg/day.
- Additional Zinc and Selenium are given to reach current recommended requirements

Acetate
- Acetate is used to increase the HCO₃ lowered in metabolic acidosis and increased renal losses.
- Very preterm babies usually need between 1 - 4 mmol/kg/day of acetate
- Term babies do not require as much acetate and there is less in the Term bag.
- Consider and individual TPN bag without acetate if the pH >7.50, HCO₃ >37.5mmol/L over a 24 hour period and the ventilation has been manipulated and maximised.

Calories
- Aiming for 85 - 110 kcal/kg/day
- Some babies will have higher requirements if they are growth restricted or have co-existing diseases
- The calories are calculated automatically on the TPN Prescription Sheet
- The dietitian (Helen Little, pager 5102) should be consulted if advice is needed.
- Babies need to be weighed as a minimum every 4 days

TPN Bags

Starter
- To be used by all babies receiving TPN on day 1 and safe for peripheral lines
- Targeted to give at least 2g/kg/day protein when receiving 90ml/kg/day fluids with other infusions
- To be used for 24 hours depending on the electrolyte status of the baby
- Not to be given at rates >120ml/kg/day
- Low sodium content and no potassium content

Standard Sodium - Peripheral
- Used for preterm infants from 24 hours of age
- Peripheral line safe, however if infusing peripherally this needs to be reviewed daily to balance the risks of potential peripheral extravasation versus insertion of a central line

Standard Sodium – Central
- Most commonly used bag for preterm infants
- Contains higher amounts of Ca and PO₄ compared to the standard peripheral bag, needs a central line

High Sodium
- For hyponaatraemic babies
- Must be infused through a central line due to the calcium content
- Often needed by day 5-7 in extremely preterm infants

Term
- Solution for term babies from 24 hours of age
- Start with a Starter bag and use a Term bags from day 2 or 3.
- Less protein and acetate compared to other bags
- If there are no Term bags available at 24 hours use a Standard Sodium Bag until Term bag is available
- Must be infused through a central line due to the calcium content and 12.5% dextrose

Low Glucose
- This bag is an alternative to use in a baby with hyperglycaemia, however, the standard practice for this unit is to treat hyperglycaemia firstly with insulin and only then reduce the dextrose content
- It is a 300mL bag which should run for 48 hours as babies needing this bag are usually <1000g
- Needs to be ordered from Pharmacy as it is not a stock item due to the infrequency of its use
- Contains higher acetate as these babies often have a metabolic acidosis
- Contains Ca and PO₄ levels at the maximum levels and so there is no need to order an individualised bag for increased Ca and Po₄ components as it won't be achievable
<table>
<thead>
<tr>
<th></th>
<th>Starter 300ml</th>
<th>Standard Peripheral 650ml</th>
<th>Standard Central 650ml</th>
<th>High 650ml</th>
<th>Term 1000ml</th>
<th>Low Glucose 300ml</th>
</tr>
</thead>
<tbody>
<tr>
<td>P</td>
<td>P</td>
<td>C</td>
<td>C</td>
<td>C</td>
<td>C</td>
<td>C</td>
</tr>
<tr>
<td>Glucose</td>
<td>10%</td>
<td>10%</td>
<td>10%</td>
<td>10%</td>
<td>12.5%</td>
<td>7.5%</td>
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<tr>
<td>Protein</td>
<td>2.9</td>
<td>4.7</td>
<td>4.7</td>
<td>4.7</td>
<td>3</td>
<td>4.7</td>
</tr>
<tr>
<td>Na mmol/kg/d</td>
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<td>4</td>
<td>8</td>
<td>3</td>
<td>4</td>
</tr>
<tr>
<td>K</td>
<td>0</td>
<td>3</td>
<td>3</td>
<td>3</td>
<td>3</td>
<td>3</td>
</tr>
<tr>
<td>Ca</td>
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<td>0.9</td>
<td>1.6</td>
<td>1.6</td>
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<td>PO4</td>
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<td>Mg</td>
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<td>0.3</td>
<td>0.3</td>
<td>0.3</td>
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</tr>
<tr>
<td>Chloride</td>
<td>0.5</td>
<td>3</td>
<td>2</td>
<td>5.3</td>
<td>4.6</td>
<td>0.9</td>
</tr>
<tr>
<td>Acetate</td>
<td>0.6</td>
<td>3.6</td>
<td>3.3</td>
<td>5.3</td>
<td>1.3</td>
<td>4.4</td>
</tr>
</tbody>
</table>

**Prescribing TPN**
- The protein, lipid and electrolyte requirements are discussed on ward rounds.
- Call Pharmacy on 80831 on Mon and Thurs to update them on the current number of patients on TPN and potential new admissions (especially multiple births or surgical babies).
- In those unstable babies who have additional requirements an individual TPN bag can be requested.
- TPN should be started as soon as there is iv access.
- The TPN bags can be hung for 48 hours.
- If iv access is changed the only time when a new bag is needed for sterility reasons is when access changes from a peripheral to a central site.
- Increase the strength of other infusions to provide more volume per day to be provided as TPN.
- Manipulations can be made in the mmol/kg/day of electrolytes provided by altering the volume of TPN given per day and altering infusion solutions.

**Prescription Instructions**
- Open the template in the TPN folder on the G:drive.
- Save the template into a new folder for the baby.
- Enter data into the white boxes only, the yellow boxes values will be calculated for you.
- Entering the ml/day of infusions that are in 0.9% saline and 0.45% saline is to calculate the extra Na these infusions will provide. **Do not include insulin** as this is over and above fluid requirements.
- Entering the ml/day of dextrose infusions is to calculate the extra kcal/kg/day that this will provide.
- If a sideline of Na is needed (see hyponatraemia section) as the Na is low but not low enough to warrant a High Na bag or this is unavailable - add the additional mmol/kg/day from the Na sideline in the appropriate box to calculate the total Na to be received in the day.
- Chart the fluids on the Level 3 fluid sheet.
- Correct charting includes writing the type of TPN bag being prescribed and the rate in ml/hr.
- If the baby is on enteral feeds chart TPN (overlapping with feeds) and the total rate in ml/hr of nutrition whether it be TPN or enteral feeds.

**Charting TPN again on the same patient**
- Open up the patients file – G:drive, TPN folder.
- Open up the form that was used the day before.
- Use the same form for the week, print the summary for the week (Page 1) on a Sunday and file in notes.
- When the baby stops TPN - Print page 1 only of the form and file it in the notes.
- When the baby needs TPN the next week open up a new template and save with the new date.
**Individual Bag Prescription Instructions**

- The need for an individual TPN bag will be decided by the consultant.
- Use the Individual TPN prescription sheet.
- It is similar to fill in as the premix bag but has less calculations done for you as you will be deciding the dextrose, electrolytes and protein that you will need.
- Print and fax the TPN worksheet to Pharmacy **ASAP and before 10am** indicating that this is individually prescribed TPN and **call Pharmacy on 80831**
- If stable try to request on Thurs – Thurs/Fri individual bags and on Fri – Sat/Sun bags
- The bag must be changed every 24 hours and it is specific to that day only.

**Prescribing Lipid**

- Standardised Lipid Syringes will be supplied by Baxter with lipid and vitamins
- SMOFlipid contains 30% soya oil, 30% MCT coconut oil, 25% olive oil, 15% fish oil
- All syringes contain the same contents (see below) and are light protected with amber syringes and tubing as peroxidation of fatty acids occurs if exposed to light
- Run through a lipid filter for up to 48 hours if volume allows
- Use the baby’s exact weight when calculating rates
- Stable for 7 days after being compounded
- Infuse for 48 hours using the lipid filter
- Stepwise increase from 1 to 3 g/kg/day and up to 3.5 or 4g/kg/day on dieticians advice
- Reduce lipid or keep at 2g/kg/day when on >80ml/kg/day enteral feeds
- Stop lipid when on >100ml/kg/day enteral feeds
- These feed volumes should be entered into the TPN worksheet as a reminder
- If the lipid rate is being kept at 1gm/kg/day then, the TPN volume should not exceed 100ml/kg/day as above this the ratio of nitrogen to non-nitrogen balance becomes unbalanced.
- Prescribe the lipid on the level 3 chart indicating g/kg/day lipid, rate in ml/hr and guardrail in mg/kg/hr

<table>
<thead>
<tr>
<th>Total Volume 50ml</th>
</tr>
</thead>
<tbody>
<tr>
<td>SMOFlipid 20%</td>
</tr>
<tr>
<td>Baxter Paed Multivitamin</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Lipid</th>
<th>Volume to be prescribed</th>
<th>Guardrail</th>
</tr>
</thead>
<tbody>
<tr>
<td>1g/kg/day</td>
<td>*0.23 ml/kg/hr</td>
<td>42 mg/kg/hr</td>
</tr>
<tr>
<td>2g/kg/day</td>
<td>*0.46 ml/kg/hr</td>
<td>83 mg/kg/hr</td>
</tr>
<tr>
<td>3g/kg/day</td>
<td>*0.69 ml/kg/hr</td>
<td>125 mg/kg/hr</td>
</tr>
<tr>
<td>3.5g/kg/day</td>
<td>*0.81 ml/kg/day</td>
<td>146 mg/kg/hr</td>
</tr>
<tr>
<td>4g/kg/day</td>
<td>*0.92 ml/kg/day</td>
<td>167 mg/kg/hr</td>
</tr>
</tbody>
</table>

**Indications to slow stepwise progression of lipid:**

- Significant jaundice - free fatty acids can displace bilirubin from binding sites on albumin
- Thrombocytopenia - hyperlipidemia can impair platelet function
- Severe PPHN keep ≤ 2g/kg/day
- Sepsis - insufficient evidence to support decreasing lipids in the presence of sepsis
Laboratory Monitoring of Babies on TPN

Extreme Preterms <26 weeks
- These babies have more electrolyte instability particularly with Na and K.
- Na can rise rapidly from increased insensible losses.
- K can rise and peak around 24 hours because of hypothermia, traumatic delivery, haemodynamic instability and metabolic acidosis.
- Electrolytes should be taken at birth and monitored 12 hourly for the first 48 hours or until stability is achieved (with NEON and gas electrolytes)
- Commence the TPN at 90ml/kg/day as the Na will rapidly rise due to increased insensible losses.
- The need for High Na TPN is predictable and is needed from day 5 to 7 due to increased renal loss

Stable Babies
- Blood gas and electrolytes once a day while they are being started on TPN.
- When the baby is clinically stable it is appropriate to space out sampling to alternate days or twice weekly.
- All babies on TPN should have liver function tests and a conjugated bilirubin checked if there is jaundice or concern regarding hepatic dysfunction.

Prolonged TPN >1month
- Referral to Prof Andrew Day Gastroenterologist
- Refer to National Intestinal Failure Service if remains on TPN at 31 days if born ≥34 weeks or on TPN at 42 days if <34 weeks at birth (liaise with Helen Little, dietician)

Stepwise TPN and Lipid Progression

Day 1  (Fluids 90ml/kg/day and Lipid 1g/kg/day)
- TPN Starter Bag – to run for 24 hours
- Lipid 1g/kg/day – to run for 24 hours

Day 2-3  (Fluids usually 90 or 120ml/kg/day and Lipid 2 or 3 g/kg/day)
- Standard Sodium TPN Change to Standard Sodium bag after 24 hours if preterm
  Bag volume will last 48 hours
- Term TPN Change to Term bag after 24 hours if term
  Bag volume will last 48 hours
- Lipid Change lipid syringe at 24 hours as TPN also needs to be changed
  If >1.5kg the lipid syringe will need to be changed after 24 hours
  If <1.5kg the lipid syringe volume will last for 48 hours

Day 3-5  (Fluids usually 120 or 150 ml/kg/day and lipid 3g/kg/day)
- Standard or High Na If >2.5kg TPN bag will need to be changed at 24 hours
- Term TPN If >3.5kg TPN bag will need to be changed at 24 hours
- Lipid If >1.5kg lipid syringe will need to be changed at 24 hours

- If the TPN bag needs to be changed remember to consider if this is a good time to change the lipid syringe
- TPN bags need to be changed at 24 or 48 hours depending on the volume used in a day and not just when the volume runs out. Try to time any TPN bag and lipid syringe changes to the same times if possible
- Remember line changes are every 4 days and factor that in to any bag/syringe changes

The Milk Bank

The Milk Bank provides pasteurised donated milk (PDM) for the NICU and the Maternity Ward when supplies allow according to the prioritisation tool below. Anthea Franks and Schol O’Bery are the Milk Bank Managers and they are supported by the executive team of: Maggie Meeks / Kristen Hougland / Bernard Hutchinson / Hazel McGregor / Nicky Clark / Graeme Webb.
The main aim of the Milk Bank is to have pasteurised milk available to NICU patients to:

- Support women as they establish their milk supply following the birth of their sick or preterm baby
- Reduce formula exposure in infants at risk of Necrotising Enterocolitis (NEC)

Parent information leaflets and consent forms are held in reception. Information is also available on the INTRANET http://cdhbintranet/WomensandChildrensHealth/ChildHealthClinicalResources/SitePages/Human%20Milk%20Bank.aspx

Milk Bank RECIPIENTS

Identifying Who Is Eligible

The diagram below shows the Milk Bank prioritisation system. The aim of this system is to ensure that when PDM is made available to a specific baby, there will be enough supplies for at least 1 week. This eligibility criteria will be updated on a weekly basis depending on PDM stock supplies.

When the prioritisation colour is GREEN (indicating ample supply)

- The priority remains with those at risk of NEC and the moderately preterm or late preterm infants where a mother is aiming to breastfeed
- Infants > 34 completed weeks at birth will also be eligible for the first week of life as their mother establishes breast milk supply
- PDM should not be offered routinely to term infants admitted from the postnatal ward who have already had formula unless they demonstrate feeding intolerance.
- PDM can be offered to infants < 34 completed weeks at birth whose mothers intend to formula feed

There are some other important points to consider when a baby has been identified as eligible for PDM:

- ALL babies should receive their mothers colostrum where possible prior to receiving PDM
- The mothers own expressed breast milk remains the first choice which is why the mothers should be encouraged and educated regarding expressions techniques and frequency of expression
- In babies < 30 weeks receiving TPN, the aim should be to delay commencement of PDM for 5 days (7 days for <28 weeks) to positively reinforce the message that the baby’s mother’s milk is first choice.
Consent for Milk Bank Recipients

- The recipient consent form needs to be signed by one of the parents/caregivers of the baby.
- Consent can be obtained by all neonatal staff that have been trained in the consenting process. A short video illustrating the process of consent can be found on the intranet under Human Milk Bank.
- The commonest question likely to be asked by the parents of a potential recipient is:
  - “What is the risk to my baby of receiving pasteurised human milk?”
    - The information leaflet addresses this question.
    - The donors are screened and tested and the Milk Bank only accept donors that meet set criteria.
    - The milk is tested before and after pasteurisation and discarded if there is evidence of infection.

Prescribing for Milk Bank Recipients

- Once the consent form has been signed, pasteurised milk can be prescribed on the nutritional additives sheet (Ref9907).
- Once PDM is commenced it is guaranteed for 7 days only but supply can be extended if stocks allow and circumstances suggest that a few extra days may allow the mother to reach the required supply, eg multiple pregnancy, lactation support strategy in place, mummy time, advice of Infant Feeding Specialist Hazel McGregor or Lactation Consultant Megan Penrose.

Administering Donor Milk

The Human Milk Bank Policy has a detailed dispensing procedure which should be followed.

Overlapping to Formula from PDM

For babies birth weight < 1250 g and/or has significant feeding tolerance issues or high risk of NEC:

Overlap over 4 days unless specified otherwise by SMO

- Day 1 start with 25% formula
- Day 2 advance to 50 % formula
- Day 3 advance to 75 % formula
- Day 4 full formula feeds
- Continue Probiotics until transition is completed

For babies with no concerns

- Day 1 If still PDM available alternate feeds with formula/any EDM available
- Day 2 full formula feeds supplemented with any EBM available

Milk Bank DONORS

Recruiting Donors

- Those that donate to the Human Milk Bank are called ‘Donors’ and are recruited from mothers who have a surplus supply of expressed breast milk for present of future needs. A Donor can be mothers from the NICU or the Community.

Enquiries About Donating

Please direct all internal enquiries to the Milk Bank or to the NICU feeding, dietetic or lactation staff. All external enquiries should be directed to the Human Milk Bank Website ([http://cdhbintranet/WomensandChildrensHealth/ChildHealthClinicalResources/SitePages/Human%20Milk%20Bank.aspx](http://cdhbintranet/WomensandChildrensHealth/ChildHealthClinicalResources/SitePages/Human%20Milk%20Bank.aspx)) or the Milk Bank Manager can be emailed at: MilkBankNICU@cdhb.health.nz

Consenting and screening donors

- All donors must meet the criteria set out on the Milk Bank Health questionnaire and should only be formally consented if this criteria is met.
- Consent of donors will primarily be completed by the Milk Bank Manager, Infant Feeding Specialist or Lactation Consultant.
- Once a mother has consented she needs to have screening bloods taken and staff should be aware that the results may take up to a week.
Community Donated Milk Pick Up Service

- A weekly community service provided by Nurse Maude volunteers picks up donations from registered donors’ homes and delivers them to the Milk Bank.

Resources

The following people are on the Milk Bank Executive Committee and should be able to be used as resources:
- Anthea Franks (Milk Bank Manager), Schol O’Bery (Milk Bank Manager), Nicky Clark (Paediatric Dietician), Bernard Hutchinson (ACNM), Hazel McGregor (Infant Feeding Specialist), Megan Penrose (Lactation Consultant), Kristen Hougland (Neonatal Consultant), Maggie Meeks (Neonatal Consultant)
NEONATAL SEPSIS AND CONGENITAL INFECTIONS

Thorough handwashing, before and after every contact with every baby is by far the most important method of preventing nosocomial infections.

Maternal Antibiotic Prophylaxis

- CWH adopts a risk based approach to GBS (as opposed to universal antenatal screening).
- Risk factors need to be identified in pregnancy/labour to dictate the course of management.
- 20-30% of women are colonised with GBS.
- GBS neonatal infection occurs early in 80% (<7 days age) and can be a devastating infection if untreated.
- If GBS is present and treated in the pregnancy do not consider that the GBS has been eradicated.
- ≥ 4 hours of intravenous antepartum antibiotics are required to provide cover to the baby.

Risk Factors for GBS

- Previous baby with GBS (including late onset)
- GBS in the maternal urine in the current pregnancy
- GBS colonisation on vaginal swabs in the current pregnancy (with the exception of a negative swab at ≥37wk using the selective broth process)
- Prolonged ROM ≥ 24 hrs (increasing risk after 18 hours)
- Preterm labour <37 weeks (with or without ruptured membranes)

Risk factors for Early Neonatal Sepsis

- prolonged rupture of membranes ≥24hrs (increasing risk after 18 hours)
- maternal illness, pyrexia >38.0 C (but any elevation >37.5 C increases risk), WBC > 15, raised CRP >1
- 0, suspected chorioamnionitis
- pathogens (e.g. GBS, E. coli) present in maternal urine or high vaginal swab
- prematurity < 37 weeks
- fetal distress, tachycardia > 160 bpm or need for resuscitation
- twin gestation

Although none of these risk factors alone has particularly good positive predictive value for sepsis, the more that are present, the lower the threshold should be to investigate and treat the baby for even minor clinical signs. In all sepsis, early diagnosis is vital.

Commencing antibiotics based on good clinical judgement will always be supported.

Risk factors may be an indication for investigation but are not in themselves an indication for antibiotics if the baby is born at term and is clinically well.

The decision process needs to balance the potential for unnecessary treatment of uninfected babies with delaying antibiotic treatment in infected babies.

Initial therapy is often commenced on the basis of clinical suspicion, since life-threatening infection can become established extremely quickly.
Neonatal Sepsis

Early Onset Sepsis (EOS)

- This is most often perinatally acquired, but nosocomial possible.
- ANZNN definition is the first 48 hours
- Group B streptococcus (GBS), E coli K1, Streptococci and Gram negative organisms are common causes
- Listeria monocytogenes can also occur in a sporadic or epidemic pattern.
- The EOS rate in VLBW infants in a study by Ting et al. 2019, was 1 in 56 with risk factors, whereas when no risk factors are present was 1 in 204.

Late Onset Sepsis (LOS)

- Usually nosocomial
- ANZNN define as from 48 hours
- VLBW infants with indwelling catheters, central lines, chest drains, etc are at higher risk
- In premature infants is predominantly caused by Coagulase negative staphylococci although since introducing the line bundle of care our rate is now very low.
- Others include Gram negative organisms (e.g. E coli, Klebsiella sp..) late onset GBS, fungal sepsis (usually Candida parapsilosis or Candida albicans) and Staph. aureus,

Clinical Features / Signs and symptoms of sepsis

- All newborn infants with early respiratory distress (tachypnoea, grunting, increased work of breathing, nasal flare) should be assessed for risk of EOS (GBS, E. Coli, other) and should be investigated.
- Where grunting is intermittent / and maybe described as “singing” they are not for transfer out from CWH OR if in primary maternity unit may need transfer into CWH.
- Use the sepsis calculator and flow chart to assist with blood tests. A Capillary Blood Gas may be helpful if there are no cord lactates.
- Temperature instability – hypothermia and hyperthermia are often due to issues with environmental temperature, but a body temp. of < 36°C or > 37.5°C for greater than 1 hour (if appropriate manoeuvres have been undertaken to correct environmental temperature) is possible sepsis and warrants clinical review and investigations
- Previously healthy baby who becomes too sleepy/lethargic to feed
- Listlessness, lethargy, pallor, mottling and irritability
- New or increased respiratory distress – tachypnoea, increased WOB, grunting, increase in oxygen requirement
- Jaundice if it develops unusually rapidly
- Abdominal distension or bilious vomiting or nasogastric aspirate suggesting ileus
- Apnoea, especially new onset or increased frequency or severity
Management of the **Asymptomatic** Baby at Risk of Sepsis ≥37 weeks

**Clinical Chorioamnionitis**
- Observations at 1hr, 4hrs and 4 hrly for 24 hrs
- Not for transfer to a birthing unit or discharge home until after 24 hrs
- Neonatal review prior to transfer/discharge

**PROM GBS + Intrapartum Ab <4hr**
- Observations at 1 hr, 4 hrs and 4 hrly for 24 hrs
- May transfer to a birthing unit after 6hrs
- Not to discharge home until
- Midwife review prior to transfer

**PROM GBS + Intrapartum Ab ≥ 4hr**
- Observations 1 hr, 4 hrs and 4 hrly for 24 hrs
- May transfer to a birthing unit any time with midwife review prior to transfer
- May transfer home any time with LMC taking over the review of sepsis risk

Management of the **Asymptomatic** Baby at Risk of Sepsis 32-37 Weeks

**Clinical Chorioamnionitis or PROM, GBS + Intrapartum Ab <4hr or ≥ 4hr**
- Observations at 1hr, 4hrs and 4 hrly for 24 hrs
- Not for transfer to a birthing unit or discharge home
- Neonatal review daily and prior to discharge which may be up to 4 days due to prematurity

If symptoms develop the neonatal team needs to be consulted to examine and investigate the baby for sepsis.

The SMO responsible for postnatal / level 5 babies is the Room 5/6/7 consultant unless unavailable and the on service/on call SMO should be contacted.
Management of the Symptomatic Baby at Risk of Sepsis < 32 Weeks

All babies < 32 weeks have a higher risk of sepsis, irrespective of clinical chorioamnionitis signs in the mother, and symptoms in the baby.

Some babies born for maternal PET, APH, maternal reasons by elective Caesarean are at low risk and we are endeavouring to reduce the use of antibiotics. If they are not requiring respiratory support they do not need antibiotics. This is an SMO decision.

Because most are on CPAP and therefore there is uncertainty we aim to stop the antibiotics as soon as possible.

Follow the sepsis flow chart and the SMO’s decision
Management of the Symptomatic Baby at Risk of Sepsis

- Follow Sepsis Flow Chart

- It is recommended to also use the Sepsis Calculator in babies from 34 weeks. [https://neonatalesepsiscalculator.kaiserpermanente.org/](https://neonatalesepsiscalculator.kaiserpermanente.org/) (validated for babies ≥34wks).
- An app is available – EOS Calculator
- There are 3 Clinical Categories for risk stratification – clinical illness, equivocal presentation and well appearing
- Divided into 3 pathways – treat empirically, observe and evaluate, and continue observation.
- Use 0.4/1000 for the incidence of EOS
- You need to know the highest maternal temp in labour (use 37.0 if unknown) GBS status and duration of AN antibiotics
- If in doubt check with SMO
Considerations
- Antibiotics for infants with sepsis can be lifesaving
- Overuse of empirical antibiotics can facilitate antibiotic resistance and prolonged use is associated with adverse outcomes.
- No antibiotic treatment is not the same as no care
- Prolonged antibiotic courses without a positive microbiology should be exception rather than the rule
- Culture media with antimicrobial neutralization properties enhance the reliability of blood cultures when the mother has received intrapartum antibiotics
- Antibiotic use has been linked to disruptions in the microbiome and is postulated to have a critical role in development of significant neonatal morbidities eg CLD, NEC, ROP & PVL, along with asthma, inflammatory bowel disease and childhood obesity

Investigations
There is no test with perfect sensitivity or specificity so the clinical scenario needs to be taken into account along with the blood test findings to decide if sepsis is present

Usual first line investigations are
- FBC and Blood Culture
- CXR
- CRP at 6hrs

FBC
- Total WBC < 5 and neutropenia < 1
- Immature/Total neutrophil ratio* >0.25 on day 1, >0.20 from day 5
  *This is immature neutrophils, (ie. bands + myelocytes + metamyelocytes) divided by the total of immature neutrophils plus the mature neutrophils.
- Toxic granulation, vacuolisation or Dohle bodies present on the film.
- Thrombocytopenia < 100 – think about Candida

CRP
- An acute phase reactant synthesised within 6-8 hours in response to tissue injury
- Non-infectious processes can also elevate the CRP ie: PROM, perinatal asphyxia, IVH, pneumothoraces, meconium, infarction, trauma, immunisation
- Levels peak at 24-48 hours
- A normal CRP at the start of an illness or at birth lacks the sensitivity to rule out sepsis but if taken at >6hrs the sensitivity improves to >90%
- A level of <10mg/L is considered normal and has a negative predictive value of 99% for infection

Blood Culture
- 1ml of blood is required for an adequate blood culture - smaller volumes may miss bacteraemia
- A negative blood culture result can be due to lack of infection, inadequate sample size or intrapartum maternal antibiotics

CXR
- AP & Lateral views for first X-ray.
- If concern re pneumothorax a AP shoot through will verify an anterior pneumothorax

Urine
- A urine should be taken in the evaluation of early onset sepsis for GBS antigen which indicates systemic GBS infection (not a GBS urine infection)
- Microscopy and culture is not required as the likelihood of a UTI is extremely low (although for late onset sepsis a UTI should be considered in the differential)
A bag urine may be sufficient but if there has been known chorioamnionitis or PROM then the baby’s skin may be colonised so a bladder puncture or catheter sample will be more appropriate to exclude a positive result from skin contamination.

If a bag urine returns a positive result for GBS antigen it should be discussed with the consultant as to whether a bladder puncture is required.

If antibiotics have been stopped after 24-48hrs and bag urine collection has been unsuccessful then it can be omitted.

**Gastric Aspirate and Surface Swabs**

- These have limited value and are not required routinely as the result only indicates colonisation.
- Gastric aspirates should be done for admissions of extremely preterm infants (<28wks) with a high index of suspicion of infection (eg chorioamnionitis).

**Ureaplasma**

- There is a significant association between Ureaplasma respiratory tract colonisation and developing bronchopulmonary dysplasia.
- It is unknown whether treatment after birth can prevent infection-mediated lung injury.
- Colonisation rates vary from 20-45% in infants <28 weeks.
- Rates of colonisation are higher the lower the gestation and with increasing length of preterm prolonged rupture of membranes and preterm labour. However, colonisation is also seen after delivery for maternal indications or for fetal factors without PPROM or labour.
- It is most likely that an inflammatory response to the colonisation starts in-utero but swabs for Ureaplasma are not a routine antenatal investigation.
- All babies <28 weeks should have a test sent for Ureaplasma colonisation on day 1.
- Babies 28-29wks should have a test sent on day 1 if there is PPROM or chorioamnionitis.
- Send an ETT aspirate if intubated for Ureaplasma.
- If not intubated then call the lab to be sent a COPAN FLOQswab and a vial of Mycoplasma transport medium to take a nasopharyngeal swab.
- Treatment is a 3 day course of oral azithromycin. This has been shown to eradicate colonisation whereas erythromycin is not as successful and thus we have changed to azithromycin (June 2020).
- Consider empiric treatment whilst awaiting the swab result if there is definite chorioamnionitis or PPROM with a high neutrophil count.

**Lumbar Puncture**

- This should be considered in a baby with a positive blood culture and in those babies with a negative blood culture but have significant blood changes or clinical signs that make meningitis a possibility.
- Should be discussed with the Consultant.
- This is never an urgent investigation (generally done in daytime hours) and can be delayed if the baby would not tolerate the procedure ie: unstable, ventilated, coagulopathy.
- Meningitis antibiotic doses are indicated until LP undertaken.
- Consider Acyclovir if delay in LP and history of herpes simplex exposure.

**Neonatal Antibiotics**

- The first choice antibiotics for suspected or proven sepsis presenting at birth or within 48 hours and admitted to NICU are:
  - Amoxycillin - 50mg/kg/dose, Q12H, IV push; High dose - 100mg/kg/dose if suspected meningitis or severe sepsis.
  - Gentamicin (IV infusion based on locally devised extended interval dosing).
- The first choice antibiotics for suspected or proven sepsis presenting at birth or within 48 hours and remaining on the postnatal ward with no requirement for NICU admission are:
  - Amoxycillin (50 mg/kg/dose, Q12H, IV push).
  - Cefotaxime (50mg/kg/dose Q12H, IV push).
  - (Gentamicin is currently not given on the postnatal ward as it is an infusion with levels required. However, discuss each case at an appropriate time with the SMO as there may occasionally be clinical factors that dictate gentamicin to be more appropriate).
• For babies who start their antibiotic course in NICU and are transferred to the ward:
  - Amoxycillin
  - Change Gentamicin to Cefotaxime only if a 5 day course is required - this needs to be charted to start at the time that the next gentamicin dose was due (60 hours)
  - These babies are reviewed daily on ward. Discuss with SMO covering room 5/6/7 is preferred contact.
• The preference is for babies on the postnatal ward to have their IV line sited on the postnatal ward.
  - Consider using the Maternity Midwife/ACNM or Clinical Support Nurse (NICU) when available to help hold and tape.
  - If it is necessary to bring the baby down to NICU for an IV line then call the NICU ACNM to coordinate where this is best to be done given NICU workloads. The intention then would be for the IV line to be placed and the baby returned to the postnatal ward for antibiotic administration

A baby on IV antibiotics must be reviewed daily. If results are normal, stopping antibiotics at 24-36 hours is good clinical practice.

If antibiotic duration required has been determined to be > 48 hours, but the baby is now well and CRP has normalised < 10, an SMO review can alter the original duration.
• For late sepsis antibiotics used are vancomycin and cefotaxime (this regime has had Paediatric Infectious Diseases approval)
• For proven or suspected necrotising enterocolitis add in metronidazole to the amoxicillin and gentamicin, or vancomycin and cefotaxime that the baby may already be on
• For Staphylococcal skin sepsis or pneumonia treat with flucloxacillin IV or oral depending on the situation and if severe use in conjunction with gentamicin
• Babies with proven or suspected UTI or renal tract anomalies should receive oral cotrimoxazole (use amoxicillin if the baby is jaundiced and change to cotrimoxazole after 5-7 days when the jaundice has settled)
• PHARMAC have stated that some antibiotics require Infectious Diseases approval. These are - imipenem, meropenem, ciprofloxacin, piperacllin, amphotericin and vancomycin
• This information is noted on the drug profile.
• The antibiotic course can be started without delay but then individual patients will need to be discussed (or email) with Tony Walls (or if unavailable Adult Infectious Diseases) during usual work hours to get approval to continue the course of antibiotics or to discuss other options.

For severe sepsis the following are sometimes indicated (discuss with consultant):
  - Immunoglobulin 500 mg/kg.
  - Exchange transfusion - e.g. if evidence of purpura fulminans (remember to "top-up" antibiotics afterwards).
  - G-CSF if the baby is neutropenic and not responding to antibiotics or immunoglobulin.

Gentamicin Levels

• Gentamicin is the first-line Gram negative cover for babies ≥750g in NICU
• Babies <750g do not clear gentamicin well and so cefotaxime is preferred in most situations
• Our NICU has researched gentamicin dosing in neonates for many years and now uses extended dosing interval treatment with gentamicin given 60 hourly (see drug profile)
• Gentamicin monitoring is important to ensure that babies receive adequate doses for bacterial kill as well as adequate clearance of the drug to minimise the risks of toxicity
• A pragmatic approach to this has been taken as most babies are receiving short courses of empirical antibiotics and we are trying to avoid unnecessary gentamicin levels/blood tests being taken
• 75% of babies receiving gentamicin at this NICU have a single dose only and so gentamicin levels are usually not required (see below**)
• Babies receiving gentamicin courses ≥5 days in length or those in certain clinical situations (see below**) require drug monitoring

Standard process for empirical gentamicin given soon after birth
• Baby is admitted to NICU after birth and gentamicin is given
• At this stage we will not know the length of the antibiotic course
• The blood taken for the CRP (0.6mL, needs a full tube) at around 6 hrs of age can be used to retrospectively run a gentamicin level when a decision is made to continue for 5 or more days and will act as the first level (this decision is usually made around 24 hrs of age)
• Call the Biochem lab on 80376 and inform them on which blood test to analyse the gentamicin level on (if not specified they default to the most recent blood received in the lab)
• Also inform the lab of the gentamicin dose, time of administration and dose intervals so that can be added to the lab report
• If there has been no blood sample taken after administration of gentamicin when the decision is made for 5 or more days of antibiotics then take a blood sample for a first gentamicin level immediately
• A second level will also need to be taken at 24-36 hours (d/w Pharmacist if unsure when to take it)
• Pharmacist will use the two levels to advise on the dose and timing of the next gentamicin dose and if further gentamicin levels are required

**Exceptional situations where gentamicin levels are required on the First Dose**
The process above works for the majority of babies but there are exceptions where close gentamicin monitoring is needed at the initiation of gentamicin:
• At birth a decision is made for at least 5 days of antibiotics prior to the Ab course starting
• Gentamicin given after 1 week of life – clearance is much higher so the dose may need to be bigger and more frequent. In this instance we are usually treating a true sepsis situation rather than empirical antibiotics after birth
• Renal impairment (congenital renal anomaly, HIE, oliguria) – at risk of poor clearance
• Hydrops, severe oedema – true body weight and volume of distribution will be affected
• Concomitant use of other nephrotoxic drugs – indomethacin, frusemide, vancomycin
• Suspected or confirmed Gram negative sepsis – need to optimise gentamicin dosing to treat infection
• Concern about clinical response to antibiotics

**Intramuscular Antibiotics**
• Ideally antibiotics are given iv however there will be situations when an iv line cannot be sited and the clinical situation will need to be discussed with the consultant
• The usual antibiotics that can be given im are amoxicillin and cefotaxime and these can be drawn up with 1% lignocaine to help with the pain after injection
• Due to the potential four-fold error in drawing up the more concentrated gentamicin (80mg/2ml) for im injection versus our usual 10mg/ml concentration a decision has been made not to give gentamicin im

**First Dose of Antibiotic**
• Baby with **signs of sepsis** and unable to site a peripheral iv line
  - Insert a UVC
• Baby with **risk factors** for sepsis but is **well** and unable to site a peripheral line
  - D/W SMO to see if a UVC is felt to be necessary to give antibiotics, or
  - D/W SMO to see if antibiotics are required or if taking FBC, CRP, blood culture and observation are appropriate, or,
  - Give IM cefotaxime 250mg/ml made up with 1% lignocaine as the sole antibiotic with Gram negative and GBS cover (do not give amoxicillin as well to avoid the baby receiving 2 im injections) and review the route of administration prior to the next dose

**Subsequent Dose of Antibiotic**
• Baby with **signs of sepsis**, peripheral iv has tissued after receiving at least 1dose of amoxicillin and gentamicin
  - Insert a UVC, or,
  - Give IM amoxicillin 250mg/ml made up with 1% lignocaine as the sole antibiotic, as initial gentamicin dose will be providing coverage for 60 hours and review the amoxicillin route of administration prior to the next dose
• Baby with **risk factors** for sepsis but is **well** and peripheral iv has tissued after receiving at least 1 dose of amoxicillin and gentamicin
  - D/W SMO to see if antibiotics are still required or if taking FBC, CRP, blood culture and observation are appropriate, or,
  - Give IM amoxicillin 250mg/ml made up with 1% lignocaine as the sole antibiotic, as initial gentamicin dose will be providing coverage for 60 hours and review the amoxicillin route of administration prior to the next dose
Culture of Lines and Tips

- There is usually a low yield for sending tips to microbiology for culture and it does not often change our management of the patient.
- In certain circumstances the clinical care may be helped by knowing what the baby is colonised with. For example: sending a longline tip would be appropriate if there were concerns for a central line infection.
- A consensus decision has been made that: UAC, UVC, LL and ETT tips – not to be sent for culture routinely but may be requested on an individual basis by the SMO.

Sticky Eyes

- The commonest cause of a sticky eye is a blocked tear duct.
- If the eyes are sticky and the conjunctiva are red and swollen, send an urgent gram stain and appropriate swab for culture to exclude gonococcal ophthalmitis (call microbiology).
- A chlamydia swab should also be taken and sent for immunofluorescence.
- Chlamydia swabs (special pink swabs) are kept in the fridge in Level 3, or may need to be requested from the laboratory. A vigorous scraping of the conjunctiva should be undertaken, prior to the baby being commenced on treatment. If the immunofluorescence is positive, commence systemic erythromycin.
- A routine bacterial culture should also be sent. Routine treatment for purulent eye discharge is chloramphenicol eye drops, one drop each eye four times a day for one week. Fusidic acid is an alternative.

Staphylococcal Infections

*Staphylococcus aureus* skin colonization

- Some babies may be colonized by *Staph. aureus* in the first 24 hours, but, only 30% of infants in one study were found to be colonized by bacteria at 6 days of age.
- Staph. colonization does not always correlate directly with incidence of infection presumably because of variable virulence of the organisms and host resistance.
- Male infants appear to have higher infection rates of bacterial infection compared to females.
- The sites most commonly colonised by *Staph. aureus* are the umbilicus, skin flexures and the nares.

*Staphylococcus aureus* superficial infections

- **Omphalitis** - erythema and/or induration with purulent discharge from the umbilical stump, due to gram+ve / gram-ve/ anaerobic organisms.
- **Paronychia** - inflammation of the nail bed.
- **Pustulosis** - localised collections of vesicopustules on an erythematous base in an otherwise asymptomatic baby. Gram stain will show Gram-positive cocci and abundant neutrophils, and culture will confirm *Staph. aureus*.

Treatment for *Staphylococcus. aureus* skin infections

- Any systemic sign of infection take blood cultures and give systemic iv antibiotics.
- Any Staphylococcal infection in a preterm infant < 35 weeks:
  - blood cultures and systemic iv antibiotics: iv for minimum 24-48 hours, after which oral antibiotics to complete a 5 day course if the baby remains well.
- Isolated Staphylococcal skin pustules in a well baby > 35/40:
  - consider chlorhexidine body wash and repeat at 24 hours if improved
  - start oral flucloxacillin if not improved within 24 hours and treat for 5 days.
- Isolated Staphylococcal superficial omphalitis:
  - oral flucloxacillin for 5 days.
  - consider adding topical treatment with alcohol wipes as well.
- Open, purulent sites may need contact precautions (gloves and aprons) in addition to universal precautions.
Chlorhexidine wash protocol

- Wet the baby’s body, face, eyes and ears with warm water.
- Spread 1% chlorhexidine white obstetric cream over the whole body except the eyes. All creases, the perianal area, periumbilical area, axillae and the neck folds should be treated.
- Massage the chlorhexidine cream gently into the scalp.
- Leave the cream in contact with the skin for 60 seconds or more.
- Wash all of the cream off gently or sponge off with warm water.
- If chlorhexidine cream accidentally gets into the eyes, gently rinse with a liberal amount of warm water only.
- An in vitro study showed that an increasing duration of exposure of *Staphylococcus aureus* to chlorhexidine 0.5% solution from 15 to 30 and 60 seconds reduced the colony count by 37%, 77% and 93% respectively.

Staphylococcal Scalded Skin Syndrome

This condition is characterised by red blistering skin which is caused by the release of two exotoxins (epidermic toxins A and B) from toxigenic strains of *Staphylococcus aureus*. Neonates are particularly at risk due to the lack of specific immunity to the toxins and an immature renal clearance system. Outbreaks in Neonatal units may be due to a staphylococcal carrier in the staff. When a baby is thought to have staphylococcal scalded skin syndrome the management will include:

- Specimen (skin swab) to be sent to the Institute of Environmental Science and Research (ESR) along with a detailed history to determine whether the *Staphylococcus aureus* is a toxigenic strain.
- Place the infant into contact isolation until the results are available (1-2 weeks)
- When the infant is being bathed they should be washed with 1% chlorhexidine obstetric cream (as above) until discharge. This is aimed at suppressing the organism on the affected infant and reduces the likelihood of transmission to other infants in the unit.
- Consider contact tracing of staff
- Strict hand hygiene is the key to prevention and further transmission.

MRSA

- MRSA stands for Methicillin Resistant *Staphylococcus aureus*
- The term is used to describe a number of strains of the bacterium *Staphylococcus aureus* which have developed resistance to antibiotics commonly used to treat staphylococcal infections therefore limiting treatment options.
- MRSA is an opportunistic bacterium which may colonise and grow readily on the skin and mucous membranes of a person, without harm to that person.
- It is commonly isolated from warm, moist body sites such as the nose, groin and perineum.
- MRSA colonisation can lead to infection such as infected skin lesions.
- The main focus of this guideline is to prevent MRSA infection in our patients and to limit spread between vulnerable patients in NICU.
- If MRSA is isolated inform the SMO on service, the Clinical Nurse Specialist Infection Prevention & Control (CNS-IPC) and Dr Tony Walls, Paediatric Infectious Diseases
- Parent information sheets are available to be given to the family and are on the Neonatal Intranet site
- The discharge letter must document if the baby tested positive for MRSA

MRSA – Screening Swabs on Baby

- A blue bacterial swab is used to sample the following sites:
  - Nasal Swab (one swab for both nostrils)
  - Groin Swab (one swab for both sides)
  - Perineum Swab (natal cleft)
  - Additional sites- urine if catheterised, wound sites or device insertion sites i.e. IV lines, drains
**MRSA – Positive Swab on Baby**

This can be either due to *colonisation* being detected incidentally with a positive swab for MRSA in a well baby, or, *infection* detected with a positive swab for MRSA from a wound/infected site

- No further swabs are required
- Isolate the baby in Contact Precautions for the remainder of the admission
  - All staff to wear gown/apron and gloves when tending to the baby
  - Practice 5 Moments for Hand Hygiene
  - Relatives do not need to wear gowns or gloves
- Decolonise the baby (see below) to suppress the MRSA to reduce the risk of cross infection in the NICU
- IV antibiotics (vancomycin) as well if MRSA is thought to be the causative organism for an active infection
- Repeat swabs after the decolonisation regime are not required
- The most likely source of the MRSA is the family and there is no need to screen the family to see if they are colonised with MRSA as the management of the baby will not change
- A risk assessment should be done, however, by asking the family members if any of them have wounds or skin infections that may need treatment by their GP or Obstetrics if still an inpatient

**Mr. Breakout**

- If there are multiple cases testing positive for MRSA, Tony Walls and the CNS-IPC would be consulted and give advice on contact tracing and screening

---

**MRSA – Positive Swab on Mother**

This can be either due to *colonisation* in the mother (within the past 2 years) or from a swab from an active *infection* site such as a wound

- No further swab are required on the mother
- The baby does not need any screening swabs to be taken
- Isolate the baby in Contact Precautions for the remainder of the admission
  - All staff to wear gown/apron and gloves when tending to the baby
  - Practice 5 Moments for Hand Hygiene
  - Relatives do not need to wear gowns or gloves

Decolonisation/Suppression Treatment:

- Mupirocin 2% nasal ointment (Bactroban) – applied to the skin covered area of the anterior nares just inside each nostril using a clean cotton bud for each application **three times a day for 7 days**
- Chlorhexidine 1% cream applied to all skin areas, including the head and then washed off with a bath or warm flannels, **once a day for 7 days**
- If the MRSA strain demonstrates resistance to Mupirocin 2% (Bactroban) alternative treatment options are available e.g. Bacitracin.
Congenital Infections

The presentation of these diseases is rarely specific and maternal infections antenatally are often asymptomatic or only mildly symptomatic. Therefore consider congenital infections in infants who have:

- IUGR, Purpura, jaundice, chronic rash, anaemia, seizures, cerebral calcification, hepatosplenomegaly, chorioretinitis, microphthalmia, pneumonitis, cataract

The investigation and treatment of these diseases is complicated and should be done in consultation with the Neonatal consultant and the Paediatric infectious disease consultant Tony Walls. At discharge discuss the follow up needs of infants with congenital infection with the consultant. Most will need developmental follow up and many will need hearing and ophthalmological assessments.

<table>
<thead>
<tr>
<th>HERPES SIMPLEX (updated from 2013 National Guidelines)</th>
</tr>
</thead>
<tbody>
<tr>
<td>(CDHB Labs no longer processes surface swab cultures and only uses PCR)</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Symptoms and Risk</th>
</tr>
</thead>
<tbody>
<tr>
<td>Only 30% of mothers of infected infants have a history of symptomatic genital herpes so need to have an index of suspicion</td>
</tr>
<tr>
<td>85% of disease is contracted during labour with only 10% being contracted postpartum</td>
</tr>
<tr>
<td>The risk of HSV infection in an infant born vaginally to a mother with a first episode of primary genital infection is 57% and so caesarean section is indicated</td>
</tr>
<tr>
<td>The risk from recurrent genital HSV i is 3% as there is some protection from maternal Ab’s</td>
</tr>
<tr>
<td>There are no absolute guidelines on how to deliver a mother with an active recurrent lesion, however, caesarean section should be offered but will not eradicate the risk of HSV transmission and is not an absolute indication (see flow charts).</td>
</tr>
<tr>
<td>Scalp electrodes and instrumentation must be avoided if there is suspicion of active HSV There may be a history of contact with herpes simplex but most symptoms are non-specific, vesicular lesions (in 40% only), pustules, fever, seizures, encephalopathy, may present with liver disease</td>
</tr>
<tr>
<td>Intrauterine disease – IUGR, chorioretinitis, skin scarring, hydranencephaly</td>
</tr>
<tr>
<td>Skin/Eye/Mouth – in 45%, good prognosis but readily disseminates if not treated</td>
</tr>
<tr>
<td>Disseminated disease – in 25%, with mortality of 30% even if treated</td>
</tr>
<tr>
<td>CNS disease – in 30%, presents with encephalitis from day 5-21</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Investigation for Mother</th>
</tr>
</thead>
<tbody>
<tr>
<td>Type specific serology testing but not often at the time as results are not immediate</td>
</tr>
<tr>
<td>Vesicle fluid sent for HSV/VZV PCR</td>
</tr>
<tr>
<td>Acyclovir from 36 weeks may decrease the risk of recurrent lesions at term (if prior outbreak earlier in pregnancy) and decrease the need for a LSCS if there are no lesions present at the time of birth. Aciclovir in this setting does not eliminate viral shedding though</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Investigation for Infant if: Suspected or Confirmed Primary HSV Infection at birth or within 6 wks of birth</th>
</tr>
</thead>
<tbody>
<tr>
<td>Delivered by LSCS and membranes ruptured for less than 4 hours</td>
</tr>
<tr>
<td>- Surface swabs of oropharynx, conjunctiva, rectum for PCR 24-48hrs after birth</td>
</tr>
<tr>
<td>- If swabs are negative – no further treatment required</td>
</tr>
<tr>
<td>- If baby becomes symptomatic with CNS signs, disseminated disease or skin lesions at any time whilst the initial surface swab results are awaited the admit and investigate as below</td>
</tr>
<tr>
<td>Symptomatic or positive surface swabs:</td>
</tr>
<tr>
<td>- Take Blood (PCR and culture), CSF (PCR and culture) prior to starting iv aciclovir</td>
</tr>
<tr>
<td>- If there are any skins lesion scrape the base of the lesion and send for PCR</td>
</tr>
<tr>
<td>- Treat for a minimum of 5 days with aciclovir until Blood and CSF (PCR and culture) results remain negative</td>
</tr>
<tr>
<td>- Treat CNS/ disseminated disease for 21 days, treat for 14 days if skin/eye/mouth disease</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Delivered Vaginally or LSCS but membranes ruptured for more than 4 hours</th>
</tr>
</thead>
<tbody>
<tr>
<td>- Surface swabs of oropharynx, conjunctiva, rectum for PCR immediately after birth</td>
</tr>
<tr>
<td>- If there are any skins lesion scrape the base of the lesion and send for PCR</td>
</tr>
<tr>
<td>- Take Blood (PCR and Culture), CSF (PCR and Culture) prior to starting iv aciclovir</td>
</tr>
<tr>
<td>- Treat for a minimum of 5 days with aciclovir until Blood and CSF (PCR and Culture) results remain negative</td>
</tr>
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</tbody>
</table>
**HERPES SIMPLEX** (updated from 2013 National Guidelines)
(CDHB Labs no longer processes surface swab cultures and only uses PCR)

**Investigation for Infant if :**

Recurrent HSV Infection

- Vaginal delivery is appropriate even in the presence of recurrent lesions.
- Often a LSCS is offered, but, it does not eradicate the low risk of transmitting HSV
- Avoid scalp electrodes and instrumentation even if no lesions are present
- If there are lesions present at delivery or a history of recurrent lesions in this pregnancy then take surface swabs of oropharynx, conjunctiva, rectum for **PCR 48hrs after birth and not before 24 hours**
- As the risk of transmission is very low the baby can be discharged from CWH and the LMC can take the swabs and follow-up the PCR results

**Asymptomatic but positive surface swabs:**

- If any of the PCR surface swabs taken after 48 hrs age come back positive, in a well baby, there are no current recommendations of what to do in this low risk population. The clinical history and risk factors need to be reviewed and the baby needs a clinical examination and repeat surface swabs taken. This is often best done with a referral to CAA if they are at home.

**Symptomatic:**

- If baby becomes symptomatic with CNS signs, disseminated disease or skin lesions at any time whilst the initial surface swab results are awaited then admit and investigate with Blood (PCR and culture), CSF (PCR and culture), if there are any skins lesion scrape the base of the lesion and send for PCR. Start on iv aciclovir and treat for a minimum of **5 days** with aciclovir until Blood and CSF (PCR and Culture) results remain negative
- If the baby has confirmed infection then treat CNS / disseminated disease for 21 days and 14 days if skin/eye/mouth disease

**Isolation**

- Contact isolation required, especially if skin lesions present.
- Advise mother about the importance of handwashing if she has active lesions

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**CYTOMEGALOVIRUS (CMV)**

**Symptoms**

Maternal symptoms: asymptomatic and/or viral illness with atypical lymphocytes.

Fetal/Neonatal signs: intracerebral calcifications, microcephaly, hydrocephaly, thrombocytopenia, haemolytic anaemia, ascites, hydrops and IUGR.

- Common. 15% of those born after primary infection of their mother will have sequelae.
- Infection and disability can occur regardless of timing in pregnancy but most severe will be primary infection in the first trimester
- 90% infants are asymptomatic at birth but are at risk of hearing impairment and learning disability

**Investigation for Mother**

- Serology CMV IgG and IgM (if these are positive in the first 20 weeks gestation, the lab will do avidity testing - low avidity means infection < 3 months ago, high avidity means infection > 3 months ago) Repeat serology required in 2 weeks time from first testing, if booking bloods are unavailable.
- Consider PCR on amniotic fluid in antenatal period (won’t confirm that the fetus is infected though).
- Obstetric specialist input required.

**Investigation for Infant**

- All infants need one urine sample for PCR taken after birth. Best transported fresh and chilled.
- The virus can be shed for up to 3 weeks so if diagnosis is delayed urine can still be collected up to 3 weeks of age
- Head ultrasound and ophthalmology review if CMV positive
- Universal hearing screening with aABR and review at 9 months and annually until 6 years if CMV positive
- If the hearing screen team take a buccal swab which is positive for CMV then a urine needs to be taken for CMV PCR to confirm the positive result

**Management**

- Treatment of CMV positive babies is intensive (6 weeks of iv ganciclovir) and not routine care.
- No clear evidence that it will improve outcomes.
- Isolation not required but strict handwashing is important
## TOXOPLASMOSIS

| Symptoms | Maternal Symptoms: sore throat, malaise, fever and lymphadenopathy  
<table>
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<tr>
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</thead>
<tbody>
<tr>
<td></td>
<td>Fetal/Neonatal signs: hydrocephalus, microcephaly, intracerebral calcifications, hepatosplenomegaly, lymphadenopathy, maculopapular rash, jaundice, thrombocytopenia, seizures, chorioretinitis. 85% of infected infants will appear normal at birth.</td>
</tr>
</tbody>
</table>

| Investigation for Mother | • Infection in first trimester is less likely to infect fetus (10%) but more likely to cause harm.  
|                        | • Infection in second/third trimester more likely to infect fetus (30-50%) but with milder effects  
|                        | • Toxoplasma IgG and IgM serology and lab will do IgG avidity testing if IgM serology is positive in the first 20 weeks gestation.  
|                        | • High IgG avidity indicates infection >3mths ago, low IgG avidity indicates infection <3mth ago  
|                        | • IgM can be detected 2 weeks after infection, peaks at 1 month and declines by 6 months  
|                        | • IgG peaks 1-2 months after infection and remains lifelong  
|                        | • PCR on amniotic fluid in antenatal period can confirm fetal infection  
|                        | • Placental tissue sample sent for toxoplasma PCR (although most positive placenta samples are also detected by other tests)  
|                        | • If toxoplasma infection is considered then treatment of the mother with pyrimethamine, sulphonamide and folinic acid may decrease the severity of the disease in the fetus.  
|                        | • Serial ultrasounds are needed to monitor the pregnancy |

| Investigation for Infant | • Serology toxoplasma IgM and a baseline IgG (only 75% of congenitally infected infants will produce detectable IgM)  
|                        | • PCR on blood, urine, CSF  
|                        | • Head ultrasound, ophthalmology review, universal hearing screening with aABR |

| Management | • Isolation is not needed  
|            | • Confirmed congenital infection  
|            | • Treat with pyrimethamine, sulfamethoxazole, folic acid for 1 year  
|            | • IgG will still be present after 1 year and titres will rise  
|            | • Not confirmed congenital infection (likely maternal infection and no transfer)  
|            | • IgM should be negative  
|            | • IgG titres will fall over time (as they are the maternal antibodies)  
|            | • Transplacental IgG from mother’s infection should disappear by 6-12 months |

## PARVOVIRUS

| Symptoms | Maternal Symptoms: Illness with rash, fever, myalgia, arthritis, +/- anaemia.  
<table>
<thead>
<tr>
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</thead>
<tbody>
<tr>
<td></td>
<td>Fetal/Neonatal Signs: Anaemic or hydropic infant noted on ultrasound scan or known maternal seroconversion in antenatal period particularly between 10 – 20 weeks gestation. Risk of fetal demise is 2-6%.</td>
</tr>
</tbody>
</table>

| Investigation for Mother | • Serology for Parvovirus IgG, IgM (positive IgM indicates infection within past 2-4mths)  
|                        | • Obstetric specialist input required.  
|                        | • Consider USS, MCA Doppler velocity monitoring and fetal blood sampling if anaemia is suspected  
|                        | • Tissue sample from placenta sent in sterile pottle with no saline. |

| Investigation for Infant | • If hydropic infant or stillborn, send tissue sample from placenta as above, and this will be tested for Parvovirus PCR |

| Management | • No specific treatment available  
|            | • Contact isolation |
### RUBELLA

| Symptoms | Maternal Symptoms: Routine antenatal screen at booking. Testing done after contact with rubella or symptoms of fever, erythema, lymphadenopathy or arthralgia.  
Fetal/Neonatal Signs: Retinal pigmentation, cataracts, glaucoma, microcephaly, sensorineural deafness, pneumonitis, hepatosplenomegaly, thrombocytopenia, blueberry muffin lesions |
| --- | --- |
| Investigation for Mother | • Rubella IgM and IgG if there is a rubella contact and/or symptoms of rubella  
• 85% chance of transmission to fetus if contact rubella in first 12 weeks of pregnancy  
• Obstetric specialist input required |
| Investigation for Infant | • Serology rubella IgM (note there can be false positives and negatives)  
• Consider sending EDTA tube from cord or infant blood for rubella PCR  
• Consider urine/CSF for rubella PCR  
• Head ultrasound and ophthalmology review  
• Universal hearing screening with aABR |
| Management | • No specific treatment  
• Can be infectious for the first year of life after congenital rubella  
• Contact isolation |

### VARICELLA ZOSTER

| Symptoms | Maternal Symptoms: chicken pox vesicles, pneumonia. Contact with chicken pox later in pregnancy carries a risk for the infant – living with a person in the same household or face to face contact for >5 minutes are considered risk factors.  
Fetal/Neonatal Signs: infection in the first trimester can cause congenital varicella syndrome in 1-2% - limb hypoplasia, skin scarring, eye and CNS anomalies |
| --- | --- |
| Investigation and management for Mother | • If vesicles evident, swab the base of the vesicle and send for VZV/HSV PCR  
• If previous history of Chicken pox is unknown – request urgent VZV IgG serology (IgM is unreliable) |
| Treatment for exposure in seronegative women: | • If mother is seronegative and she presents within 4 days from chicken pox contact, mother should get ZIG to attempt to prevent infection developing.  
• If consultation is greater than 4 days from chicken pox contact, no ZIG is required.  
• Oral aciclovir is given in the 2nd half of pregnancy, in the immuno-compromised, in a smoker or a woman with underlying lung disease. |
| Treatment of women with active chicken pox | • If seen within 24 hrs, mother to get oral aciclovir.  
• If seen after 24 hrs, no aciclovir.  
• If seen after 24hrs and considered high risk/ at risk of complications, mother to get IV aciclovir.  
• If mother develops chicken pox 5 days prior to 2 days after birth, infant should receive ZIG. |
| Management of Infant | • If maternal chickenpox onset is 5 days prior to delivery or develops within 2 days of birth, infant to have ZIG  
• If maternal chickenpox onset is greater than 7 days prior to delivery, no ZIG necessary  
• Healthy term infants exposed to chicken pox outside these timeframes in a mother who has not had chicken pox do not need ZIG but should seek medical review if any lesions occur as there is a risk of severe disease that may need treatment. |
| Treatment for Infant: | • In an infant who develops chickenpox and is very preterm, has respiratory disease and or severe chicken pox disease, this infant to have IV aciclovir administered.  
• ZIG to be given if the neonate is preterm and there is no maternal history of chicken pox  
• ZIG to be given if <28wks or <1000gm regardless of maternal chicken pox status  
• Isolate (contact and airborne precautions) if the baby has active lesions until they crust over  
• Infants with embryopathy at birth do not need isolation |
| Symptoms                  | Maternal symptoms: Fever, encephalitis, myositis, Hand Foot and Mouth disease.  
|                          | Fetal/Neonatal Signs: Nonspecific but can include apnea, sepsis, meningitis, hepatitis |
| Investigations for Infant | • Call Microbiologist to discuss appropriate testing required.  
|                          | • Samples can be sent for PCR or culture  
|                          | • Nasopharynx/throat/rectal swabs may be done after discussion with microbiology  
|                          | • If doing an LP send the CSF for PCR |
| Management               | No specific treatment |
There has been a rapid rise in syphilis cases among women in recent years with a corresponding increase in cases of congenital syphilis. Incorrect or delayed management or inadequate follow up can result in increased morbidity

There should be information in the antenatal folder regarding women that are known to have syphilis

| Definitions | Acquired syphilis can be divided into **primary** (ulcer or chancre stage), **secondary** (systemic dissemination), **early latent** (> 2 years since acquisition with no symptoms), **late latent** (> 2 years since acquisition with no symptoms) and **tertiary** syphilis (symptomatic late syphilis e.g. gummas, cardiovascular and neurological involvement).

The risk of congenital syphilis is very high during the first 4 years after acquisition of syphilis and is negligible after 8 years of infection.

| Maternal and Antenatal Risk factors | The risk of congenital infection for untreated pregnant women is 100% for primary syphilis and secondary syphilis, 80% for early latent and 10% for late latent syphilis. The risk of congenital syphilis in women treated during pregnancy is between 1-2%.

| Antenatal Scans | Ultrasound signs of congenital syphilis in the fetus include hepatomegaly, placentomegaly, polyhydramnios, ascites and elevated middle cerebral artery peak systolic velocity. Less frequent findings include bowel dilatation and long bone abnormalities. Abnormal ultrasound finding prior to treatment is associated with treatment failure and delivery of a neonate with congenital syphilis.

| Understanding Investigations |  
| --- | --- | --- | --- |
| **Reactive** | Reactive | Reactive | Confirmed syphilis infection |
| **Reactive** | Reactive | Non-reactive | Evidence of past treated syphilis or latent infection OR very early infection |
| **Reactive** | Non-reactive | Reactive | Biological false positive OR very early infection Repeat in 2 weeks |
| **Reactive** | Non-reactive | Non-reactive | Possible early primary, latent or false-positive, retest in one month |
| **Non-reactive** | Not tested | Not tested | No evidence of syphilis, or too early, retest in one month if strong suspicion based on clinical evidence |

A decrease of RPR titres after treatment from 1:16 to 1:4 is indicative of adequate treatment. Conversely for example, a rise in RPR titres after treatment from 1:2 to 1:8 is indicative of re-infection or treatment failure.

| Symptoms |  
| --- | --- |
| Babies born to all women treated for syphilis in current pregnancy require evaluation at birth. |

The newborn infant should be examined for the following clinical features:

- IUGR / Unexplained enlarged placenta.
- Hepatomegaly/splenomegaly
- Necrotising funisitis – inflammation of the umbilical cord
- Fever / Jaundice.
- Non immune hydrops fetalis (NB: check for parvovirus).
- Generalised lymphadenopathy.
- Snuffles, haemorrhagic rhinitis.
- Bullous lesions, palmar/plantar rash,mucous patches.
- Condylomata lata.
Investigations that may suggest diagnosis
- Osteochondritis/periostitis.
- CNS signs, elevated cell count or protein in CSF and no other cause found
- Haemolytic anaemia, DIC, thrombocytopenia.
- Pneumonitis, Nephrotic syndrome.

**Investigation for infant**
The interpretation of syphilis serology in neonates requires specialist input as no single test can be used to diagnose congenital syphilis. Passive transfer of maternal antibodies makes interpretation of neonate serology more complex
- Paired venous blood samples: RPR serology paired with mother
  - Send a neonatal venous blood sample for syphilis serology: request serum treponemal EIA, RPR, treponemal IgM (available through select laboratories in NZ). Take blood from the neonate, not the umbilical cord.
  - Send a maternal venous blood sample for serum RPR at the time of delivery if no result available within the past 4 weeks from the same lab
- Send further tests as clinically indicated below

**Management**
See below
<table>
<thead>
<tr>
<th>Category</th>
<th>Findings</th>
<th>Evaluation</th>
<th>Treatment</th>
<th>Follow Up</th>
</tr>
</thead>
<tbody>
<tr>
<td>Proven, or highly probable congenital syphilis</td>
<td>Abnormal physical examination consistent with congenital syphilis OR A serum RPR titre fourfold high than the mother’s titre on 2 occasions (e.g. mother’s RPR 1:4, infants 1:16) or infant IgM positive OR T. pallidum PCR assay of lesions or body fluids reactive</td>
<td>• CSF analysis (VDRL, cell count, protein) • FBC, EUC, LFT • Long-bone X-Rays • Other tests if needed: - Chest X Ray - Neuroimaging • Ophthalmologic examination • Formal audiologic examination • Examination • Placental histology and syphilis PCR</td>
<td>Benzylpenicillin 50,000U (30mg)/kg/dose IV every 12 hours during the first 7 days of life AND every 8 hours thereafter for a total of 10 days *</td>
<td>1) Paediatric review at 6wks, 3mths, 5-6 mths and 12-18 mths of life. 2) RPR expected to be negative at 6 months 3) If congenital neurosyphilis diagnosed at birth- repeat CSF analysis every 6 months until normal parameters 4) If infant RPR increasing or not decreasing may need repeat LP / retreatment</td>
</tr>
<tr>
<td>Asymptomatic possible congenital syphilis</td>
<td>Normal clinical examination AND serum RPR equal to or less than fourfold the maternal titre AND ONE OF THE FOLLOWING Mother not treated, inadequately treated or no documentation of treatment OR Mother treated with a non-penicillin regimen OR Mother received recommended treatment &lt;4 weeks before delivery</td>
<td>• CSF analysis (VDRL, cell count, protein) • FBC, EUC, LFT • Long-bone X-Rays • Placental histology and syphilis PCR</td>
<td>Benzylpenicillin 50,000U (30mg)/kg/dose IV every 12 hours during the first 7 days of life AND every 8 hours thereafter for a total of 10 days * Note: For some infants where CSF examination and other investigations normal and where follow up can be assured, then benzathine benzylpenicillin tetrahydrate 50,000U/kg IM as a single dose may be used after discussion with Paediatric ID specialist #</td>
<td>1) Paediatric review at 6wks, 3mths, 5-6 and 12-18 mths of life with repeat RPR 2) RPR expected to be negative at 6 months 3) If congenital neurosyphilis diagnosed at birth- repeat CSF analysis every 6 months until normal</td>
</tr>
</tbody>
</table>
| Congenital syphilis less likely | Normal infant examination | None needed | Repeat serology at 6 weeks, 3 and 6 months
| | | | OR
| | | | If any concern regarding follow up or lack of required maternal testing then GIVE benzathine benzylpenicillin tetrahydrate 50,000U/kg IM as a single dose #
| | | | 1) Repeat syphilis serology at 3 months – if all negative – discharge
| | | | 2) If syphilis serology reactive then repeat at 3 monthly intervals until negative
| | | | 3) RPR is expected to be non-reactive at 6 months
| | | | - any passive cross over of treponemal antibodies will be negative by 15 months of life.

Candidal Infections

- The rate of invasive candidiasis for ELBW infants is about 2%
- Our incidence of systemic fungal infections (candidaemia, candiduria, candida meningitis) is low
- Invasive infection usually occurs after the first week ie: is a late infection
- Untreated the mortality is >80% and treated mortality is still up to 60%
- Infants who have had systemic candida have a higher morbidity for outcomes such as ROP, CLD and developmental delay
- Candida may be the cause of chorioamnionitis which would require the use of early systemic antifungal agents

Risk Factors

- VLBW
- Prolonged intubation
- Antibiotic use (especially 3rd generation cephalosporins)
- Parenteral nutrition
- Indwelling catheters
- H2 receptor antagonists
- Vaginal delivery
- Postnatal steroids

Colonisation

- Superficial infections (particularly in napkin area) increase the longer an infant is in the unit and are common
- Transmission may be vertical ie: acquired from the vagina during birth, or nosocomial.
- Colonisation at birth is particular troublesome for extremely preterm infants (< 28 weeks) so review of maternal swabs in the days before birth and of the initial swabs and gastric aspirate may identify a baby at particular risk.

Clinical Features

- Nonspecific
- Lethargy, respiratory distress, hyperglycaemia, abdominal distension, blood in stools
- Thrombocytopenia is common but not diagnostic

Treatment

- Cutaneous candida should be treated with topical cream as well as oral suspension
- Systemic candida should be treated with intravenous antifungals eg: fluconazole or amphotericin

Prophylaxis

- Systemic candida infection is associated with extreme prematurity, BWT < 1000g, prolonged antibiotic treatment, prolonged TPN use and systemic steroids.
- All infants <28 weeks gestation should receive prophylactic oral nystatin until discharge from level 3.
- Infants 28 – 32 weeks should receive nystatin prophylaxis if they are colonised with candida.
- All infants should receive nystatin prophylaxis if they are on a prolonged antibiotic course for > 7 days, on postnatal steroids or have long-term central access and TPN use or on H2 receptor antagonists
Management of Babies born to Hepatitis B, C and HIV Positive Mothers

- Wear gloves when handling the baby until they have been bathed
- Bath the baby in warm water only to remove any maternal secretions
- Clean the skin with an aqueous chlorhexidine (alcohol-free) swab prior to giving im Vitamin K
- Hep B positive mothers – baby to receive Hep B immunoglobulin and vaccination as soon as possible (see immunisation section)

HIV Positive Mothers

- Pregnant women are routinely offered antenatal HIV screening with the first trimester bloods. Late presenters with high risk factors should be screened after this time.
- The probability of an HIV positive woman transmitting the virus to her baby during pregnancy, labour, delivery or breastfeeding ranges from 15-25% in an industrialised country with no intervention. (Msellati et al 1995, Kreiss 1997).
- It is now well established that combining elective caesarean section and antiretroviral drug therapy and not breastfeeding reduces the risk of vertical transmission to < 1%. (Connor et al 1994, European Collaborative Study 1999, BHIVA 2014).
- In pregnancies where mothers are treated with anti-retrovirals transmission occurred in 0.8% delivered by elective caesarean at 37 weeks. (Mandelbrot et all 1999). More recently where there is undetected viral load before delivery vaginal delivery is deemed acceptable (BHIVA 2014).
- The British HIV association guideline 2014 is a good resource. Recent cohort and RCT data from Africa show maternal ART treated women with suppressed viral load who breast feed have reduced but not abolished transmission. Avoidance of breast feeding remains our recommendation.
- Management of HIV infected women during pregnancy and child birth, is a document found on the CDHB Intranet under Maternity guidelines

Neonatal Management

- The mother will receive antiretroviral therapy starting between 14-34 weeks and will also receive intrapartum antiretroviral therapy as determined by the ID. An ID nurse specialist follows the women closely.
- Written notification of the case antenatally by obstetric staff to Dr Nicola Austin who is responsible for the follow up of these infants
- Notification to Dr Austin and the Infectious Disease specialist involved should occur at birth and especially if delivery is earlier than planned/expected
- An elective caesarean section or vaginal delivery if undetected viral load is offered at 37-38 weeks.
- The baby should be bathed ASAP after birth in warm water to remove any secretions and blood
- The baby will be seen and examined by the neonatal team after birth and should be handled with gloves until washed.
- Vitamin K is recommended for all babies. If formula fed then a single oral dose (2mg) is sufficient. If the IM route is chosen it should be done after the baby has been bathed and the skin must be cleaned with an aqueous chlorhexidine (alcohol-free) swab prior to any injection

Drug Dosing (also refer to Neonatal Drug Profiles)

- With the parents permission the baby should receive the first dose of oral antiretroviral syrup within 4-6 hours of birth. Aim for dosing at 0800 and 2000 if possible and from then on dosing should be given strictly to time
- Zidovudine (AZT) is the usual drug given:
  - ≥ 35 weeks  4mg/kg/dose orally 12 hourly, or 1.5mg/kg/dose IV 6 hourly
  - < 35 weeks  2mg/kg/dose orally 12 hourly for 2 wks then 3mg/kg/dose orally 12 hourly for 2 wks, or 1.5mg/kg/dose IV 12 hourly
- Treatment should continue for 4 weeks total
- Liaison with pharmacy should have occurred prior to delivery of the infant to ensure this is available. A supply is kept on Birthing Suite when cases are known.
- Cotrimoxazole prophylaxis against Pneumocystis carinii may commence at 4-6 weeks of age if there is a recent history in the parents or the HIV PCR is positive after birth. The dose is 120mg (0.5mL of 240mg/5mL syrup) once a day three days a week eg. Mon, Weds, Fri
Blood Monitoring

- Day 2 – FBC and differential baseline and HIV Proviral DNA.
  At least 2 but preferably 3 EDTA tubes are needed and are sent to Auckland
  ** To ensure the blood tubes for HIV DNA testing are sent to Akld and not the smaller FBC tube please put the 2 full tubes for HIV DNA in a separate bag with a separate form to the FBC tube
- 2 weeks - FBC and differential (AZT can cause anaemia and neutropenia) liver function, renal function
- 6 weeks – FBC and differential and HIV Proviral DNA
- 4 months – FBC and differential and HIV Proviral DNA
- HIV Proviral DNA taken preferably on a Mon-Wed before 1200 for the 6 wk and 4 mth time periods.

Feeding

- Recommend formula feeding as a further means of reducing risk of vertical transmission
- The ID specialist obtains funding for formula where appropriate. See MOH, Breastfeeding by women with HIV 2016.

Follow-Up

- Dr Austin will follow up these infants at 2 weeks, 6 weeks of age and 4 months on the neonatal unit in the assessment room.
- BCG or live vaccines should not be given until after the 4month HIV PCR test result is known. The exception is Rotavirus which can be given if the first HIV PCR is negative and the lymphocytes are adequate at 2 weeks.
\textbf{HAEMATOLOGICAL DISORDERS AND JAUNDICE}

\textbf{Red Flags for Jaundice}

The following situations are where babies need bilirubin levels to be taken

- Known maternal blood group sensitisation with antibodies detected eg: Rhesus isoimmunisation, ABO incompatibility, other antibodies
- Family history of significant jaundice eg: due to blood group incompatibilities, hereditary spherocytosis, G6PD deficiency in males
- Preterm infants
- Any baby with visible jaundice in the first 24 hours
- Birth trauma eg: bruising, cephalhaematoma
- Polycythaemia (can occur after deferred cord clamping)
- Poor feeding and dehydration
- Sepsis
- HIE or other casues of acidosis
- Low albumin levels
- Dark pigmented skin (unable to assess jaundice levels visually)
- Ethnicity – increase risk in Asians, Mediterranean, African, Middle Eastern due to skin colour + risk of G6PD

Due to the high risk nature of the babies admitted to NICU a pragmatic approach has been taken so that every baby having a 48 hour Guthrie test in NICU will also have a screening bilirubin level taken with a blood gas.

\textbf{Phototherapy Charts}

- These are to be used as a guide for when to start phototherapy or consider an exchange transfusion
- We have phototherapy charts for different gestations but setting light levels and exchange levels should also be based on the clinical situation. These may alter depending on the presence of asphyxia, hypoxaemia, acidosis, hypothermia and sepsis
- The treatment lines have been calculated with evidence from the AAP and NICE guidelines and all data for phototherapy and exchange transfusion is based on experience with severe rhesus haemolytic disease only.
- There are up to 3 lines per chart - a level for considering an exchange transfusion, a level for standard phototherapy and a level for babies ≥35 weeks with haemolytic disease (there is no evidence to create a similar line for lower gestations)
- Completing all the parts of the phototherapy chart ensures that at a glance all the information is present to make decisions on starting or stopping treatment
- The phototherapy lines are indications to start phototherapy and do not guide when to stop phototherapy. This decision is made taking into account the risk factors for jaundice, the rate of rise or fall of the bilirubin, the number of lights the baby is on and how far below the treatment line the bilirubin is.
- It is best to have the bilirubin significantly under the treatment line before stopping lights or the baby may rebound quickly back on to lights. There is no need to have 2 results below the line if the result is well below the line ie: >50μmol/L under.
- Keep in mind that there is no good specific evidence as to what constitutes a dangerous bilirubin level in a particular baby, so it is impossible to come up with sound, evidence based rules. Clinical judgement will be required. If in any doubt discuss with a consultant.

\textbf{Even in a well infant, investigation and phototherapy should be considered in the following circumstances}

<table>
<thead>
<tr>
<th>Birth Weight and Maturity</th>
<th>Rate of Rise of Bilirubin in First 48 Hours, Averaged over 8-12 hours</th>
<th>Peak Bilirubin Above</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>1\textsuperscript{st} 24 hours</td>
<td>2\textsuperscript{nd} 24 hours</td>
</tr>
<tr>
<td>&lt;1000 g</td>
<td>&gt; 3 mmol/hour</td>
<td>100 μmol/L</td>
</tr>
<tr>
<td>1000-2000 g</td>
<td>&gt; 4 mmol/hour</td>
<td>120 μmol/L</td>
</tr>
<tr>
<td>&gt; 2000, Premature</td>
<td>&gt; 6 mmol/hour</td>
<td>150 μmol/L</td>
</tr>
<tr>
<td>Term</td>
<td>&gt;8 mmol/hour</td>
<td>170 μmol/L</td>
</tr>
</tbody>
</table>
How to use phototherapy charts

- Ensure you have the correct chart using the birth gestation at all times.
- It is important to ensure the correct phototherapy chart is used as the treatment levels vary according to the infant’s gestation.
- Fill in the top box with date and time of birth, maternal blood group, evidence of antibodies or haemolysis.
- When deciding if there are red flags for jaundice refer to the back of the phototherapy page under red flags and then indicate if these are present yes or no in the top box and list the red flags.
- Careful thought about the aetiology of the jaundice and appropriate investigation is at least as important as phototherapy, and may lead to identification of another specific therapy.
- In the right hand column ensure the date, time and result of the SBR or bilirubinometer reading are recorded and plot on the graph (each square is 2 hours).
- Record number of lights or bilibed in the box - number of lights. This is important to help assess the response to treatment.
Physiologic jaundice

“Physiologic” jaundice does not need phototherapy, but frequent feeds (preferably breast feeds) should be encouraged. There is good evidence that the frequency of breast feeding is as, or more important than the exact volume the baby receives. Infants with physiologic jaundice will be:

- Healthy term infant, weight loss <10% birth weight
- No blood group “set up” for haemolysis
- Rate of rise less than 8 µmol/hour - i.e. not visible in 1st 36 hours
- Normal ‘Guthrie’ screen
- Peak less <300 µmol/L

Investigation of Jaundice

- jaundice in the first 24 hours
  - Full blood count, film and reticulocyte count.
  - Group and Coombs, cross match
  - Maternal group.
  - CRP, NEON
  - Blood culture
  - Urine culture and LP if clinically indicated

- Jaundice requiring phototherapy on day 3-5 in a term baby
  - Full blood count, film and reticulocyte count.
  - Group and Coombs
  - CRP, NEON
  - Blood culture if clinically indicated
  - Make sure Guthrie card has been sent and check results with the National Testing Centre
  - Consider G6PD deficiency screen if male infant of African, Mediterranean, Middle Eastern or Asian ancestry.
  - Review current weight versus birthweight and feeding history
• Jaundice approaching exchange level
  - Full blood count, film and reticulocyte count.
  - Group and Coombs, cross match
  - Maternal group.
  - CRP, NEON, Liver function tests including conjugated bilirubin
  - Blood culture
  - Make sure Guthrie card has been sent and check results with the National Testing Centre

• Jaundice in a sick infant or jaundice that improves and then recurs
  - Group and Coombs
  - FBC, film and WBC differential, reticulocyte count
  - Blood cultures, urine cultures, CSF examination, CRP.
  - Consider G6PD screen if male infant is of African, Mediterranean, Middle Eastern or Asian ancestry.
  - Conjugated bilirubin and liver function tests
  - Make sure the Guthrie card has gone and check results with the National Testing Centre

• Prolonged jaundice:
  - Defined as:
    Term infants >14 days age with visible jaundice or an unconjugated bilirubin >150umol/L
    Preterm infants >21 days age with an unconjugated bilirubin >150umol/L
  - Liver function tests including conjugated and unconjugated bilirubin
  - FBC, film and reticulocyte count
  - Urine culture only if clinically likely that a UTI is present
  - T4/TSH
  - Make sure the Guthrie card has gone and check results with the National Testing Centre
  - Potential causes are biliary atresia, hypothyroidism, hypopituitarism, TPN cholestasis, other endocrine or metabolic causes

Conjugated Hyperbilirubinaemia

Conjugated bilirubin > 20 mmol/l or >20% of total bilirubin level

The most common cause of this in NICU is liver cholestasis from prolonged TPN use.

The list of investigations to rule out other causes is long and should be done in a step-wise fashion and with referral to the Paediatric Gastroenterologist when conjugated jaundice is identified.

First set of investigations to be done by NICU team before referral to Gastroenterology:
• FBC, film
• Group and Coombs
• LFTS – including AST, ALT, GGT, ALP
• Bilirubin – conjugated and unconjugated
• Albumin
• Coagulation profile
• TFT
• Urine CMV
• Maternal antenatal bloods results for rubella, toxoplasma, Hepatitis B
• Check Guthrie result for CF screen
• Liver ultrasound

Further investigations will be dictated by Gastroenterology after their review and may include:
• Alpha-l-antitrypsin phenotype
• Ferritin
• Cortisol
• Cholesterol/Triglycerides
• Cystic Fibrosis genetic screen (by PCR)
• Sweat test
• HIDA scan is now rarely required and only after discussion with Paediatric Gastroenterologist
Management Options

- Lipid infusion over 12 or 20 hours may be advised by Gastro and there are TPN templates to prescribe this.
- Vitamin supplements may be recommended and Vit A and E levels may be checked on these babies to ensure they are not too low despite supplementation.
  - Vitadol C (high dose 1mL daily)
  - Vitamin E (high dose 0.5mL daily)
  - Vitamin D (0.03-0.05 microgram/kg/day)
  - Vitamin K 2mg daily if INR abnormal (dose may need to increase if coagulation does not improve)
  - Ursodeoxycholic acid (10mg/kg/dose bid) if requested by Gastro

Bilirubinometer

- The transcutaneous bilirubinometer is a non-invasive screening device.
- The meter measures the yellowness of the subcutaneous tissue of the infant by measuring the difference in optical density of two different wavelengths of light through the skin.
- Accuracy is dependent on correct usage of the device, therefore all staff are required to have completed Point of Care (POC) training if using the Konica Minolta JM-103® bilirubinometer.
- Refer to Transcutaneous Bilirubin measurements - Maternity guideline for checking and use of the device.
- Measurements are taken by placing the instruments measuring probe against the infants forehead or sternum and gently pressing downwards.
- This was introduced into use in the postnatal ward March 2017.

Criteria for Bilirubinometer Screen

- ≥ 35 weeks gestation
- More than 24 hours of age

Criteria for SBR Testing only and not Bilirubinometer Screening

- Jaundice < 24 hours age
- Preterm infants < 35 weeks
- Significant bruising over the forehead where the TcB will be measured
- During or after phototherapy

Process for TCB Measurement Recording

- Record all measurements in clinical notes and plot on the phototherapy chart with notation “TcB” with recorded level. If TcB level, requires SBR to be performed then midwives to take the SBR, plot the result on the phototherapy chart and contact the Neonatal Ward staff with the result if phototherapy required.

<table>
<thead>
<tr>
<th>Jaundice Onset</th>
<th>TcB</th>
<th>Action</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt; 24 hours</td>
<td>Not applicable</td>
<td>Perform SBR</td>
</tr>
<tr>
<td>24-48 hours</td>
<td>&gt; 140 µ mol/l</td>
<td>Perform SBR</td>
</tr>
<tr>
<td>48-72 hours</td>
<td>&gt; 200 µ mol/l</td>
<td>Perform SBR</td>
</tr>
<tr>
<td>&gt; 72 hours</td>
<td>&gt; 250 µ mol/l</td>
<td>Perform SBR</td>
</tr>
</tbody>
</table>

Haemolytic Disease of the Newborn

- We should be made aware antenatally of all pregnancies where is evidence of **significant haemolytic disease**.
- Inform the consultant of all deliveries of babies whose mothers have significant antibody titres.
- Two units of blood for exchange should be cross matched against the mother and be available before delivery.
- Not all atypical maternal antibodies cause haemolytic disease in the newborn, regardless of titre. However, they can present problems with crossmatch.
Frequency of Haemolytic Disease of the Newborn

<table>
<thead>
<tr>
<th>Category</th>
<th>Antibody</th>
</tr>
</thead>
<tbody>
<tr>
<td>Common</td>
<td>D, c, E, Kell</td>
</tr>
<tr>
<td>Uncommon</td>
<td>e, Ce, cE Kpa, Kpb, k S, Wra,FyA</td>
</tr>
<tr>
<td>Very rare</td>
<td>S, U, M, N Fyab, Doa, Coa Dp, Dp, Lu a, Yt a, Jka, Jkb</td>
</tr>
<tr>
<td>No occurrence</td>
<td>Le a, Le b, P</td>
</tr>
</tbody>
</table>

- Cord blood must be analysed for blood group, DCT, bilirubin, full blood count and reticulocyte count.
- In all but mildly affected babies, consideration should be given to inserting umbilical arterial and venous catheters soon after birth, because the success rate of insertion is much higher then and exchange transfusion is much more difficult without them. Discuss this with the consultant.

Timing of Exchange Transfusion

Decided on the basis of the cord haemoglobin results, the rate of fall of the Hb and the rate of rise of the bilirubin:

- Hb < 100 g/L Immediate exchange likely to be necessary
- Hb 100-120 g/L Exchange likely to be required in the first day
- Hb > 120 g/L Wait. Treat with phototherapy.

Bilirubin Estimations

- The bilirubin should be estimated every 4-6 hours.
- If the bilirubin is rising at > 17 mcmol/lour, or if it exceeds 340 µmol/L in first 24-48 hrs, an exchange should be considered.
- See section on procedures for details on how to perform an exchange transfusion.
- There will be differences in the bilirubin levels when measured in the lab and on our gas machine in those babies who have received intrauterine blood transfusions or exchange transfusions. This is because their circulating blood will have components of adult Hb.
- The more accurate bilirubin measurement after a baby has received adult blood (intrauterine or exchange transfusions) is a formal lab measurement

Immunoglobulin

- Intravenous immunoglobulin has been shown to reduce the need for exchange transfusions in rhesus haemolytic disease and ABO incompatibility.
- Immunoglobulin 0.5-1g/kg and repeated after 12 hours should be considered if high antibody titres and considered high risk for an exchange or the bilirubin is rising towards exchange transfusion levels despite intensive phototherapy.
- Blood Bank have 2 products – Intragam P (60mg/mL) and Privigen (100mg/mL)
- Intragam is supplied by volunteer NZ donors with supplies being less available and so the preference from Blood Bank is to use this product for patients requiring immunoglobulin for over 3 months
- Privigen is supplied by paid USA donors and the preference from Blood Bank is to use this product for patients receiving short-term immunoglobulin infusions

Late Anaemia

- Late anaemia is a feature of haemolytic disease of the newborn except anti A and anti B HDN.
- Infants who are susceptible should have Hb checked periodically in the first month of life.
- Top up transfusions should be considered if the haemoglobin is < 70 g/l, especially if the reticulocyte count is low.
- Erythropoietin to prevent the need for transfusions for late anaemia has been used on individual babies. The main indication currently is where parents have objections to transfusion eg. Jehovah’s Witness.
- If erythropoietin is started too soon in haemolytic disease to prevent late anaemia, it could increase jaundice to dangerous levels, so this is probably not a useful therapy in babies whose bilirubin levels are still more than about half the exchange level.
Folate deficiency is common and all infants with Rh HDN or any other significant haemolytic anaemia should have supplementary folate (50 mcg daily for 3-4 months).

**Exchange Blood Transfusion Procedure**

**Indications**
- Serum bilirubin (unconjugated) in exchange transfusion range (this is usually 50–100 µmol above the phototherapy level depending on gestational and chronological age).
- Check conjugated bilirubin and subtract from total bilirubin, important if baby had in-utero transfusions. Lower exchange levels may be set for sick or extreme preterms, or if there is hypoalbuminaemia (albumin < 25g/L discuss with consultant).
- Rhesus disease - consider if cord Hb <100, cord bilirubin > 80µmol/l, or bilirubin will rise >340µmol/L
- Bilirubin increasing despite maximal phototherapy and fluid rehydration
- Drug / toxin / metabolite / immune factor removal

**Aims**
- Reduce the risk of brain damage and prevent kernicterus (by decreasing unconjugated bilirubin level)
- Prevent further haemolysis by exchanging sensitised red blood cells and removing maternal RBC Ab’s
- To control blood volume and relieve heart failure
- To improve anaemia and increase the oxygen carrying capacity

**Potential complications**
- Infection (bacterial or blood-borne)
- Embolism (air / clot)
- Electrolyte disturbances (including low calcium and magnesium)
- Haemodynamic (overload, shock, anaemia, polycythaemia)
- Clotting abnormality (EDTA in stored blood binds calcium)
- NEC (especially in “single-operator” method)
- Blood reaction
- Rebound hypoglycaemia
- Temperature instability
- Death (mortality 0.5%)

**Methods**
- A consultant must be present for an exchange transfusion.
- The retrieval registrar will need to be called in to respond to other calls from delivery suite or NICU.

**Two catheters / Two operators plus nurse**
- Simultaneous withdrawal and delivery using 10 or 20ml aliquots depending on size of infant.
- Each operator drawing in (designated “in”) and pushing out (designated “out”) blood by using three-way tap attached to their syringe

**Venous catheter / Single operator plus nurse**
- Only use this method if there is only venous access
- 5ml (<1kg); 10ml (<2kg) or 15-20ml (>2kg) aliquots
- Start with “out” and replace.

**Vascular Access**
- One artery and one vein (for “two-operator” exchange)
- UAC or peripheral artery (“out” volumes)
- UVC or large vein (eg. cubital fossa) (“in” volumes)
- Umbilical vessels may be used up to 10 days after birth
- Single umbilical vein (for “single-operator” exchange)
UAC/UVC position checked with X-ray
UVC preferably in IVC (not in hepatic branch) – use a 5 Fr single lumen catheter

**Volume to be exchanged**
- Volume = 2 x 85 ml x Weight (kg) (twice infants blood volume)
- Removes about 85% of baby’s red blood cells.

**Blood**
- In most exchanges for rhesus disease the blood is cross-matched against mother’s blood and available before delivery.
- Check with blood bank that they are aware of the mother being in labour (we receive a monthly summary of expected deliveries).
- If it is an exchange for a non-rhesus cause occurring after delivery, the blood needs to be cross-matched against the baby
- A baby <1500g or any baby who received intrauterine transfusions must have irradiated, CMV negative, leucocyte depleted blood. This is a preference for all other babies as well but not a necessity in cases where blood is needed urgently.
- In an emergency when urgent O Rh negative blood is used the appropriate leucodepletion filter will need to be used if the blood leaves blood bank before being leucocyte depleted
- Best to use fresh blood <5 days old
- Stored blood has high K⁺ (up to 15 mMol/l; ideally < 10 mMol/l)
- Irradiated blood must be used within 24 hrs as the potassium level increases post irradiation
- Best to use blood with a haematocrit of 0.55 (compared to the usual haematocrit of 0.7 as used for top-ups)
- Anticoagulant is used - citrate phosphate dextrose (binds Ca²⁺ and high dextrose)

**Equipment and setup**
See nursing OPS N 292 and 112. WHD 9743
- If not ventilated, nasal O2 if sats fall (this is usually needed during procedure as stored blood is poorly saturated)
- Monitoring - cardiorespiratory, saturations, blood pressure, temperature (every 15 mins throughout procedure)
- Strict timing of “in” and “out” volumes
- Strict recording of volumes and drugs
- Fresh blood reservoir above level of patient (for easier drawing up)
- Waste blood reservoir below level of patient (to prevent flowing back)
- Donor blood warmed to 37°C and shaken every 100ml stage during procedure (as red cells settle in the bag).
- Prophylactic antibiotics not indicated unless specific concerns about sterility.
- Full sterile procedure throughout – sterile gloves and gowns to be worn
- Glasses for all staff involved.
- Temperature, BSL, saturations and blood pH need to be optimal prior to commencement.
- Consent (written informed consent) from parent/guardian.
- Check infant ID and blood pack/s.
- Prepare all tubes and equipment before scrubbing up.
- Gastric contents must be evacuated - 8FG catheter on free drainage
- Place a urine bag
- Baby may need to be pacified during procedure.
- Suction and resus trolley need to be available in case of emergency.
- Lines need to be sited prior to procedure with X-ray position checked
Two Operator Procedure

**Step 1**

- Two doctors or CNS(ANP) and nurse recorder with consultant present
- Staff who begin the procedure must remain until procedure completed
- Blood tests from the first “out” specimen:
  - electrolytes, calcium, magnesium, LFT’s, SBR, conjugated bilirubin
  - blood culture
  - FBC, diff, film
  - ABG, BSL
  - Blood group and Coomb’s (if not already done)
  - Guthrie card
  - Saved specimens: clotted (plain) as well as EDTA samples (in case of further tests that may be needed at a later stage)
- Aim for a rate of 100ml per 15mins (or slower)
- Draw up calcium gluconate in a separate sterile syringe before commencing.
- Check ionised calcium on an ABG halfway through
- 1ml of 10% Calcium gluconate may be added to infused blood exchanged or injected slowly, watching pulse rate and rhythm.
- “In” operator will need to disconnect syringe to draw up calcium (directly from separate syringe and diluted with blood prior to injecting)
- ABG, BSL, calcium, K+ half-way through exchange (or more frequently in case of concerns eg. ECG changes)
- Blood tests from the last “out” specimen:
  - electrolytes, calcium, magnesium, SBR, conjugated bilirubin
  - blood culture
  - FBC (this is unreliable for a few hours after exchange)
  - ABG, BSL

**Subsequent monitoring and management**

- Blood sugar - this may drop (rebound result due to high glucose content of the anticoagulant
  - check hourly for 2 hours post exchange, then 3 hourly for 24 hours.
  - must have a continuous infusion of 10% dextrose post exchange.
  - Infants usually NBM
- SBR
  - post-transfusion expected to be around 50% of pre-transfusion level
  - By 4 hours post-transfusion: ↑ to 75% of pre-transfusion level
  - May continue increasing, check at 6-hourly intervals.
  - Continue intensive phototherapy
- Na
  - banked blood has high Na content
- FBC
  - major abnormalities with incorrect “out” & “in” balance
  - anaemia may be due to not shaking donor blood bag (ie. concentrated blood exchanged earlier, and thus removed, and dilute blood subsequently)
  - haematocrit more reliable after 4-8hrs post exchange (fluid shifts)
• Other
  – Hourly nursing obs for 6 hours (longer if abnormal)
  – Feeds - careful as ↑ risk of NEC (give breast milk feeds)
  – Inform parents as to outcome of exchange.

Blood Transfusion

• Many babies admitted to the NICU (mainly extreme preterms or surgical babies) should have a pink EDTA specimen sent to the Blood Bank for a Group and Hold and Coombs test
• Blood is supplied in " Paed Packs", one unit divided into 5.
• "Split Packs " are also available, these are one unit divided into 2
• For exchange transfusion the blood should be as fresh as is possible with a maximum age of 5 days.
• For top-up transfusions blood can be up to 35 days old.
• Before transfusing a baby > 1 month of age check that the blood bank still have a valid group and hold and the parental consent form is up to date.

Please write on the form if the baby is less than 1000 grams or is expected for any other reason to require multiple transfusions. The Blood Bank will then attempt to set aside all the segments of a unit for that baby, in order to minimise exposure to multiple donors.

If a baby changes their name after the blood has been sent for Group and hold then the request for blood QMR022A or blood products QMR0022B needs to use the label with the original name on it or a new sample will be requested by blood bank. The two common examples are: where the surname changes from mother’s to father’s after the mother is discharged from maternity or they are call “baby of X Y” and they then get a Christian name.

Reasons for Red Blood Cell Transfusion

• Anaemia of prematurity occurs in most premature babies. It is hypoerythroblastopenic anaemia associated with low erythropoietin levels and it lasts longer and results in a lower nadir Hb levels than the “physiologic” anaemia of infancy.
• It is hard to maintain iron sufficiency via parenteral/enteral nutrition in the first weeks in very premature infants.
• Sick babies are also at risk of anaemia of chronic illness.
• Vitamin E deficiency places some babies at risk of chronic haemolysis
• The greatest reason by far to transfuse babies is the "anaemia of chronic investigation". Therefore, keep blood tests to a minimum, and never draw more blood than is needed.
• However, the haemoglobin should initially be checked weekly in babies who were < 1500 g at birth.

Transfusion Thresholds

A guideline is given below, however, the clinical scenario, birth gestation, postnatal age, Hb level and reticulocyte response all need to be taken into consideration when deciding whether a transfusion is required

120g/L Term baby with acute blood loss in Week 1
  eg: fetomaternal transfusion, Rhesus isoimmunisation
  Ventilated
  BiPAP in oxygen

100g/L BiPAP in air
  CPAP
  Humidified high flow oxygen
  Nasal cannula oxygen

Clinical deterioration due to anaemia ie: lethargy, poor feeding, apnoea

80g/L Well baby over a week of age on no respiratory support
  Includes babies up to one month corrected age

See section below regarding reticulocyte response
Reticulocyte Count
- The reticulocyte count should be checked from 4-6 weeks of age - a good response is 4% or an absolute count of >100 and indicates bone marrow response
- If the reticulocyte count is adequate/ improving from previous levels it may be appropriate to not transfuse a well baby who is > 4 weeks old and just monitor the Hb weekly or earlier if there are clinical concerns.

Transfusion Volume
- Transfusion amount is usually 15mls/kg over 3 hours unless the baby requires acute volume expansion.
- In severe anaemia where transfusion volume is 20mls/kg or greater, blood < 5 days old is advised to avoid the effects of hyperkalaemia
- Frusemide (1 mg/kg) may be given if the blood is not being transfused to expand blood volume, although avoid it in the first week in ELBW babies because of literature suggesting that it may increase the risk of symptomatic patent ductus arteriosus. Always check with the consultant.

The Lamson tube system is now used for receiving blood product requests, send request by fax. A light will go on above the Lamson when something has arrived and it should be retrieved promptly and light cancelled.

Blood Product Requirements for Neonates
- Blood products used in neonates should be CMV negative.
- These guidelines outline situations in which it is recommended that blood products should be irradiated.
- Irradiation prevents the risk of transfusion associated graft versus host disease which is a serious and usually fatal complication of transfusion of which neonates are at increased risk. The majority of TA-GVHD cases reported in neonates have been in apparently immune-competent infants who have had intrauterine transfusion followed by exchange transfusion.

Emergency Blood Transfusion
- Usually requested from delivery suite at birth after an unexpected large fetomaternal haemorrhage
- Emergency blood for a neonate has different requirements from an adult. Blood should be <14 days old and CMV negative preferably. Blood allocated to the mother cannot be used for the baby.

Process
- If the CCO is not present call a 777 or press emergency bell and inform them urgent blood is needed
- Complete the QMR022A form (bold greenstrip on side) for the Nurse/Midwife/Dr to fill in
- Write DOB, surname and first name as Baby of ……, do not write in maternal details
- No NHI will be allocated so leave blank as this is not necessary
- Request 1 unit emergency O negative blood
- CCO will nominate a "runner" to run to Blood Bank LG floor Chch Hospital
- CCO will call Blood Bank to inform them that urgent blood is needed and a runner is on their way
- Anticipate this process should not take more than 10 minutes

Intrauterine and Exchange Transfusion
- All blood for intrauterine transfusion must be irradiated.
- It is essential to irradiate blood for exchange transfusion if there has been a previous intrauterine transfusion or the donation comes from a relative.
- For other exchange transfusions irradiation is recommended if it does not delay an emergency transfusion.
- Blood less than 5 days of age should be used for intrauterine and exchange transfusion and must be transfused within 24 hours of irradiation.

Top Up Transfusion in Neonates
- It is not necessary to irradiate blood for routine top up transfusions of premature or term neonates unless there has been a previous intrauterine transfusion or the blood has come from a relative.
Platelet Transfusions
- Irradiation must be performed on platelets transfused in-utero to treat alloimmune thrombocytopenia
- Irradiation must be performed on platelets transfused after birth to infants who have received either red cells or platelets in utero
- Irradiation must be performed when the platelet transfusion has come from a blood relative.

Granulocyte Transfusions
- These are used rarely but in all circumstances the granulocytes must be irradiated and transfused as soon as possible after irradiation.

Suspected Congenital Immunodeficiency
- Irradiation of cellular blood products is recommended for all infants with suspected T cell immunodeficiency
- There is no indication for irradiation of cellular blood products in children who are HIV antibody positive or who have AIDS.

Fresh Frozen Plasma
- When requested, this will be thawed by Blood Bank, and, once thawed has a shelf-life of 24 hours.
- If the transfusion cannot occur within 30 mins of receiving the product then return it to Blood Bank.
- Once started, the transfusion should be completed within 4 hours.

IV Immunoglobulin
- There is a national process to request immunoglobulin (IVIG) to streamline requests and to ensure appropriate clinical use in line with national guidelines
- The prescriber needs to be registered as an approved clinician (SMO, Reg, NNP) and this can be set up in advance at https://igo.nzblood.co.nz/account/login. The mobile number and email are required as that is how you will be contacted to inform you of your approval to prescribe and if your patient has been approved to receive immunoglobulin. Transfusion Medicine Specialists (TMS) must approve all applications.
- Once a clinician has been approved a request for IVIG needs to be completed from the IgO site. It involves putting in patient details and picking an indication from a drop-down list. Please note that you cannot leave any details blank - if a detail is not available/relevant, you can just put “xxx” to allow the form to go through.
- The request is then sent to the phone of the local TMS (or on-call if after hours) and is reviewed. After hours the TMS will need to be called to expedite the process. It is not automatically approved once the application is submitted. Once approved you will receive a text and the product can be charted on a fluid chart and requested from blood bank.

Blood Donation by Parents of Babies
- The NZ Blood service policy statement (Feb 2000) does not support the practice of directed donations on the basis that there is no evidence that such components lead to improved patient care nor that they reduce the risk of acquiring transfusion associated infections.
- Because of the possibility of shared HLA haplotypes within families, donations from family members poses particular risk of transfusion associated GVHD especially when the recipient is a neonate.
- This blood is never available in an emergency situation
- Parent blood donation is not available at Christchurch Women's Hospital

Thrombocytopenia
Reference: British Journal of Haematology 2011: 156; 155-162
- Commonly seen in neonates admitted to NICU and most are mild and self-limiting
- In the neonate usually defined as <150x10⁹/L, severe thrombocytopenia is <50x10⁹/L
- No exact level where the risk of haemorrhage can be predicted as the clinical scenario is also important
- Higher risk of bleeding if <50x10⁹/L with the commonest site being intracranial then gastrointestinal or pulmonary haemorrhage
- Falsely low platelet counts can be seen from cord blood or heel prick samples so a venous sample should be taken to confirm thrombocytopenia
Aetiology

- Artefact
  Clot in specimen.
- Dilutional
  After exchange transfusion or cumulative top-ups with platelet poor adult blood
- Hypoxia
  Placental insufficiency, PET, HELLP, IUGR, HIE
- Immune disorders
  Neonatal Alloimmune Thrombocytopenia (NAIT)
  Autoimmune Thrombocytopenia (maternal ITP, SLE)
- Consumption
  DIC, IVH, Kasabach-Meritt (giant haemangioma), hypersplenism.
- Infection
  NEC, Bacterial (GBS), viral (CMV, Toxo, Rubella, Coxsackie), fungal,
- Congenital
  Thrombocytopenia Absent Radius, Fanconi, Trisomy
- Metabolic disorders
- Leukaemia

Investigation

- Look for a possible cause listed above according to the clinical situation.
- Maternal platelet count
- Coagulation profile
- If NAIT is suspected see below for investigations

Placental Insufficiency – Common

- Platelet count falls to a nadir of >50x10^9/L by day 4-7 and spontaneously recovers in 2 weeks
- Associated transient neutropenia, increased nucleated red blood cells, polycythaemia
- As this is usually self-limiting do not need to investigate further unless the count remains <50 at 2 weeks

Neonatal Alloimmune Thrombocytopenia (NAIT) – Uncommon but severe consequences

- Most important cause of thrombocytopenia to rule out in term babies
- 10-20% will have an intracranial bleed
- Severe thrombocytopenia with no clear aetiology (see list above)
- Due to transplacental passage of maternal antibodies to fetal platelets with paternal HPA antigens that the mother does not have
- Usually resolves in 1-2 weeks but some cases are prolonged
- Screening head ultrasound is required
- Treatment is usually required immediately and before a diagnosis is confirmed
- Investigations (blood sent to Akld to National Tissue Typing Lab, D/W Haematologist)
  - identify maternal anti-HPA antibodies
  - genotype both parents blood for the common HPA antigens (1a,2,3,5b,15) anti-1a and anti-5b are the cause in 95% Caucasians
  - despite doing these tests only about 20% of cases are confirmed in the lab as maternal antibodies can be hard to detect or the HPA antigen may not be one of the common ones that is tested
- Management
  - Platelet transfusion – see below
  - Immunoglobulin – 1g/kg/day iv for 2 days
    Blood Bank have 2 products – Intragam P (60mg/mL) and Privigen (100mg/mL)
    Intragam is supplied by volunteer NZ donors with supplies being less available and so the preference from Blood Bank is to use this product for patients requiring immunoglobulin for over 3 months. Privigen is supplied by paid USA donors and the preference from Blood Bank is to use this product for patients receiving short-term immunoglobulin infusions

Platelet Transfusion

- Adult size packs suitable for neonatal use will be available from Blood Bank for urgent transfusions
- Split packs will be provided for neonates only after a specific request for this. It will take between 1-2 hours for this process to occur and so this can be requested for a non-urgent transfusion and/or for when a baby is identified with ongoing need for platelet transfusions. A single donor’s platelets will then be split into smaller aliquots and will have a 5 days expiry
- The aliquots for a split pack will on average be 45mL (range 30-60mL) and if larger volumes are needed this needs to be clearly requested on the QMR022B form
- Platelets only need to be irradiated if there have been any intrauterine platelet transfusions. It takes an additional 2 hours for the irradiation to be completed.
- Use 170-200 micron filter (to remove white cells), Platelets are CMV negative
- Generally give 10 mL/kg over 1 hour
- It is standard NZ blood service protocol to culture every bag of platelets, these cultures are observed for 5 days. Any hint of a positive culture is reported to the service caring for that patient and recommendation for commencing broad spectrum antibiotics. There are a number of false positive reports.
- Neonatal platelets are never frozen they are stored at room temperature. They are not suspended in the preservative solution but suspended in fresh plasma. There is the potential for increased risk of contamination. The shelf life as a consequence is 7 days. The stock level is kept low because there is limited demand and they have a short expiry date of 7 days.
- Any platelets or blood products being transported from other Blood transfusions services in NZ such as Auckland are required to follow stringent transportation regulations.

<table>
<thead>
<tr>
<th>Platelet Count</th>
<th>Clinical Indication</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt;20x10⁹/L</td>
<td>All Neonates</td>
</tr>
<tr>
<td>&lt;30x10⁹/L</td>
<td>&lt;1kg</td>
</tr>
<tr>
<td></td>
<td>First week life</td>
</tr>
<tr>
<td></td>
<td>Clinically unstable</td>
</tr>
<tr>
<td></td>
<td>Minor bleeding – ooze puncture site, petechiae, blood stained secretions</td>
</tr>
<tr>
<td></td>
<td>Abnormal coag profile</td>
</tr>
<tr>
<td></td>
<td>Surgery</td>
</tr>
<tr>
<td></td>
<td>Exchange Transfusion</td>
</tr>
<tr>
<td>&lt;50x10⁹/L</td>
<td>Major haemorrhage</td>
</tr>
</tbody>
</table>

- If NAIT suspected the preference is to have HPA-1a, HPA-5b negative platelets but there should not be any delays in transfusing the baby so use neonatal platelets first. For subsequent transfusions we need to discuss with the Transfusion Medicine Specialist about the possibility of NAIT and the availability of appropriate antigen-negative platelets
- If NAIT is suspected then after the first transfusion check the platelet count 1 hour later, then 12 hours later to document the peak and decay of the platelets. This does not need to repeated after every transfusion
- If concentrated neonatal platelets are required due to volume overload this is possible but takes longer. If required request these through Blood Bank who will discuss with the Transfusion Medicine Specialist. It takes up to 4 hours to prepare these and they expire within 4-5 hours of arrival at NICU. The volume will be 10-20mL and will be decided on by the Transfusion Medicine Specialist.

**Neutropenia**

- Defined as an absolute neutrophil count of < 1.5 x10⁹/L.
- Venous counts are 80% of capillary counts.
- Common causes related to infection e.g. bacterial, viral (especially CMV, HSV, parvovirus, enterovirus, or other viral and non-viral congenital infections in the neonatal period. Later hepatitis A and B, RSV, influenza A and B, measles, rubella, varicella become more prominent). Maternal toxaemia is also a possible cause.
- Treatment usually directed at the cause.
- G-CSF (granulocyte colony stimulating factor) can be considered if neutropenia accompanies significant systemic infection (dose of 5 micrograms/kg).
- Occasionally very healthy term babies will have extremely low neutrophil counts due to maternal anti-neutrophil antibodies. These babies may require only careful follow-up.
Polycythaemia

Risk factors
- delayed cord clamping
- cord stripping
- holding baby below mother at delivery
- twin to twin transfusion (typically polycythemic larger and anaemic smaller twin)
- fetal hypoxia due to maternal toxaemia, smoking, cyanotic heart disease, severe lung disease.
- infants of diabetic mothers with poor glycemic control.
- infants with syndromes including Down syndrome, Beckwith Weideman syndrome, trisomy 13, trisomy 18
- infants with certain endocrine/metabolic disorders - congenital adrenal hyperplasia, neonatal thyrotoxicosis.

Diagnosis
- Definition usually taken to be Hb > 220 g/l or PCV >0.65.
- However, viscosity is not linearly related to PCV, so if the baby has no symptoms, consideration can be given to avoiding reverse exchange, whereas if a baby has significant symptoms at PCV 0.60-0.65, reverse exchange may be indicated.
- Capillary samples tend to give PCV results that are 5-20% higher than central venous samples.
- Warming the heel before obtaining the specimen will give a better correlation.
- If the capillary PCV is above 0.65, a fast flowing venous PCV should always be done.

Consequences
- Poor feeding, lethargy and hypotonia, jitteriness, respiratory distress, cardiac failure, cyanosis, jaundice, hypoglycaemia, hypocalcaemia, convulsions, venous thrombosis, thrombocytopenia.

Treatment
- Partial exchange transfusion with saline, using from 20 ml/kg up to 30 ml/kg, with the intention to reduce PCV to about 0.55. Formula for calculating exchange transfusion is as follows:
  \[
  \text{Volume to exchange} = \frac{(\text{actual PCV} - \text{desired PCV}) \times 80 \times \text{wt (kg)}}{\text{actual PCV}} = \text{mls}
  \]
- In infants with high PCVs who are managed without reverse exchange, careful attention to fluid balance (avoidance of dehydration) is essential.

Haemophilia

Mothers who are known to be carriers should have received antenatal counselling with the support of a haematologists and obstetrician. Some of these parents may have opted for antenatal gene testing. A clear management plan for labour and delivery should be in place. Management of the newborn with known haemophilia is outlined below as per the National Guidelines. Close liaison should be maintained with Dr Mark Smith, Haematologist who will guide management.

Protocol
- take blood from the umbilical cord (or peripheral vein if cord blood specimen unobtainable/unsatisfactory) for urgent (result <3 hours) factor VIII/IX level.
- if urgent factor VIII/IX assay unavailable, do coagulation screen (upper limit normal APTT in newborn - 36 seconds)
- avoid heel pricks for coagulation studies or factor assays.
- suggest factor VIII/IX level is done on females born to carrier mothers to detect the occasional carrier female with low levels at risk of symptomatic bleeding.
- oral Vitamin K prophylaxis is effective in preventing classical haemorrhagic disease of the newborn, but ineffective in preventing late HDN. Increasing the dose or giving it weekly for a longer period increases the efficacy of the oral prophylaxis.
- alternatively, IM Vitamin K can be given, especially after factor replacement administered, providing pressure is maintained for a minimum of 5-10 minutes.
If factor assay indicates severe (<1%) or moderate (1 - 4%) factor VIII/IX deficiency

- sensitive communication of result to parents by experienced staff
- take further factor VIII/IX level
- consider immediate replacement with a single vial of recombinant factor VIII or factor IX without waiting for second result (caution if family history of inhibitors). Doses should be guided by advice of haematologist.
- follow newborn closely for a minimum of 7 days after birth through daily phone contact from Haemophilia Centre or GP and frequent midwife visits.
- educate parents regarding symptoms of intracranial haemorrhage - poor feeding, irritability, listlessness, full fontanelle, convulsions, pallor.
- US head if clinical suspicion of intracranial haemorrhage or at 24 hour.
- if confirmed intracranial haemorrhage - treat according to National Guidelines for Factor Replacement in Haemophilia

Suspected Coagulopathy

In infants where the diagnosis is not known antenatally but a coagulopathy is suspected the following approach should be used.

- Investigate for a coagulopathy in any infant with significant intracranial haemorrhage, subgaleal haemorrhage, easy bruising (including significant cephalohaematomas) or prolonged oozing from venepuncture sites, petechiae (especially if more generalised than simply over face which can be induced during a normal delivery) or a family history of a coagulopathy.
- Do not be dissuaded from considering haemophilia if haemorrhage is a presenting feature of a newborn's illness, even if:
  - coagulation screen suggests DIC
  - thrombocytopenia coexists with prolonged APTT
- Initial coagulopathy screen should include:
  - CBC
  - Group and hold
  - Coagulation screen (if APTT is prolonged go on to request Factor VIII/IX levels).

Thalassemia

- Thalassemias are generally inherited in an autosomal recessive fashion and are more common in communities from the Mediterranean, Middle East, South East Asia and the Pacific.
- Hopefully mothers with thalassemia will have received antenatal genetic counselling and the results of the infant’s father’s Hb electrophoresis will be known
- At birth the predominant circulating haemoglobin is Hb F ($\alpha_2\gamma_2$).
- Adult haemoglobin, Hb A ($\alpha_2\beta_2$), becomes the predominant haemoglobin after birth.
- Therefore, infants with $\alpha$-chain abnormalities tend to be symptomatic at birth (or before)
- Those with $\beta$-chain problems develop symptoms usually after 4-6 months.
- The cord blood red cells from infants with either $\alpha$-thalassemia trait or HbH Disease are microcytic, and the level of Hb Bart’s (an abnormal haemoglobin composed of only $\gamma$-chains) in the cord blood is raised

Investigations

- Cord blood for haemoglobin electrophoresis, particularly Hb Bart’s quantitation if $\alpha$-thalassaemia is suspected
- Venous blood for haemoglobinopathy study.
- If $\beta$-globin defect is suspected the specimen should usually be taken at 3 months at the earliest.
- FBC (may be normal in thalassaemia traits)

Beta Thalassemia Minor (trait)

- Mild reduction in beta chain synthesis leading to a reduction in HbA and mild microcytic anaemia
- If both parents have beta thal trait they have a 25% risk of having a child with beta thalassemia major
- If only one parent is affected there is no risk of having a child with beta thal major
- Diagnosis is confirmed by having elevated HbA2 on Hb electrophoresis
Beta Thalassemia Major
- Marked decrease in beta chain synthesis with accumulation and precipitation of alpha chains
- Fetus is unaffected in utero and newborn is haematologically normal
- Symptoms develop in the first 6-12 mths with severe microcytic, hypochromic anaemia, failure to thrive, hepatosplenomegaly and jaundice
- Electrophoresis shows a predominance of HbF with absence or only small amounts of HbA

Alpha Thalassemia – Silent carrier (one alpha gene deleted)
- Clinically and haematologically silent – of no clinical significance to offspring unless partner also affected

Alpha Thalassemia Trait (two alpha genes deleted)
- Causes mild microcytic anaemia
- 5-8% Hb Barts seen on Hb electrophoresis for first 3-6 months but then normal

Hb H Disease (three alpha genes deleted)
- Excess beta chains accumulate and precipitate in red blood cells causing a chronic moderately severe microcytic haemolytic anaemia

Hydrops Fetalis (no alpha chains)
- Causes still birth or immediate post natal death.
The testing programme aims to detect the below diseases for which effective treatment is available. Four dried blood spots are collected on filter paper and analysed for evidence of these diseases. In each case, early detection, before the child has developed symptoms, has the potential to reduce mortality and/or long term morbidity. Since there are about 60,000 births per year in NZ, there are about 30-35 true positive tests. Many babies with these disorders will be symptomatic before the test results become available, therefore, clinical suspicion and appropriate laboratory testing is still essential. The fax and telephone numbers for the National Testing Centre are in the rolodex at the NICU ward clerk’s desk.

If you need results urgently:
- Arrange for the card to be sent urgently to the NTC.
- Call Dianne Webster, the Director of the NTC and explain the urgency.
- If the card has already gone and you need results urgently, call and ask for them, rather than waiting for them to appear.

About 1 in 100 tests needs to be repeated. The most common reasons are due to timing of the first sample, or incorrectly filled blood spots. Most babies with an abnormal test should be evaluated by a doctor, even while awaiting the results of a repeat test. The action needed may involve repeating the test and/or urgent treatment

<table>
<thead>
<tr>
<th>Disorder</th>
<th>Incidence</th>
<th>Test Measures</th>
<th>Why Detect It?</th>
<th>Other Comments</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cystic Fibrosis (autosomal recessive mutation of the CFTR gene)</td>
<td>1 in 3000</td>
<td>Immunoreactive trypsin then PCR for CFTR mutations by PCR</td>
<td>CF causes malabsorption, chest infections, early death. Early detection allows genetic counselling and early treatment (antibiotics, enzyme supplements) delays symptoms, prolongs life.</td>
<td>High false positive rate for IRT. False negatives can occur in babies with minimal or very advanced pancreatic disease at birth.</td>
</tr>
<tr>
<td>Hypothyroidism (mixture of genetic and non-genetic causes)</td>
<td>1 in 4500</td>
<td>TSH</td>
<td>Before screening hypothyroidism in infancy was the leading cause of severe but preventable mental retardation. If treatment is begun soon after birth, later intelligence is normal.</td>
<td>Clinical signs and symptoms are difficult to detect until irreversible brain damage has occurred. NB: Test will miss hypothalamic hypopituitarism and primary hypopituitarism.</td>
</tr>
<tr>
<td>Phenylketonuria - autosomal recessive disorders of amino acid metabolism</td>
<td>1 in 15,000</td>
<td>Phenylalanine</td>
<td>PKU causes autism, mental retardation and seizures, due to build-up of the amino acid phenylalanine. Consequences can largely be prevented by a special (low phenylalanine) diet throughout childhood.</td>
<td>Test should be done after minimum of 2 days of protein feeding. Highest incidence among Irish and Polish (1 in 6000).</td>
</tr>
<tr>
<td>Congenital Adrenal Hyperplasia (autosomal recessive disorders of steroid synthesis)</td>
<td>1 in 20,000</td>
<td>17 OH progesterone</td>
<td>Life-threatening abnormalities of fluid and salt balance, high blood pressure and abnormal sex hormones occur in different variants of CAH. Steroid medications effective.</td>
<td>Premature babies commonly have elevated levels of 17 OH progesterone. They need a repeat test.</td>
</tr>
<tr>
<td>Biotinidase Deficiency (autosomal recessive disorder of biotin synthesis)</td>
<td>1 in 50,000</td>
<td>Biotinidase activity</td>
<td>Biotin is essential for many metabolic pathways in the body. Deficiency causes vomiting, lethargy, coma and often death.</td>
<td>Treatment with biotin results in a good prognosis.</td>
</tr>
<tr>
<td>Galactosemia (autosomal recessive disorders of)</td>
<td>1 in 120000</td>
<td>Galactose and gal-1-PO4</td>
<td>Babies with the condition cannot break down galactose (one of the two types of sugar molecules in</td>
<td>Testing should be done after 2 days of milk feedings.</td>
</tr>
<tr>
<td>Disorder</td>
<td>Incidence</td>
<td>Test Measures</td>
<td>Why Detect It?</td>
<td>Other Comments</td>
</tr>
<tr>
<td>---------------------------------------------------</td>
<td>-----------------</td>
<td>--------------------------------</td>
<td>--------------------------------------------------------------------------------</td>
<td>--------------------------------------------------------------------------------</td>
</tr>
<tr>
<td>Disorder breakdown of galactose</td>
<td></td>
<td></td>
<td>mammalian milk). Build up of galactose causes mental retardation, seizures, liver failure, serious infections, cataracts and death if untreated.</td>
<td>Treatment is a milk free diet (e.g. soy formula in infancy).</td>
</tr>
<tr>
<td>Maple Syrup Urine Disease (MSUD - autosomal recessive disorder of metabolism of branched chain AA's)</td>
<td>1 in 250,000</td>
<td>Branched chain amino acids.</td>
<td>Build up of branched chain amino acids leads to coma, seizures and death, or mental retardation in survivors.</td>
<td>Treatment is with a special diet low in branched chain amino acids.</td>
</tr>
<tr>
<td>Fatty acid oxidation and mitochondrial disorders</td>
<td></td>
<td>Multiple</td>
<td>Associated with SUDI's</td>
<td>Referral / discussion with metabolic consultant Starship</td>
</tr>
<tr>
<td>SCID (Severe Combined Immune Deficiency)</td>
<td>1 in 50,000</td>
<td>Levels of TREC (T-cell receptor excision circles)</td>
<td>Rare disorder caused by a deficiency or absence of T cells. Risk of recurrent life threatening infections and death in the first year if untreated.</td>
<td>Can be managed with early antibiotics and isolation and consideration of stem cell transplant if the condition is diagnosed. This improves survival and health outcomes</td>
</tr>
</tbody>
</table>

**Investigation of Suspected Cystic Fibrosis**

- Immunoreactive trypsin levels are checked as part of the Guthrie screen.
- The IRT level is high at birth in carriers and most patients with CF but declines after the newborn period.
- The current process following a positive IRT result is as follows.

<table>
<thead>
<tr>
<th>Positive Guthrie – IRT in highest 1% level</th>
<th>CF genetic screen undertaken from the Guthrie for three common mutations.</th>
<th>If 1 or 2 mutations are found the LMC and CF team are notified to inform family and arrange review</th>
<th>Clinical assessment + sweat test + extended gene probe + pancreatic function + family carrier screening</th>
<th>Carrier or possible CF or definite CF – depending on sweat test and genetic results</th>
</tr>
</thead>
<tbody>
<tr>
<td>3 mutation screen – at least 1 present in 98% of NZ CF patients therefore 2% missed</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Screening - 8% false negative rate (a low IRT in those with minimal pancreatic insufficiency and no CFTR genetic mutations detected)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Screening - 92% false positive rate from high IRT result (75% of those with a high IRT will be carriers only)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Infants with meconium ileus should have CF genetic mutation analysis and a sweat test organized regardless of the newborn screen result as they often can have a low IRT and be missed on the newborn screen.</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Most useful screen for pancreatic insufficiency (PI) is faecal elastase. &gt;200 is normal, 100-200 is moderate PI and &lt;100 is severe PI.</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>In cases where pancreatic insufficiency is suspected infants require supplementation with pancreatic enzymes.</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Infants who are &lt; 37 weeks or &lt; 2.5kg receive Pancrease while term infants &gt; 2.5 kg receive Creon. There is detailed information regarding the use of these supplements in the NICU drug folder.</td>
<td></td>
<td></td>
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</tr>
</tbody>
</table>
Investigation of Neonates with a Family History of Metabolic Conditions

In these situations it may be necessary to investigate the baby earlier on rather than waiting for a Guthrie result to be available. If there are any concerns talk to the Neonatal Consultant who may need to discuss testing with the Metabolic Specialists or other Specialists at Starship.

- Cystic Fibrosis – take blood for CF gene testing at the time of the Guthrie test, or earlier if parental concern
- Hypothyroidism – take Guthrie test as usual at 48 hours and only do further TFT’s if Guthrie is abnormal
- Phenylketonuria – take blood for amino acids on day 1
- Congenital Adrenal Hyperplasia – take blood for 17-hydroxyprogesterone on day 1
- Galactosaemia – take Guthrie at 48 hours of age but comment on the card that a sibling has galactosaemia to ensure an enzyme assay is also done
- Biotinidase deficiency – take Guthrie at 48 hours, no other specific tests required
- Maple Syrup Urine Disease – take Guthrie at 48 hours of age, need to wait until then for baby to become catabolic
- Other metabolic conditions need to be individually discussed with the Metabolic Team to decide on the most appropriate timing of tests. Often if there is a family history a plan will have been made prior to birth. An example is MCAD and Auckland have an extensive guideline on this.
Ordering the most appropriate investigation to diagnose a potential genetic abnormality is a rapidly changing field. If in doubt ask.

Referrals to the genetic service in Christchurch is by email to genetics.servicenz@cdhb.health.nz

Antenatal Investigation

In pregnancies where there have been concerns about possible genetic diagnoses there should be letters under the mother's NHI. If they have seen a neonatal paediatrician there will be a letter or notes on health connect as well as in the neonatal anomaly folder in reception.

The radiology investigations should be reviewed and these include the screening ultrasound scans as well as specific investigations such as MRI.

The two approaches to genetic investigations are classified as:

- Non Invasive Prenatal (genetic) Screening Testing or NIPS
  - This is a screening test and possible diagnoses should be confirmed with a karyotype or microarray after birth.
- Invasive (amniocentesis or chorionic villous sampling)
  - This is a diagnostic test and does not require repeating after birth.
  - Amniocentesis results can be filed under laboratory or letters if done outside Christchurch.

Following birth tests include:

- Fluorescent In Situ Hybridisation (FISH) (1 ml Green top)
  - This is a preliminary test that is able to look for extra genetic material (such as when there is an extra chromosome in Trisomy 21 / 18 or 13), or absent material (eg 22q deletion). Definitive tests are always needed to confirm the results.
  - Please discuss with the SMO before giving parents any potential results, including provisional results.
  - These results must be confirmed using a karyotype or microarray.
- Karyotype (1 ml green tube)
  - This refers to microscopic examination of the numbers of chromosomes and their appearance. It picks up aneuploidy (abnormal numbers of chromosomes (eg Trisomy or Monosomy) as well as observable deletions / translocations and duplications. The report includes % cells affected and whether likely mosaic.
  - It is recommended that we wait until the report is signed off in the lab before giving parents the results.
- Microarray (2 full EDTA + Green)
  - This refers to a more detailed examination of chromosomes where the chromosomes are examined as small sections of DNA material rather than as whole chromosomes. This is now the usual first line test where a range of indeterminable dysmorphic features are present. Decision is by the lead or ward round SMO. Consent from the parents is required. There is a patient information sheet available – Consent and information for parents about microarray analysis.
  - The result takes 5-10 working days. Don't over promise the timing of a result.
  - If a microarray was taken antenatally, a repeat postnatal is not required. The result needs to go under the baby's NHI (a work in progress to achieve).
  - Always discuss the finding with the lead SMO. If unavailable contact the consultant of the week.
  - A referral to the geneticist is usually made and they will visit once the result is through. If the baby's condition is critical inform them so that they can advise on additional tests.
- Specific Genetic Studies
  - If a baby has been reviewed by a Geneticist they may request specific studies looking for a known genetic mutation.
- Whole blood exome testing
  - Whole blood genome testing is arranged by genetics in consultation with the lead SMO, and occasionally needs additional approval for high cost tests from the CD.
- Sample for stored DNA, for later use.
  - (Pink/EDTA)
  - Suggest this is taken ASAP if a metabolic or genetic condition is suspected and survival not expected. Often requested by the geneticist (when they are unable to determine a condition clinically).
## GENETIC TESTING REQUEST FORM

<table>
<thead>
<tr>
<th>Surname</th>
<th>Given Names</th>
<th>Extra copy to</th>
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<tr>
<th>Location</th>
<th>Consultant</th>
<th>Charge to (HIC Code)</th>
<th>TAKEN BY</th>
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</thead>
</table>

Canterbury Health Laboratories will rely on the requestor to obtain informed consent for the requested tests, and any additional related tests, to be performed by the laboratory.

**Clinical details:**

- Sendaway shipping instructions:
  - [ ] Hold pending sendaway form

Two independent samples required? [ ]

<table>
<thead>
<tr>
<th>Microarray</th>
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<th>DMD</th>
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<td>Karyotype</td>
<td>Hereditary deafness (Cx26 &amp; 30)</td>
<td>Fragile X</td>
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<tr>
<td>FISH</td>
<td>Mitochondrial point mutations</td>
<td>BRCA 1/2</td>
</tr>
<tr>
<td>DNA (Extraction and store)</td>
<td>Hereditary pancreatitis</td>
<td>Long QT Syndrome</td>
</tr>
<tr>
<td>Other (please specify condition or gene):</td>
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</table>

Blood tube collection key:  
- Lithium heparin [ ]
- EDTA [ ]
**NEUROLOGY**

**Hypoxic Ischaemic Encephalopathy**

- Evidence is accumulating that whole body cooling started soon after birth in infants with hypoxic ischaemic encephalopathy (HIE) will improve survival free of major sensorineural disability.
- Intrapartum hypoxia affects 3 - 5 per 1,000 live births with subsequent moderate or severe HIE in 0.5 -1 per 1,000 live births.
- Of these, between 10-60% die and at least 25% of the survivors have neurodevelopment sequelae.
- Hypothermia is the most promising, clinically feasible manoeuvre to provide neurological rescue for neonates with HIE.

**Pathogenesis**

Following a reversible hypoxic ischaemic global insult, neuronal death occurs in two major phases, each characterised by distinct pathophysiological processes influenced by the nature and severity of the insult.

- **Primary neuronal loss** is related to cellular hypoxia, which leads to exhaustion of high-energy metabolism (primary energy failure) and cellular depolarisation. There may be immediate neuronal death if the insult is severe, however, many neurons do not die during this primary phase of injury.
- **Secondary neuronal loss** occurs due to a cascade of pathologic processes and delayed neuronal death is initiated leading to further loss of neurons starting hours after the initial insult and extending over days. This accounts for a significant proportion of final cell loss. The mechanisms involved include hyperaemia, cytotoxic oedema, mitochondrial failure, accumulation of excitotoxins, active cell death (analogous to developmental apoptosis), NO synthesis and cytotoxic actions of activated microglia. The delayed phase is associated with increased seizure activity and marked encephalopathy. This was predictive of both mortality and neurodevelopmental outcome at both one and four years of age.

Therefore a therapeutic ‘window of opportunity’ exists in the interval following resuscitation of the asphyxiated newborn, before the secondary phase of impaired energy metabolism and injury. From animal experimental data it appears that the earlier hypothermia is commenced the better the outcome. It must be commenced before 6 hours to be of any benefit.

**Predictors of Adverse Outcome**

- Moderate or severe encephalopathy appears to be the most sensitive single predictor of adverse outcome, with a 44% risk of death and/or disability.
- An Apgar score of ≤3 at 10 minutes has been associated with a 23% risk of death or severe disability.
- A base deficit of >12 mmol/L was associated with a 39% risk of adverse neurological outcome at 1 year.
- Combinations of perinatal events have also been used - a 5 minute Apgar score of ≤5 in association with encephalopathy and seizures has been associated with a 70% risk of death or disability.
- The triad of delivery room intubation with a 5 minute Apgar score of ≤5 and a cord pH of ≤7 and/or postnatal base deficit of ≥14 or more combined with early moderate or severe encephalopathy at 3 hours age predicted 83% of deaths or abnormal neurological examinations at discharge.
- The combination of encephalopathy and low Apgar scores and/or significant acidaemia will predict 30-80% of deaths or significant disability in survivors.

**Current Evidence for Outcome after Cooling**

The Cochrane systematic review of the effect of systemic cooling on outcome was updated in January 2013. Eight randomised controlled trials of 638 term infants with moderate/severe encephalopathy and evidence of intrapartum asphyxia were reviewed.

- Therapeutic hypothermia resulted in a statistically significant and clinically important reduction in the combined outcome of mortality or major neurodevelopmental disability to 18 months of age (typical RR 0.76, 95% CI 0.65, 0.89; typical RD -0.15, 95% CI -0.24, -0.07; NNT 7, 95% CI 4,14).
- Cooling resulted in statistically significant reductions in mortality (typical RR 0.74, 95% CI 0.58, 0.94; typical RD -0.09, 95% CI -0.16, -0.02; NNT 11, 95% CI 6,50) and in
- Cooling resulted in statistically significant reductions in neurodevelopmental disability in survivors (typical RR 0.68, 95% CI 0.51, 0.92; typical RD -0.13, 95% CI -0.23, -0.03; NNT 8, 95% CI 4,33).
- Some adverse effects of hypothermia included an increase in the need for inotrope support of borderline significance and a significant increase in thrombocytopenia.
They concluded as have 2 other meta-analyses by Shah and Schulzke that therapeutic hypothermia is beneficial to term infants with moderate and severe HIE, with reduced mortality without increase in major disability in the survivors. The benefits outweigh the short term adverse effects. Further trial results are expected and will provide additional information.

Adverse Effects of Hypothermia
- Mild hypothermia appears to be well tolerated.
- There have been no reported serious adverse effects in the three pilot studies of hypothermia in human newborns.
- Adverse effects such as sinus bradycardia, increased blood pressure and increased oxygen requirement were all transient and reversible with re-warming (when the rectal temperature remained within 33.0°C – 34.0°C, these were less likely to occur).
- There has been a median increase in FiO₂ of 10–15% reported in infants with severe respiratory failure (oxygen requirement greater than 80%) up to 6 hours of age and because of this they are not eligible for cooling.
- Increase in the rates of thrombocytopenia

Cerebral Function Monitor (Brainz Monitor)
- Amplitude-integrated EEG recordings with a “cerebral function monitor”, obtained continuously from two biparietal electrodes, have been shown to be useful in the early prediction of the severity of brain injury.
- The Brainz monitor should be placed early on any baby with HIE or encephalopathy.
- A daily report should be written in the patient’s notes about the background brain activity and presence of seizures.

We acknowledge that much of the content of this information has been obtained from the ICE study. For references see the information sheets in the Hypothermia box in Room 1 Level 3.

Hypothermia Protocol

Inclusion Criteria
- Infants of 35 weeks gestation or more
- Evidence of moderate or severe encephalopathy (Sarnat 2 or 3)
- Evidence of intrapartum hypoxia, at least two of:
  - Apgar score of ≤5 at 10 minutes
  - Mechanical ventilation, CPAP or ongoing resuscitation at 10 minutes
  - Cord pH <7.00, or
  - Arterial pH <7.00 or base deficit of ≥12 within 60 minutes of birth

Exclusion Criteria
- Cooling cannot be started within 6 hours of birth
- Birth weight less than 1.8 kg
- Major congenital abnormalities
- Severe hypotension, pulmonary hypertension or coagulopathy unresponsive to treatment
- Infants requiring inspired oxygen >80%
- Infants in extremis and not expected to survive

Cord Blood Gases
- Should be requested for any delivery where the baby requires resuscitation of (> 2 minutes duration.)

Placental Histology
- Should be requested for any baby admitted with Apgar <6 at 5 mins or who you think may be considered for cooling.

Documentation
- Precooling assessment sheet is to be filled in by the clinician making the decision to cool
- This can be found in the Neonatal Handbook folder under “other guidelines” and can be used as an aide to help deciding whether to cool or not
- This form also documents relevant information such as timing of cooling, grade of HIE, æEEG findings, dates for follow-up MRI and developmental exam
Sarnat Stages for HIE

Always discuss with a consultant when a baby meets criteria for HIE as Sarnat scoring can be difficult.

Stage 1: Hyperalertness, hyper-reflexia, dilated pupils, tachycardia, seizures uncommon

Stage 2: Lethargy, hyper-reflexia, miosis, bradycardia, seizures, abnormal tone, weak suck and Moro reflex. EEG shows abnormal background activity

Stage 3: Stupor, flaccidity, small to mid-position pupils which react poorly to light, decreased stretch reflexes, hypothermia and absent Moro, seizures often absent but may have decerebrate posturing

Management of Babies Commenced on the Hypothermia Protocol

A detailed discussion between the parents and consultant must occur prior to commencement of cooling.

Cooling can occur on a retrieval as long as the case has been discussed with the consultant and the management deemed appropriate for that baby. The cooling equipment would need to be taken out on the retrieval so discussion with the consultant before departing would be prudent to assess whether cooling might be an option for the baby.

All infants should have venous and arterial catheters inserted.

Cooling protocol
- All infants will be nursed on an overhead.
- Core temperature to be lowered to 33.0°C–34.0°C.
- Temperature will be measured continuously by a thermistor inserted 5 cm into the rectum.
- Cooling will be started and continued for 72 hours.
- Hypothermia will be achieved primarily by turning the radiant warmer off and exposing the infant to ambient temperature.
- Cooling should preferentially be done with the Meditherm mattress which has a feedback mechanism from the baby's rectal temperature and the cooling machine.
- Cooling can also be achieved by using cool packs of around 10°C be applied to the back of the neck and head, and across the torso. Active cooling will be reduced when the rectal temperature falls below 34.5°C and stopped when below 34.0°C. If the temperature falls below 33.5°C, the heater output on the radiant warmer will be manually adjusted to maintain the target rectal temperature at around 33.5°C.
- Active cooling will be reduced if the inspired oxygen increases by more than 20%
- Cooling will be stopped if there is:
  - persistent hypoxaemia in 100% oxygen
  - life threatening coagulopathy
  - an arrhythmia requiring medical treatment (not sinus bradycardia)
  - parents withdraw consent to continue hypothermia
- After 72 hours, rewarming will occur at a rate not exceeding 0.5°C every 2 hours and stopped when the core temperature is 37 degrees

Monitoring protocol
For 72 hours infants will be carefully monitored by:
- continuous invasive blood pressure measurements
- oxygen saturation
- heart rate
- respiration rate
- urine output
- ECG

Investigations
The following will be done at least daily, as discussed on the ward round, and more frequently if abnormal:
- blood gases and lactate
- glucose
- electrolytes, urea, creatinine, calcium, magnesium (NEON)
- liver function tests
- full blood count
- coagulation profile
Neonatal Seizures

Notify the consultant of the admission of any baby with suspected or proven seizures. There are multiple possible causes, including:

- Birth asphyxia
- Intracranial haemorrhage eg: IVH, intracerebral, subarachnoid, subdural,
- Metabolic disturbance
  - Hypoglycemia
  - Hypocalcemia
  - Hypo/hypernatremia
  - Hypomagnesemia
  - Hyperammonemia
  - Secondary to inborn errors of metabolism
- Infection
- Viral eg. HSV
- Bacterial
- Protozoal eg. toxoplasmosis
- Anatomical abnormalities
- Intoxication (e.g. inadvertent injection of local anaesthetic into fetus)
- Drug withdrawal

Presentation

This may be quite subtle e.g. dusky spells, apnoea, or with obvious clonic or tonic-clonic movements.

The following table is from the ADHB guidelines.

<table>
<thead>
<tr>
<th>Type</th>
<th>Physical Features</th>
<th>EEG</th>
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</table>
| Subtle| • Eye signs - eyelid fluttering, eye deviation, fixed open stare, blinking  
  • Apnoea  
  • Cycling, boxing, stepping, swimming movements of limbs  
  • Mouthing, chewing, lip-smacking, smiling  
|       | • Often no EEG changes.  
  • EEG changes most likely to occur with ocular manifestations |
| Tonic | • Stiffening  
  • Decerebrate posturing  
|       | • EEG variable |
| Clonic| • Repetitive jerking, distinct from jittering  
  • Can be unifocal or multifocal  
|       | • Usually changes identified |
| Myoclonic| • Rare  
  • Sleep myoclonus is benign  
|       | • EEG often normal  
  • Background EEG can be abnormal |

Investigations

- FBC,Electrolytes, Blood glucose
- Blood cultures, urine cultures, CSF (don’t forget contraindications). Send sufficient specimen for viral studies, including HSV PCR. It may be helpful to send an extra tube of CSF and ask the lab to freeze it, especially if an infectious or metabolic cause is suspected.
- Throat swab, stool specimen for virology
- Aspirate and swab any skin lessons. Discuss with virology. In general they will send a vial of viral transport medium. A swab tip can be broken into this.
- Immunofluorescence specimens are collected by scraping the base of a vesicle and smearing the material onto a glass slide (supplied from virology).This slide should be transported in a rigid plastic container.
- Blood gases
- CXR, cranial ultrasound, urgent CT if suspicion of a subdural haemorrhage.
- Investigation of suspected inborn error as above.
• EEG / BRAINZ monitor
  - It is not necessary to defer therapy until an EEG can be obtained.
  - EEG may assist in confirming that subtle phenomena are seizures or to determine if a paralysed infant is having seizures.
  - The interictal EEG (full 16 lead) may be useful in estimating prognosis particularly in HIE.

Diagnosis and Management
• Involves a full history and examination particularly with reference to the above aetiologies.
• Management directed to the cause if known.
• Avoid hyperthermia – see protocol. Maintain temperature within 35.5 to 36.5 °C
• Ventilation if apnoea or hypoventilation secondary to seizures or anticonvulsant therapy.

Anticonvulsants
• Phenobarbitone; Give 20 mg/kg initially, then further 10 mg/kg doses can be given if seizures recur. We often use up to a total dose of 40 mg/kg, (doses up to 50−60mg/kg have been used discuss with a consultant, ventilation for apnoea is likely to be necessary). If maintenance therapy is required (where there is a high risk of recurrent seizures) the dose is 3-6 mg/kg/day given at least 12 hrs after the load
• If seizures not controlled discuss use of the following with a consultant;
  - Phenytoin. 15mg/kg IV slow infusion, further 5mg/kg doses may be given up to 25mg/kg. IV Maintenance therapy may be used initially but continuing with maintenance phenytoin is uncommon due to the erratic levels achieved with oral solutions.
  - Midazolam 0.1 mg/kg IV (see drug protocol) IV bolus, can be followed by an infusion if fits persist.
  - Levitracetam is a second line anticonvulsant that can be used in consultation with the Paediatric Neurologists. A loading dose of 30mg/kg is required followed by maintenance starting at 10mg/kg 12-24 hourly.
  - Paraldehyde 0.1 mg/kg deep IM injection or rectally. (need to mix with oil if given PR to avoid proctitis and risk of sterile abscess given IM) Paraldehyde can be a very useful drug, but as it is excreted (and can be absorbed) via the lungs, for reasons of safety and comfort of staff, its use should be limited
  - Consider pyridoxine deficiency in intractable seizures. Give pyridoxine 50 mg IV. Should be done with EEG monitoring. Risk of apnoea.
• Drug levels should be monitored when maintenance therapy is thought to be necessary.

Brainz Monitor
• The BRAINZ monitor is a type of cerebral function monitor but does not give the same information as a standard 16 lead EEG.
• It does have the advantage of easy use in a neonatal intensive care situation and has been proven to be of use in the detection of neonatal seizures and significant cerebral dysfunction.

Indications
The following infants should be considered for BRAINZ BRM2 monitoring.
• Infants ≥ 34/40 with:
  - Definite or possible seizures
  - Muscle relaxed infants at risk of seizures that may not be clinically apparent
  - Moderate perinatal asphyxia (Neonatal encephalopathy, Sarnat 2 or 3)
  - Unexplained apnoea
• Monitoring on other babies should not be commenced unless directed by a specialist.

Application
• Sensors (needles preferred, or electrodes if still stocked) to be applied by Newborn Medical and Nursing staff who have undertaken BRAINZ BRM2 training.
• See instructions on monitor and make sure to enter the correct date and time
• Needles
  - Clean the skin with 2% chlorhexidine and part the hair
  - Insert needles in the appropriate spots, attach the reference sensor on to the back
  - Dry the area around the needles and secure with a steristrip
• Electrodes – the key to good application is patience and a stepwise approach
- Ensure the purple end has straight (not bent) connecting “teeth”
- Warm the sensors in the incubator or under the radiant warmer
- Use the ruler provided to ascertain where the sensors should be placed
- Place them around any injury ie: cephalhaematoma and mirror the position on the other side
- Prepare the skin by parting the hair and rubbing a small amount of abrasive gel 10-15 times over a larger area than is required to fit the sensor on
- Place the sensor in the correct spot and hold it there firmly for 30 seconds to allow the hydrogel to warm and bond with the skin
- Only add water form the outside of the sensor if needed if the hydrogel dries out

- See basic EEG atlas pictures of BRAINZ abnormalities on G: drive / brainz.

Documentation
- Assessment of the background activity, presence of seizures and R/L asymmetry are important to note.
- There are stickers to put in the notes (in the BRAINZ box) whenever a baby is being monitored.

Intraventricular Haemorrhage

- Incidence in babies <1500gm is up to 25%, and is highest in gestation < 26 weeks
- Caused by bleeding into the subependymal germinal matrix ± extension into the ventricle or parenchyma
- The bleeding occurs mainly in the first few days after birth and can be detected by head US:

  Day 1 – 50% detected
  Day 2 – 75% detected
  Day 3 – 90% detected

  Grade 1 - Bleeding limited to the germinal matrix
  Grade 2 - Blood extending into the ventricle but not distending the ventricle
  Grade 3 - Blood extending into the ventricle and filling (over 50%) and distending the ventricle with blood, ventricular dilatation is present.
  Grade 4 - Parenchymal bleeding with or without blood in the ventricle

Risk Factors
- Lower gestational age
- Lack of antenatal steroids
- Unstable haemodynamics
- PDA
- Sepsis

Monitoring
- Head US scans on day 2-3, 7 and at, 6 weeks in all <1500gm and <32 week gestation babies
- Head US scans on day 14 and 28 as well if significant abnormalities
- See checklist for babies <1500gm

Post Haemorrhagic Ventricular Dilatation

Definition
- Posthaemorrhagic ventricular dilatation (PHVD) is defined as progressive ventriculomegaly after an intraventricular haemorrhage (IVH).
- The ventricular width at the intraventricular foramen exceeds 4mm over the 97th centile for gestational age (see attached charts in the Neonatal Handbook Folder – Levene index).
- The overall risk of PHVD after an IVH is about 25%. However, the risk increases with the severity of the IVH: (Arch Dis Child Fetal Neonatal Ed 2002;87:F37-41)

  Grade 1 IVH 4%
  Grade 2 IVH 12%
  Grade 3 IVH 74%
Pathogenesis

- CSF is produced by the choroid plexus and reabsorbed by the arachnoid villi and across the ependyma into small blood vessels. After an IVH there are multiple small blood clots that prevent CSF reabsorption. Plasminogen in the CSF attempts to fibrinolyse the blood but is often ineffective.
- Over time transforming growth factor β1 (TGFβ1) is released into the CSF and causes deposition of extracellular matrix proteins and this can further inhibit reabsorption leading to communicating hydrocephalus.
- The extracellular proteins can also cause obstructive hydrocephalus by blocking the exit to the fourth ventricle.

Management

- Infants with progressive ventricular dilatation are managed with the aim of preventing secondary damage due to raised intracranial pressure and avoiding the need for a permanent shunt if at all possible.
- About 50% of infants will show resolution of their dilatation and not require surgery.
- Studies have reviewed a number of non-surgical treatment modalities and they have been shown to be ineffective or high risk procedures - these include repeated lumbar punctures or ventricular taps, diuretic therapy with acetazolamide and frusemide and intraventricular fibrinolytic therapy.

Surgery

- Surgical interventions for PHVD are the mainstay of treatment and are needed if the ventricular dilatation progresses and is associated with raised intracranial pressure.
- About 30% of infants with PHVD will need surgery.
- External ventricular drains can be placed subcutaneously in the scalp with a catheter in the ventricle and allow easy access for repeated drainage of CSF to remove blood and protein and to normalise intracranial pressure.
- The risks include infection, skin breakdown and blockage with blood clots.
- This is usually a temporary measure and be all that is required if the ventricular dilatation stops progressing.
- A permanent ventriculoperitoneal (VP) shunt is needed if the ventricular dilatation is unremitting.
- These can not be placed early as there is a high risk of blockage if the protein component of the CSF is >1.5g/L.
- VP shunts carry a high risk of infection and blockage.

Outcome

- The outcome after PVHD is poor.
- About 20% of infants will die and those that survive have high rates of neurodevelopmental disability.
Hypoglycaemia

Definition
- Data suggests that there may be sequelae from blood sugar values < 2.6 mmol/L (Lucas 1988), however, there is no clear consensus as to what threshold to use in treatment of hypoglycaemia. In a recent survey of all units in the Australia New Zealand Neonatal Network < 2.6 mmol/L was used in all units and is the cut off used in a New Zealand collaboration of clinical practice guidelines on treatment of neonatal hypoglycaemia with oral dextrose gel.
- If any baby shows symptoms that could be due to hypoglycaemia, a blood sugar level should be measured immediately.

Background
- Healthy term infants are able to mobilise energy stores through a process known as counter regulation and are unlikely to suffer any ill effects if fed infrequently in the first 24-48 hrs. Some babies however are less able to mount this response and these babies are at greater risk of hypoglycaemia.
- Regular assessment and documentation of all infants should occur including assessment of feeding regardless of risk factors. All assessments should be documented on the neonatal observation chart if on the postnatal ward.
- Healthy term infants do not require routine monitoring of blood sugars but at risk babies need to be identified and monitored accordingly.
- Hypoglycaemia may be asymptomatic or symptomatic, but both can result in adverse outcome such as brain injury, neurodevelopmental delay and death.
- Symptoms are wide ranging and include poor feeding, altered level of consciousness, cyanosis, jitteriness, seizures, apnoea, tachypnoea, irritability, hypotonia, sleepiness and floppiness.
- Hypoglycaemia needs to be suspected or prevented and actively investigated and treated in any unwell baby.
- Please be aware that colostrum and milk mixture are not equivalent. Give whatever colostrum is available and recheck before considering infant formula. Women who have collected colostrum antenatally will have a limited resource available.
- Dextrose gel has been shown to be better than feeding alone to reverse neonatal hypoglycaemia in babies from 35 weeks gestation and under 48 hours of age. This in turn reduces maternal infant separation by reduced admission to the Neonatal Unit and encourages the establishment of breast feeding.

Infants at high risk of hypoglycaemia
- Preterm < 37 weeks
- Small for gestational age <9th percentile (on UK-WHO growth chart ref 6505)
- Macrosomic babies >98th percentile (on UK-WHO growth chart ref 6505)
- Baby of a mother with diabetes
- Hypothermic infants
- Severe intrapartum fetal distress or cord lactate > 5.8mmol/L.
- Asymmetric growth (weight centile more than 2 centile lines lower than head and length centile) in conjunction with another risk factor e.g. intrapartum fetal distress or meconium
- Unwell infants
- Sepsis
- Ultrasound abnormalities in utero such as agenesis of the corpus callosus, septic-optic dysphasia or absent cavum septum pellucidum (See Antenatal Ultrasound Abnormalities)
- Infants diagnosed or suspected of Beckwith Weideman Syndrome

Transient Hypoglycaemia
- Decreased glucose production or increased utilisation
  - Asphyxia, starvation, sepsis, congenital heart disease, hypothermia, SGA
- Transient hyperinsulinism
  - Infant of a diabetic mother, LGA, SGA, rhesus haemolytic disease, Beckwith-Weideman syndrome.
Persistent or Severe Hypoglycaemia

- Hyperinsulinism
  - Congenital hyperinsulinism, pancreatic adenoma, leucine sensitivity.
- Decreased production of glucose
  - Glucagon deficiency, congenital hypopituitarism, cortisol deficiency (CAH), inborn errors of metabolism.

RED FLAG: A baby with severe or persistent hypoglycaemia without a clear common cause such as, a baby of a diabetic mother, or being growth restricted should be an alert to a less common cause which could be endocrine or metabolic in origin

Management of Hypoglycaemia on Postnatal Wards (BSL <2.6 mmol/L)

Indication to Call Neonatal Team Immediately

- Apnoea
- Cyanosis
- Altered level of consciousness
- Seizures
- Abnormal tone
- Blood glucose <2.0mmol/L

Management

- Care to be provided on the postnatal ward unless the baby is symptomatic in which case refer and transfer to the neonatal unit (see flow chart below for babies ≥35 weeks gestation within 48 hours of birth).
- Infants at risk of hypoglycaemia should be fed as soon as possible, preferably within the first hour and then fed at least 3 hourly
- The first blood sugar should be checked pre-feed 3-4 hours after birth (this is to avoid the natural nadir in BSL levels) and should continue to have 3 hourly feeds until a total of 3 consecutive pre-feed measurements are ≥2.6mmol/L
- If any recording of BSL <2.6 has occurred, feed the baby at least every 3 hours with blood sugars prior to feeds until there have been 3 subsequent and consecutive BSL’s ≥2.6 and the baby is feeding well without supplemental feeds
- If a baby has not had low sugars but changes from breastfeeds with top-ups to fully breastfeeding a pre-feed BSL is recommended
- The Accu-chek monitors is used for blood sugar measurement and is accurate at low levels. If there is any concern regarding the result use the blood gas analyser as it is more reliable. The Maternity Clinical Coordinators are able to process blood samples through the blood gas analyser to confirm hypoglycaemia prior to giving iv glucose. Do not delay treatment.
- Give colostrum and recheck BSL before considering the use of formula
- If there are concerns about the adequacy of the first feed or concerns about milk transfer then discuss with the parents the option of EBM, harvested colostrum, pasteurised donor breast milk or infant formula
- Prior to any administration of infant formula, ensure mothers have had lactation support including LMC/Midwife or lactation consultant and are regularly expressing. Ideally babies should be skin to skin with their mother whilst feeding by any method
- If formula is indicated give 5-10ml/kg per feed and assess response
- Hypoglycaemia usually resolves in the first 24-48 hours depending on the cause
- The desired outcome from this treatment is that the blood glucose is restored quickly to the normal range without disruption to the establishment of breast feeding and maternal-infant bonding.
- Babies at risk of hypoglycaemia can discontinue blood glucose measurements when they are feeding well without the use of dextrose gel, and without additional supplementary feeds or top ups AND have three consecutive pre-feed blood glucose concentrations ≥ 2.6mmol/L

Dextrose Gel

- To be used only in the first 48 hours after birth for those babies ≥35 weeks gestation
- 40% dextrose gel solution
- Dose is 200mg/kg (0.5mL/kg), prescribed by the neonatal team. Verbal order is allowed if there are delays in prescribing on the drug chart
- Use oral single use syringes to withdraw doses. See Neonatal Drug profile for further details.
• Use non-sterile latex free gloves, dry the baby's inner cheek with sterile gauze and then massage the dextrose gel into the buccal mucosa.
• Administration of dextrose gel should be at least 30min apart
• Maximum of 6 doses of dextrose gel in 48 hours

**Referral Criteria for Neonatal Assessment**

• If blood glucose is below 2.0 mmol/L at any stage to consider NICU admission
• If blood glucose is below 2.6 mmol/L after the second or third feed despite dextrose gel administration
• Any clinical signs of hypoglycaemia
• If more than 4 doses of dextrose gel are required in the first 48 hours
• Request neonatal daily review for babies who develop hypoglycaemia (2.0-2.5mmol/L) even if the post-feed BSL improves according to the feeding protocol
• If the clinical picture is of significant concern that may indicate the baby is unwell regardless of the blood glucose level
Babies at Risk of Hypoglycaemia Flowsheet

NOTE: Oral dextrose gel is used to treat neonatal hypoglycaemia ≥ 35 weeks and < 48 hours after birth

FEED IN FIRST HOUR THEN AT LEAST 3 HOURLY, KEEP WARM INCLUDING SKIN TO SKIN

Blood glucose before second feed and not before 3 hours age

- Blood glucose ≥ 2.6 mmol/L
  - Breastfeed at least 3 hourly
  - Blood glucose 3 hourly before feeds until 3 consecutive results are 2.6 mmol/L or above

- Blood glucose 2.0 to 2.5 mmol/L
  - Immediate neonatal review
  - Give oral dextrose gel 0.5 mL/kg (verbal order if needed)
  - Ensure baby is fed, ideally breastmilk
  - Further feeding advice after neonatal review

- Blood glucose < 2.0 mmol/L
  - Neonatal review within 30 minutes
  - Ensure baby is fed, ideally breastmilk. Formula may be indicated – 5-10 mLs/kg

Recheck blood glucose 30 minutes after dextrose gel administration

- Blood glucose ≥ 2.6 mmol/L
  - Feed at least 3 hourly – ideally breastmilk
  - Blood glucose 3 hourly before feeds until 3 consecutive results are 2.6 mmol/L without top-ups or dextrose gel

- Blood glucose 2.0 to 2.5 mmol/L
  - Neonatal review within 30 minutes
  - Ensure baby is fed, ideally breastmilk. Formula may be indicated – 5-10 mLs/kg

- Blood glucose < 2.0 mmol/L
  - Neonatal review
  - Admit to NICU

If recurrent hypoglycaemia reconsider oral dextrose gel

Neonatal review if:
- More than 4 doses of oral dextrose gel required in 48 hours
- The clinical picture is of significant concern regardless of blood glucose

PRACTICE POINTS
- If any concerns regarding the Accu-check monitor blood glucose result, use the blood gas analyser
- If the feeding regime changes (ie. from breastfeeds with top ups, to fully breastfeeding) a blood glucose measurement is recommended before the next breastfeed
Management of Hypoglycaemia in NICU

- If care on the postnatal ward cannot maintain the BSL’s or baby is symptomatic admit to NICU
- Recheck the BSL on the gas analyser to confirm hypoglycaemia before making a management plan as glucometers are less accurate at lower blood glucose levels
- If severe hypoglycaemia and iv access is difficult or delayed consider using an im glucagon bolus (200 mcg/kg) to elevate the blood sugar whilst working on iv access as this can elevate the BSL for up to 2 hours

BSL <1.5mmol/L or symptomatic hypoglycaemia

- Immediate intervention is required with iv therapy
- If able, take the bloods for a hypoglycaemia screen at iv insertion but do not delay treatment if there is difficulty in obtaining the samples
- Give 2ml/kg 10% dextrose at 2mls/min - be aware of rebound hypoglycaemia
- A bolus must be followed by a continuous infusion of dextrose
  - If IUGR <9th% start at 90ml/kg/day
  - If normally grown start at 65ml/kg/day and increase up to 90ml/kg/day if the subsequent BSL remains < 2.6mmol/L
- If already on a dextrose infusion then the rate or concentration of the infusion needs to be increased after a subsequent bolus
- Recheck the BSL in 30 minutes
- Total fluid volume in day 1 should be limited to 100ml/kg/day otherwise issues arise with hyponatraemia from free water overload. Other options include increasing the dextrose concentration from 10% to 12.5% to 15% to maintain the BSL. Central iv access is required if concentration is ≥12.5% but a peripheral iv with 12.5% can be used temporarily until central access is obtained
- Calculate the glucose load in mg/kg/min with each alteration in fluid rate or concentration (Click Here)
- If manipulating dextrose concentrations and rates > 10 mg/kg/min are not sufficient to maintain BSL’s then additional pharmacological management may be required after consultation with Endocrinology and a cause for the hypoglycaemia found
- Medications may include – glucagon infusion, diazoxide, hydrocortisone, octreotide
- To discuss on ward rounds with the SMO when it is safe to start weaning iv fluids – usually when BSL’s have been stable for 12 hours and there is no feed intolerance
- Gradually wean and overlap to enteral feeds – be aware of the severity and persistence of hypoglycaemia to guide the rate at which to wean. If in doubt discuss with the SMO
- Use the glucose calculator as a guide to the drop in mg/kg/min when alterations are being made to ensure changes are not too aggressive
- Make sure to check the glucose intake if a drip tissues and a plan is made to increase enteral feeds faster and not replace the line as the glucose intake may be significantly lower running the risk of hypoglycaemia recurring
- BSL must continue to be monitored when weaning (see below)

BSL 1.5-2.5mmol/L

- If the baby is asymptomatic and has not responded to dextrose gel, breastfeeds or top-ups on maternity then place a nasogastric tube and feed enterally between 65-90ml/kg/day depending on their age and previous milk intake volumes as a 1-2 hourly feed volume
- Breastmilk volumes will likely be insufficient and so pasteurised donor milk (PDM) or formula will be required. PDM should be offered first but if this is declined then consent for formula will be needed or iv fluids used. If parents are concerned because of a family history of cow’s milk allergy then Peptijunior or Neocate may be used in the short-term but this needs to be discussed with the SMO and dietician.
- If BSL 1.5-1.9mmol/L - consider a 2ml/kg 10% dextrose at 2mls/min bolus and start an IV infusion of 10% dextrose at 65ml/kg/day or if already on a dextrose infusion increase the rate or concentration and recheck the BSL in 30 minutes
- If BSL is 2.0-2.5mmol/L consider increasing the feed volume if tolerated or start an iv infusion as above and recheck the BSL in 1 hour
- Calculate the glucose load in mg/kg/min with each alteration in fluid rate or concentration (Click Here)
- Wean iv fluids when BSL have been stable for 12 hours as per the section above.
BSL Monitoring
- Every 30 minutes until ≥2mmol/L, then,
- Hourly until ≥2.6mmol/L, then,
- 4-6 hourly prefeed but this is very dependent on the clinical context and will be individualised
- **Transient hypoglycaemia** - stop BSL when there are 3 consecutive BSL ≥2.6mmol/L and
  - is off iv fluids
  - has transitioned to 3 hourly enteral feeds without additional caloric supplements
  - is not on medication to support the blood sugar level
- **Persistent or Recurrent hypoglycaemia** - BSL monitoring will need to be continued 4-12 hourly depending on the clinical scenario for a longer period and needs to continue until:
  - has transitioned to 3 hourly enteral feeds
  - nutritional supplements are weaned
  - is not on medication to support the blood sugar level
  - requires 3 consecutive BSL ≥3.5mmol/L
  - a 6 hour safety fast is required before discharge

**Indications for a Hypoglycaemia Screen**
- **Severe Hypoglycaemia** - BSL <1.5mmol/L
  - Not all babies with a one off/admission low BSL will need a screen - consider the clinical context and risk factors
- **Hypoglycaemia associated with seizures**
- **Glucose requirement >10 mg/kg/min**
- **Early onset hypoglycaemia** - that persists or recurs after 72 hours despite adequate iv or enteral feed volumes
  - Take a screen even if the BSL is not significantly low ie: <3mmol/L as the results will still be helpful for the Endocrine team. These babies should have a BSL ≥3.5mmol/L at this age and so we should not accept BSL <3mmol/L as being normal.

Packs with the appropriate tubes and prewritten form are available in the Lab room along with a sticker for documentation of the results.

Check the blood sugar on a gas prior to taking “hypoglycaemia bloods” as glucometers are less accurate at lower blood glucose levels. Take the bloods before a feed or additional iv dextrose is administered but do not stop the current iv infusion.

<table>
<thead>
<tr>
<th>Hypoglycaemia Bloods</th>
<th>Blood tube</th>
<th>Lab</th>
<th>Result Time</th>
<th>Blood Tube Summary</th>
</tr>
</thead>
<tbody>
<tr>
<td>Formal Glucose</td>
<td>Green 0.6mL</td>
<td>Daily</td>
<td>2-3hrs</td>
<td>Minimum requirements:</td>
</tr>
<tr>
<td>Beta-hydroxybutyrate</td>
<td>Green 0.6mL</td>
<td>Daily</td>
<td>1-2hrs</td>
<td>3 full EDTA pink tubes (one on ice). A 4th tube would be helpful if able</td>
</tr>
<tr>
<td>Cortisol</td>
<td>Green or Pink 0.6mL</td>
<td>M-F, not w/e or holidays</td>
<td>1-4 days</td>
<td></td>
</tr>
<tr>
<td>Plasma Insulin</td>
<td>Pink 0.6mL on ice</td>
<td>M-F, not w/e or holidays</td>
<td>1-4 days</td>
<td></td>
</tr>
<tr>
<td>C-peptide</td>
<td>Pink 0.6mL on ice</td>
<td>2x a week M-F, not w/e or holidays</td>
<td>3-7 days</td>
<td></td>
</tr>
<tr>
<td>Ammonia</td>
<td>Pink 0.6mL on ice</td>
<td>Daily</td>
<td>1-2hrs</td>
<td>1 blood gas</td>
</tr>
<tr>
<td>GH</td>
<td>Pink 1mL</td>
<td>2x a week M-F, not w/e or holidays</td>
<td>1-5 days</td>
<td></td>
</tr>
<tr>
<td>Free fatty acids</td>
<td>Pink 1mL</td>
<td>M-F, not w/e or holidays</td>
<td>1-4 days</td>
<td></td>
</tr>
<tr>
<td>Blood gas - lactate</td>
<td>Capillary tube</td>
<td>NICU</td>
<td>Immediate</td>
<td></td>
</tr>
<tr>
<td>TSH, T4 (Biochem)</td>
<td>Green 0.6mL</td>
<td>Daily</td>
<td>2-3hrs</td>
<td></td>
</tr>
</tbody>
</table>

Urine metabolic screen can be an additional test if the hypoglycaemia bloods do not point to an obvious cause. The urine does not need to be collected when a baby is hypoglycaemic.

These tests are to show what the baby’s biochemical and hormonal response to hypoglycaemia. This may provide a diagnosis or guide further investigation.
Documentation of the results needs to be on the prepared results sticker and placed on the Investigation sheet for interpretation and sign off by the lead SMO. It also has advice for referral to Endocrine and the need for further tests such as MRI and a 6 hour fast pre discharge.

Results in the normal range may not actually be normal for the clinical situation and reference ranges cannot be relied upon in these circumstances.

Results needs to be discussed with the SMO and Endocrinologist as they need to be interpreted along with the clinical situation

Criteria for Endocrinology Referral

1. Glucose requirement of >10 mg/kg/min
2. Severe hypoglycaemia - BSL <1.5mmol/L at any time
   - Not all babies with a one off/admission low BSL will need a referral - consider the clinical context and risk factors
3. Persistent hypoglycaemia – BSL <2.6mmol/L that persists past 72 hours age
4. Recurrent hypoglycaemia - BSL <2.6 that recurs after seeming to resolve
5. Hypoglycaemia in a baby with no obvious maternal or neonatal risk factors such as maternal diabetes, IUGR, LGA, perinatal stress
6. For interpretation of a hypoglycaemia screen result
   Transient hypoglycaemia is common in the NICU but there will be a group of patients with an underlying condition such as a metabolic or endocrine disorder that present with severe, recurrent or persistent hypoglycaemia and these babies need specialist input and investigation

Endocrine - Hyperinsulinism, Hypopituitarism
Metabolic - Galactosaemia, Fatty acid oxidation disorders, Glycogen storage disease, Maple Syrup Urine Disease, Organic acidaemia, Tyrosinaemia

Criteria for a Safety Fast

1. All the conditions listed above that warrant an Endocrine referral, plus
2. Confirmed hyperinsulinism
3. Confirmed hypopituitarism
   - To be done 5-7 days prior to discharge, in the daytime – taking into account weekends/public holidays and results turnaround
   - No sucrose or milk to be given to the baby to settle them during the fast
   - IV access is not required prior to the test
Time 0  Baseline BSL then breastfeed or bottle feed baby a 3hourly amount
3hrs  BSL on gas machine
4hrs  BSL on gas machine
5hrs  BSL on gas machine
6hrs  BSL on gas machine

BSL <3.0mmol/L at any time: take bloods for a hypoglycaemic screen, then STOP the fast, feed the baby and consider an im glucagon bolus depending on how low the glucose level is

BSL ≥3.0 mmol/L: continue the fast after the 6 hour BSL feed the baby and no bloods are needed

Hyperinsulinism

- Rare but significant cause of severe, recurrent hypoglycaemia in infancy
- Insulin secretion is inappropriately high and poorly regulated
- Insulin inhibits: glycogenolysis, gluconeogenesis, lipolysis, ketogenesis, glucagon and cortisol responses
- Therefore the body is unable to use counter-regulatory hormones in the face of hypoglycaemia
- The triad to diagnose hyperinsulinism includes – absent ketones and a detectable insulin level in the presence of hypoglycaemia
- If this diagnosis is considered take blood tests listed above at the time of hypoglycaemia
- Caused mainly by mutations involving the SUR1 and Kir6.2 proteins in the K\(^{+}\)ATP channel in the pancreatic beta cell membrane that regulates insulin secretion
- Mutation in the GLUD-1 gene causes activation of glutamate dehydrogenase that also causes an increased release of ammonia so it is imperative to check for ammonia if hyperinsulinism is being entertained
- Note that not all hyperinsulinaemic babies will be macrosomic

Diagnosis

- BSL <2.6mmol/L
- Inappropriately high insulin or C peptide for the glucose concentration (note the level of the insulin does not predict the severity of the hypoglycaemia)
- Increased glucose requirements >10 mg/kg/min
- Low free fatty acids
- Absent ketones
- Cortisol will be slightly lower than expected
- Rapid response to glucagon - expect a rise of >1.5mmol/L after administration

Management

- Secure good iv access with a central line
- Provide frequent feeds or iv dextrose as needed to maintain BSL’s
- Aim to keep BSL >3.5mmol/L as these babies have no other energy source for their brains to utilise
- Document the mg/kg/min of dextrose that is required to maintain BSL’s
- Use glucagon 200 mcg/kg im if an iv tissues and there is difficulty re-siting another – it acts by glycogen being released in the liver into glucose to be utilised
- Diazoxide – start at 2mg/kg/dose 8 hourly and increase to 3.5mg/kg/dose 8 hourly then up to a maximum of 5mg/kg/dose 8 hourly if needed. Diazoxide works by inhibiting insulin secretion. Most responders will do so at the lower dose. Higher doses will have the risk of more side effects such as fluid retention and oedema.
- Diazoxide is albumin bound so if a baby is hypoalbuminaemic the starting dose of diazoxide would be lowered to 1mg/kg/dose 8 hourly or run the risk of excessive fluid overload
- Diazoxide can worsen pulmonary hypertension and so consider withholding treatment until a baby is stable and has transitioned appropriately
- Chlorothiazide – used as a diuretic to counteract the fluid retention caused by diazoxide but is also synergistic with diazoxide by inhibiting insulin secretion by other methods. Must be started any time diazoxide is used.
- Octreotide for sugars resistant to diazoxide
- Surgical treatment only if persistent – partial or 95% pancreatectomy
Hypopituitarism

- Another rare cause of persistent hypoglycaemia
- Can be seen in association with temperature instability, hypotonia, roving nystagmus, prolonged jaundice, conjugated jaundice, micropenis, undescended testes, midline defects such as absent corpus callosum or cavum septum pellucidum
- Consider this as a cause of persistent or recurrent hypoglycaemia if there are no clear risk factors in the maternal or neonatal history

Diagnosis

- Recurrent low glucose which but may not be severely low. Remember a BSL of >3.0 mmol/L after a few days of age is not normal
- Low cortisol
- Low GH
- Low TSH
- Appropriately low insulin and C-peptide levels
- Normal ketones

Management

- This will be directed by the Endocrine team
- Hydrocortisone replacement must start 24 hours prior to thyroxine replacement
- Note there is a chance of diabetes insipidus after starting medical management. This would be detected by a large increase in the urine output.
- Thyroid function tests will need to be taken frequently ad dose adjustments made as directed by the Endocrine team
- Referral to Opthalmology to examine the optic discs which may be pale if there is an association with septo-optic dysplasia.

Hyperglycaemia

- Blood glucose above 7 mmol/L are elevated.
- Very low birthweight infants are at risk of developing hyperglycaemia due to reduced glucose tolerance and insulin resistance.
- Hyperglycaemia is associated with increased fluid and electrolytes losses, IVH, sepsis and mortality
- Occurs in those usually <28 weeks or <1000g during periods of stress, sepsis, instability or when steroids are given
- Consider and investigate for sepsis as a cause of the hyperglycaemia rather than just putting it down to prematurity
- Management includes
  - reviewing the glucose intake to make sure it is not excessive ie: >9mg/kg/min (see glucose calculator under TPN Folder) and if so alter the fluid volume or change from 12.5% to 10%
  - change any extra infusions eg dopamine/morphine into 5% dextrose or saline
  - start insulin
  - decreasing dextrose concentration to 7.5% but this will impact on nutrition and growth
- When 2 consecutive blood sugars 4 hours apart are >10 mmol/L, and/or there is dextrose present in the urine, insulin should be commenced after discussing with the SMO.
- An arterial line for frequent blood sampling is advisable if there is hyperglycaemia.
- Blood sugars to be checked 2-4 hourly initially and if very stable then 4 hourly and never check less frequently than 6 hourly

Insulin Infusion

- Babies who receive insulin should be entered in the insulin computer for insulin dosing predictions
- Enter the baby on the computer prior to starting insulin – hyperglycaemia is not an emergency
- Chart insulin on the insulin infusion sheet
  - Commence at 0.05 units/kg/hr in most cases
  - If BSL >20mmol/L consider starting at 0.1 unit/kg/hr
• Occasionally in a resistant baby the infusion will need to be made “double strength” to limit the volume
• The Registrar or CNS/ANP must be informed about every blood sugar that is taken when a baby is on an insulin infusion
• The aim is for a gradual weaning of the BSL below 10mmol/L and to stop glucose leaking in to the urine - it is more dangerous to have a low BSL than a high BSL
• The target range for the insulin computer is 4-8mmol/L
• The insulin infusion must always run through the same iv line as the dextrose infusions so that if the line tissues the infusions stop simultaneously
• If the baby’s insulin dosing is not via the insulin computer then
  – decrease the rate when the BSL is 5-8mmol/L
  – stop the insulin infusion when the BSL is <5.0mmol/L

Insulin Computer
• Stored in a labelled drawer in the equipment room
• Place on a separate trolley and secure to the trolley with the lock – code 999
• Always have it plugged in to save power
• Turn on and select STAR-GRYPHON icon
• STAR (Stochastic TARgeted) and GRYPHON (Glucose Regulation sYstem to Prevent Hyper and hypoglycaemia in Neonates)
• STAR setting – can choose between training mode and clinical mode – ensure the training mode is OFF if using on a patient
• Select New Patient
• Enter all the fields, making sure that the times for the blood sugars, feeds and infusions are correct
• Working weight is the birth weight until it is surpassed
• Insulin concentration is automatically calculated form the working weight and will round down to 2 decimal points. The clock defaults to 30min after the blood sugar was taken to allow time for the infusion to be prepared – if there are further delays then update the time that the infusion is expected to start
• The first prediction is always 0.05units/kg/hr or 0.2mL/hr (if baby <1000g 0.05units/kg/hr will be 0.21mL/hr and the pump will also show this so do not worry). Choose when you want the next blood sugar to be taken in 2,3 or 4 hours time acknowledging that it takes 1-2 hours for the infusion to reach the baby
• When the next sugar is taken – click Calculate treatment to be taken through the steps
  – Insulin – check working weight is the same, if not update the weight and click Calc Conc to update the concentration and check that this working weight is being used for the insulin infusion recipe. If not then rechart the infusion with the new concentration and have a new bag prepared
  – Parenteral Nutrition – check that the infusion rates of dextrose containing fluids are accurate. If there have been any changes since the last sugar was taken then adjust the nutrition rates and/or add a new infusion in
  – Enteral Nutrition – need to enter the feed regime each time as it can change frequently, ensure that the time of the next feed is correct. If there is no feed regime then click the ON button to say OFF
  – Blood Glucose – enter the latest value
  – Recommendations – options for dosing are given depending on when you want the next sugar to be taken. There is also an option for stopping insulin but that is usually left for when a baby is better and trialling off insulin. The graph shows 95% confidence intervals for where the blood sugar will be in the next 2,3 or 4 hours time to help you decide which option to pick. Chart the new dose and rate on the infusion sheet
  – Repeat the process with every blood sugar taken
• When to stop insulin
  – SMO decision taking into account the stability of the blood suagrs, the rate of insulin (usually <0.05 units/kg/hr) and the clinical picture
  – When insulin is off, continue to monitor blood sugars in case the baby rebounds and needs insulin again
  – If within 8 hours of stopping insulin the baby starts insulin again then update the information on the current episode and continue
  – If insulin is needed after being off for more than 8 hours click “Start a new episode” which will autofill some data and allow you to quickly update the nutritional data
• To save a summary of the insulin course after it has been stopped
  – Click Star data helper, choose patient, view graphs, print summary, then view tables and print summary, connect tablet to the computer using the charger cables
  – My computer / Portable media player / tablet / Patient summaries / Right click and copy / Save in Insulin Computer folder G:drive under Patient summaries / Rename with patient details and print a copy for the notes
Inborn Errors of Metabolism

- Presentation is usually non-specific, with poor feeding, lethargy, vomiting and tachypnoea.
- Acute encephalopathy is the predominant presentation of metabolic disease in the newborn.
- Early consultant input and consultation with a metabolic paediatrician is essential in suspected cases as irreversible neuronal injury occurs rapidly.

Think of the possibility of inborn errors of metabolism in the presence of:
- Consanguinity
- Family history of unexplained neonatal death, or a sibling with unexplained mental retardation
- Unexplained illness in a term baby, usually but not exclusively, after commencing feeding
- Sudden onset of overwhelming illness

### Metabolic Differential Diagnosis of Acute Encephalopathy

<table>
<thead>
<tr>
<th>1. Aminoacidopathy</th>
</tr>
</thead>
<tbody>
<tr>
<td>2. Organic Acidemia</td>
</tr>
<tr>
<td>3. Disorders of fatty acid oxidation</td>
</tr>
<tr>
<td>4. Urea cycle defects</td>
</tr>
<tr>
<td>5. Defects of pyruvate metabolism</td>
</tr>
<tr>
<td>6. Defects of mitochondrial electron transport chain</td>
</tr>
</tbody>
</table>

Differential diagnosis includes:
- Septicaemia
- Congenital heart disease eg. hypoplastic left heart.

### Presentation

- Acid-base disturbance
  - Suspect if tachypnoea, grunting and apnoea with a normal CXR (Kussmaul breathing).
  - Conditions presenting in this way include the organic acidemias.
  - If acute encephalopathy with unexplained lactic acidosis consider disorders of pyruvate and citric acid cycles, disorders of mitochondrial electron transport chain.
  - Defects in gluconeogenesis may also cause a raised lactate.
- Hyperammonemia and encephalopathy
  - consider organic acidemias, urea cycle defects, transient hyperammonemia of newborn
- Intractable newborn seizures
  - nonketotic hyperglycinemia, pyridoxine dependent seizures, sulfite oxidase deficiency, molybdenum cofactor deficiency
- Unexplained hypoglycaemia.
  - effects in gluconeogenesis
- Jaundice, hepatomegaly, bleeding diathesis,
  - galactosemia, fructosemia, and tyrosinemia type 1
- Unusual smells
  - PKU, MSUD, isovaleric acidemia.
- Neonates with galactosemia may also present with overwhelming gram negative sepsis.

### Investigations

Discuss specimen collection with the laboratory (especially Chris Leaver, Head Technician in Special Biochemistry)

It is very important to try and make the diagnosis for the family for genetic counselling
- Blood gases
- Urea and electrolytes
- Glucose
- Ammonia
- Urine for reducing substances and ketones.
• Blood and urine amino acids
• Urinary organic acids and acylcarnitine profile
• Pyruvate and lactate if the baby is acidotic and/or hypoglycaemic

If the infant dies, try to ensure appropriate specimens are taken:
• Plasma - deep freeze 5 mls in aliquots at -20 degrees.
• White cells - deep freeze for DNA.
• Urine - deep freeze 10 mls.
• Skin biopsy - store at 4 degrees.
• Liver biopsy - a needle or open biopsy: A needle biopsy may be taken for histology, EM and enzyme studies. This should be deep frozen (preferably snap frozen).

**Initial Management in Suspected Metabolic Encephalopathy**
• Stop protein feeds
• Stop catabolism (give IV fluids with good concentration of glucose eg: 10% and lipids at 1g/kg/day)
• Specific medications and/or haemofiltration as guided by metabolic paediatrician

**Maternal Thyroid Disease**
• Maternal stimulating or inhibiting thyroid antibodies may cross the placenta and transiently affect the baby
• Occasionally some of these infants will need treatment in the neonatal period.
• When there are more than two generations affected by thyrotoxicosis or more that one first degree relative consider the even rarer autosomally dominantly inherited activating mutations of the TSH receptor. This is important as the neonatal thyrotoxicosis is more persistent
• Any symptomatic baby needs a medical review
• Note that the Guthrie will only detect hypothyroidism

**Maternal Hypothyroidism secondary to ectopic thyroid or thyroid aplasia**
• There is only a slightly increased risk in the neonate and a Guthrie card test should suffice.

**Maternal Graves’ Disease**
• In mothers with a history of Graves’ disease will have circulating thyroid stimulating antibodies which can cause a transient hyperthyroid state in the neonate that may require treatment.
• This can occur even if the mother has been rendered hypothyroid by surgery or radioiodine treatment.
• Both carbimazole and propylthiouracil cross into breast milk but do not appear to affect the neonate if maternal doses are less than 15mg per day of carbimazole and less that 150mg per day for PTU
• Although rare neonatal thyrotoxicosis is associated with a high mortality.
• The incidence of Graves’ is estimated to be about 0.2% and only 1-10% of infants will subsequently be affected. The highest risk is in those whose mothers are receiving antithyroid treatment at the time of delivery.

**Maternal Hashimoto’s**
• In mothers with Hashimoto’s thyroiditis there may be circulating stimulating or inhibiting antibodies that may transiently affect the neonate.

**Investigations**
• The optimum timing of thyroid function tests in the newborn is debatable.
• We know that there is a natural physiological surge in TSH and subsequently T4 at about 30 minutes post delivery. The TSH falls over the next 5 days with the T4 gradually declining over the next 2 weeks.
• Babies at high risk of (mothers with active Graves’ in pregnancy, known maternal thyroid antibodies on blood tests, or family history of activating mutations of TSH receptor) or with symptoms of hyperthyroidism (tachycardic, weight loss, loose stools, increased temperature/hot, jittery/hyperactive) should have thyroid function tests done at 2 days, in conjunction with the Guthrie (or earlier if symptomatic) and repeated at 1 week with close observation for symptoms.
- Babies at **low but some risk** of hyperthyroidism (past history of Graves’ disease or Hashimoto’s thyroiditis) or hypothyroidism (Hashimoto’s thyroiditis) should have thyroid function tests checked at 5-7 days and parents advised of symptoms of thyroid disease.
- Babies at **low risk** (mothers with an ectopic or aplastic thyroid, or on thyroxine replacement with no antibodies) should simply have a Guthrie done.
- Babies whose mothers are on thyroxine but have no thyroid antibodies only need a Guthrie.
- TFT’s (TSH, free T4, free T3) are done in the biochemistry lab at Canterbury Health Lab for screening in low risk/asymptomatic babies or as part of a prolonged jaundice screen. **Fill one green tube to the top line, this can be done by the midwife by heelprick along with the Guthrie.** Results available that day.
- TFT’s (TSH, total T4, total T3, free T4, free T4 index) should be sent to the Endo lab **(fill 2 Red tubes so recommend this is done by venepuncture and not heel prick)** if the baby is high risk/symptomatic or if an Endocrine referral has been made. Note that the Endo lab does not process bloods daily or at weekends and so if a result is needed that day then the blood should be sent to the Canterbury Health Lab as above.

**Process**

- LMC identifies in pregnancy that the baby will need screening blood tests after birth.
- LMC is responsible for ensuring the blood form is completed and their name on the form to check the result.
- Liaison with the Neonatal team at any stage is appropriate but these babies are frequently discharged prior to any of these blood tests being taken and so needs LMC oversight.
- LMC to ensure the bloods are taken (if on day 2 then combine with the Guthrie and if in CWH then to be discussed with core staff about who will take the blood).
- LMC is the primary person responsible for checking the result. If the results are normal then no more action is needed. If the results are abnormal then that is the time to contact the Neonatal team on 5039 (weekdays) or 5019 (weekends) for help in interpreting the result.
Murmurs

- The following recommendations are based on the fact that the majority babies will have an audible murmur (often quite transiently) some time in the first 24 hours, caused by closure of the ductus arteriosus or other circulatory changes related to the perinatal transition.
- The murmur of normal ductal closure in a well term infant is typically a systolic murmur with blowing or "whooshing" quality. It can be reasonably loud but should never be accompanied by a precordial lift or thrill, abnormal peripheral pulses, cyanosis. It is usually short, as opposed to the holosystolic or machinery murmur of a persistent patent ductus arteriosus in an older baby, presumably because the pulmonary vascular pressures are still relatively high. With experience, you will get used to these innocent murmurs and will distinguish them from murmurs that sound more pathological in origin.
- However, it is also important to recognize that several of the most serious congenital heart defects that present in the first week of life can be associated with soft or insignificant sounding murmurs. Thus, in excluding serious congenital heart disease, the rest of the cardiovascular examination is just as important as auscultating the heart.
- If the baby is well and has a normal examination, and is less than 24 hours old, re-examine in 24 hours.
- Examine the baby daily up to day 4 or until the baby is discharged. **If the murmur is still present on day 4 or at discharge:**
  - Perform pre (right wrist) and post ductal oxygen saturations (feet) prior to discharge. The oxygen saturation should be read once a satisfactory trace has been obtained (for at least 5 minutes)
  - Infants in whom the oxygen saturation is <95% on either of the recordings or where there is a significant difference (≥3%) between the two readings should have an echo performed prior to discharge
  - Infants in whom the oxygen saturation is <90% or in whom there is clinical concern should be seen by the neonatal registrar/CNS-ANP and admitted to the NICU
  - Infants in whom the oxygen saturation is ≥95% on both of the recordings should be booked in to the Echocardiogram clinic on a Wed afternoon in CWH Radiology.
- **Antenatal VSD's** - Babies with a membranous VSD of any size, or, a muscular VSD of 3mm or more should have an outpatient echo arranged (see below). Muscular VSD's of 1 to 2mm in size only need to be referred for a scan if there is a murmur present as often these have closed antenatally. If a murmur becomes audible within the next 4 weeks the GP or LMC can refer back to the Neonatal team to arrange a heart scan at that stage.
- Order echo by completing an electronic radiology request on Health Connect South. Consultants will triage and arrange a Wed appointment within 1-2 weeks
- If an echo is requested by the Consultant to be performed by the echo technicians from cardiology, complete an Echo request form and fax (81449)
- Perform CXR, ECG, 4 limb BP if baby unwell or echo indicates a significant shunt.
- When doing an ECG on a baby, place the leads as follows:
  
  Use the:  
  
  V1 lead as V4R  
  V2 lead as V1  
  V3 lead as V2  
  V4 lead as V3  
  V5 lead as V4  
  V6 lead as V6

**Neonatal ECG Guide**

Please note: many values are different to paediatric or adult guidelines

Each 1mm square on the ECG equates to 40ms or 0.04sec

- Rate: 1500/R-R interval (in mm)
- Rhythm: Sinus (if p waves present)
- P axis: 0-90°
- QRS axis: 60-180°
- T axis: 1st 72 hours: +ve in V1 and V4R.  
  After 72 hours: -ve in V1 and V4R  
  Similar progression in V2 and V3)
- P waves < 3mm in any lead, ≤ 0.08s
• QRS complex 0.06-0.08s
  RS progression: gradual progression up to 3 yrs
  “Neonatal pattern” (after 72 hrs): R>S in V1; S>R in V5 and V6
  “Infantile/neonatal” pattern (after neonatal period): R may be > S in V1 as well as V5-6
  S in V6 < 15mm by 1 wk; < 10mm by 6mths; < 5mm by 1yr
• Q wave May be present and normal in V1 and V4R.
  Always less than 25% of R in same lead.
  < 6mm in aVF and V5, < 5mm in V6, ≤0.03s.
• ST segments ST depression up to 2mm (precordial) may be normal
• PR interval 0.08-0.12s
• QT interval < 0.45s under 6 months, then < 0.44s, if >0.49s then needs urgent cardiology review

Corrected QT interval QTc=QT/√R-R (all in seconds)

<table>
<thead>
<tr>
<th>Condition</th>
<th>Rate/rhythm</th>
<th>P</th>
<th>QRS</th>
<th>ST</th>
<th>T</th>
<th>Other</th>
</tr>
</thead>
<tbody>
<tr>
<td>HyperK+ Ventricular arrhythmias</td>
<td>Small/flat</td>
<td>↑, small R</td>
<td>↓↓</td>
<td>Tall, pointed</td>
<td></td>
<td></td>
</tr>
<tr>
<td>HypoK- arrhythmias</td>
<td>↑</td>
<td>↑</td>
<td>↓↓</td>
<td>↓</td>
<td>U waves</td>
<td></td>
</tr>
<tr>
<td>RV Hypertrophy</td>
<td>R axis deviation</td>
<td>R V1 &gt; 30mm</td>
<td>S V6 &gt; 15mm</td>
<td>R/S V1 &gt; 6.5mm</td>
<td>Up in V1 &gt;3days</td>
<td></td>
</tr>
<tr>
<td>LV Hypertrophy</td>
<td>S V1+R V6&gt;30mm</td>
<td>R V6 ≥ 15mm</td>
<td>S V1 &gt; 20mm</td>
<td></td>
<td></td>
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<tr>
<td>Acute myocarditis</td>
<td>Frequent ectopics</td>
<td>Q waves</td>
<td>↓</td>
<td>flat</td>
<td>↑QTc</td>
<td></td>
</tr>
<tr>
<td>Hypothermia Sinus brady</td>
<td>↑PR</td>
<td>↑PR; ↓</td>
<td>↓</td>
<td>↓</td>
<td>↑QTc J waves</td>
<td></td>
</tr>
<tr>
<td>ASD Incomplete RBBB</td>
<td>↑</td>
<td>↑RV hypertrophy</td>
<td>Primum: Lt axis deviation</td>
<td>Secundum: Rt axis deviation</td>
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<td></td>
</tr>
<tr>
<td>Hypothyroid Sinus brady</td>
<td>↑PR; ↓</td>
<td>↓</td>
<td>↓</td>
<td>↓</td>
<td>U; ↑QTc</td>
<td></td>
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</tbody>
</table>

**Persistent Pulmonary Hypertension of the Newborn**

**Primary PPHN**
- Primary dysfunction of the pulmonary endothelium
- Usually idiopathic

**Secondary PPHN**
- Birth asphyxia
- Meconium aspiration
- Diaphragmatic hernia
- Sepsis
- Severe hyaline membrane disease

**Diagnosis**
- A 20 mmHg difference between pre and postductal oxygen, if the ductus arteriosus is open
- Ultrasonographic presence of a right to left shunt at ductal level, through a patent foramen ovale, or more subtle echocardiographic signs (ie: TR, septal flattening in systole) can also indicate PPHN is present.
Management
- Establish adequate monitoring with pre and post ductal transcutaneous oximetry and arterial lines
- Liberal oxygen use as this is a pulmonary vasodilator, aim for a PaO₂ 100-120mmHg
- Avoid noxious stimuli and keep quiet
- Use analgesia as needed to calm the baby and achieve better matching of tissue oxygen supply/demand
- Maintain PCV > 0.35 to ensure good oxygen carrying capacity but treat polycythemia
- Aim for a normal pH 7.30-7.40.
- Consider using a bicarbonate infusion e.g. 0.5-1 mmol/kg/hour if necessary.
- Use muscle relaxants if necessary to reduce metabolic demands and promote stability.
- Treat ventricular failure and maintain adequate systemic blood pressure with volume and pressors, e.g. dobutamine, milrinone, dopamine as necessary.
- A higher than normal systemic pressure may be needed to reduce right to left shunting and maintain pulmonary blood flow.
- Consultants will guide optimum MAP/ pCO₂/ pO₂ in each case.
- Inhaled nitric oxide should be used if there is evidence of PPHN and the baby is receiving maximal respiratory support and failing to oxygenate well (eg: oxygenation index 30-40)
- NO improves oxygenation markedly in primary PPHN
- Sildenafil can also be an adjunct to nitric oxide in severe pulmonary hypertension and is available po and iv with an iv infusion sheet available on the Intranet
- Prostaglandins may be required to maintain duct patency to offload right heart strain
- Intravenous vasodilators e.g. nitroprusside or nitroglycerine have a role.

\[
\text{Oxygenation Index} = \frac{\text{MAP} \times \text{FiO}_2 \times 100}{\text{PaO}_2}
\]

<table>
<thead>
<tr>
<th>Condition</th>
<th>Oxygenation Index</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal Infant</td>
<td>2.1 - 2.5</td>
</tr>
<tr>
<td>Infant in 100% O₂, Paw; 25, PaO₂; 50</td>
<td>50</td>
</tr>
<tr>
<td>Infant in 21% O₂, Paw; 10, PaO₂; 95-100</td>
<td>2.1 - 2.2</td>
</tr>
</tbody>
</table>

Nitric Oxide
- Inhaled nitric oxide (iNO) is currently the standard vasodilatory agent used for treatment of PPHN
- Multiple clinical trials conducted since 1992, including a Cochrane review in 2009, have demonstrated its benefit in treating PPHN
- Nitric oxide is recommended for use in the term and late preterm population. Its role in the extreme preterm infant is controversial but may be considered for temporary management of acute hypoxaemia whilst awaiting other treatments to take effect
- Nitric oxide diffuses across alveoli and causes vasodilation by relaxing smooth muscle in pulmonary arterioles. This decreases pulmonary resistance and improves oxygenation
- The benefit of inhaled nitric is its ability to be delivered directly to the lungs with no or minimal systemic action, therefore no risk of systemic hypotension. However, prolonged and unnecessary use of iNO is not recommended
- The formation of byproducts such as methemoglobin and nitrogen dioxide can have adverse and toxic effects.
- iNO is expensive drug so should be judiciously used and weaned as early as possible

Indications for commencing Nitric Oxide
- This is a consultant decision based on a clinical diagnosis of PPHN supported by echocardiography confirmation and an oxygen index of >20 (however consider nitric oxide at an OI of 15)
- Echocardiography can confirm high pulmonary arterial and right ventricular pressures based on tricuspid regurgitation, as well as exclude cyanotic congenital heart disease, a major differential diagnosis
- Ideally an echo should be done prior to starting iNO but in an emergency iNO should be started as soon as possible followed by an echo in 1-2 hours
- A chest X-ray should be done prior to ensure adequate lung recruitment
Commencing Inhaled Nitric Oxide

- After confirming the diagnosis of PPHN and taking baseline observations and blood gas (preferably arterial) iNO should be commenced at 20ppm. Higher doses have not been found to offer any additional benefit and carry increased risk of toxicity
- A repeat arterial blood gas should be taken after 30-60 minutes of commencing therapy to assess response. A positive response is suggested by an increase in PaO2 ≥ 20 mmHg, increase in sats by 10% or being able to drop FiO2 ≥ 0.2
- If a positive or partial response is not seen within the first hour, the options include immediately stopping the iNO, or continuing for 12-24 hours before weaning. This is in view of evidence suggesting that 35-40% of newborn infants may not respond to iNO
- It is unsafe to stop abruptly if iNO has been administered for longer than one hour due to the risk of rebound PPHN. If a positive or partial response is seen then iNO should continue for four hours, weaning the FiO2 as able
- Methaemoglobin (F MetHb) levels (%) are automatically measured and reported in all the gases. Levels should be reviewed on the first gas after commencing and then 24 hourly. Levels of MetHb up to 2.5% are considered normal and safe. Levels above that should be closely watched and iNO should be discontinued if levels >10%.

Weaning of Nitric oxide

- Once iNO has been commenced, consideration of timely and appropriate weaning is important
- Rebound pulmonary hypertension following prolonged use and abrupt withdrawal of iNO is a significant risk due to the down regulation of any endogenous NO production
- Weaning should be considered at four hours if FiO2 is <0.6. Decreasing iNO from 20ppm down to 5ppm quickly has been found to be safe. This should be done by halving the iNO from 20ppm, to 10ppm to 5ppm, every 1-2 hours if the patient remains stable
• Indications not to wean include an increase in FiO2 of >0.2, a decrease in oxygen saturations of >5% or a pre/post ductal saturation gradient of >10%. In this situation iNO should be returned to previous dose and continued for four hours until any further attempts at weaning are made.
• Once the patient is stable at 5ppm iNO, weaning should occur in 1ppm steps, every 1-2 hours until off.
• The old Aeronox system can wean to 1ppm then off.
• The new Aeronox 2 system can wean to 2ppm only then off.
• At discontinuation, it may be necessary to increase the FiO2 by 0.2 temporarily to counter rebound PPHN.
• Again, if at any point there are criteria fitting weaning failure the iNO should be returned to the previous dose for four hours before attempting further weaning.
• If the patient remains on iNO at 96 hours, review of differential diagnosis is recommended.
Criteria for Weaning Failure:
- ↑FiO2 by 0.20
- ↓SpO2 by >5%
- Pre/post ductal SpO2 gradient of >10%

Stop weaning and return to previous dose of NO

Weaning Considerations:
- If weaning failure criteria are met, wait 4 hours before reattempting wean
- If >96 hours on NO consider repeat ECHO and consider differentials for pulmonary hypertension

Discontinue NO if using Aeronox 2:
At discontinuation it may be necessary to increase FiO2 by up to 0.20

Discontinue NO if using Aeronox:
At discontinuation it may be necessary to increase FiO2 by up to 0.20
Patent Ductus Arteriosus

- The incidence of haemodynamically significant PDA (hsPDA) varies on the definition, the time of assessment and gestational age.
- In normal term infants, 90% of PDAs are closed by 48 hours and 98% by 96 hours.
- Closure can be delayed with increasing prematurity.
- The difficulty is determining in which babies closure will be delayed which may then lead to significant left to right shunting and adversely affect pulmonary function.
- Early prophylactic administration of indomethacin reduces the need for symptomatic treatment of PDA but in randomised controlled trials has not affected pulmonary or developmental outcome (TIPP study NEJM 344(26):1966-72).
- As a result we use targeted treatment of PDA’s based on clinical features or echocardiogram diagnosis.

Clinical signs to assess significance of PDA

- systolic murmur, full radial pulses
- active praecordium
- wide pulse pressure on invasive monitoring
- systemic hypotension.
- clinically significant but silent PDAs (no murmur) are common especially in the extremely premature baby and in the first 3-5 days of life
- late features (after day 3-5) include increasing ventilation requirements, cardiomegaly, hepatomegaly and pulmonary plethora on the CXR.

Echocardiogram

Gold standard for diagnosis and the following all contribute to the echo assessment of the PDA. Those in bold have more recently been used to determine haemodynamic significance.

- Increased LA dimension
- Increased LA:AO ratio
- Increased LV end diastolic dimension
- Absent or reversed flow in the descending aorta
- Non-restrictive pattern on PDA doppler flow
- Turbulent main pulmonary artery peak diastolic flow >0.2m/sec
- Left pulmonary artery peak diastolic velocity >0.3m/sec
- Absolute dimension of Ductus Arteriosus.

Timing of assessment has implications on significance.

- Nick Evan’s group use a “targeted early closure” approach
  - Treating PDA’s > 2.0mm in first 12 hours
  - Other groups target a day 3 assessment which will identify silent PDA’s.

CWH Protocol

- Infants < 28 weeks’ gestation will have an echo between day 1-3 depending on index of suspicion, hypotension and respiratory factors.
- Infants 28 weeks or more will have an echo based on clinical and respiratory features and index of suspicion eg. no antenatal steroids, hypotension, need for ventilation

Treatment

- Will be decided by the consultant after consideration of contraindications to indomethacin
- Various Indomethacin courses can be used. A Cochrane review has shown that there is no benefit of prolonged course (ie: 6 days) over shorter courses (ie: 3 days)
- For infants < 1kg 0.1mg/kg/dose 24 hourly for 3 – 6 doses with an echo after the 3rd dose to assess the need for continuing treatment.
- For infants > 1kg or on a second course the dose is usually 0.2mg/kg 24 hourly for 3 doses.
- A single course of Paracetamol can be considered if there are contraindications to using indomethacin or after a failed indomethacin course within the first month of life. Refer to the drug profile for more details.
Contraindications to Indomethacin

- Duct dependent cardiac lesion, pulmonary hypertension
- Platelets < 80 x 10^9 /l
- Abnormal bleeding, necrotising enterocolitis
- Renal failure eg: oliguria < 0.5ml/kg/hr, creatinine >120mmol/l.

Surgical Closure of PDA

Surgical closure of a PDA may be required if the duct has not closed after two courses of indomethacin and the baby remains ventilator dependent.

When the decision has been made for surgical repair of a PDA it is the Neonatal Team’s responsibility to prepare the baby appropriately for surgery. This will include:

- Liaison with the surgical team for the date and time of the procedure
- Liaison with the anaesthetist for any pre-op requirements
- A current group and hold
- Intubation, ventilation and stabilisation prior to surgery
- Ventilate on a machine with the capacity to deliver high frequency ventilation
- Arterial line access
- Dobutamine infusion prepared and commenced after d/w the Neonatal consultant and/or anaesthetist

Follow-Up

- Any baby who has a PDA that remains open at discharge needs Paediatric follow-up
- The PDA may have been detected on a heart scan at discharge or be a persisting murmur in a baby who has previously had a heart scan confirming a PDA
- Arrange for follow-up by the lead SMO in the Paediatric Outpatients clinic
- Note that all babies <32 weeks will already be followed up so it is babies 32 weeks and older where follow-up will need to be specifically considered

Hypotension

Hypotension is defined as a mean systemic BP < GA in the first few days.

BP increases with postnatal age so that by day 3 the MAP is usually GA + 5-10 mmHg.

Causes of Hypotension

- Hypovolaemia
- Anaemia
- Haemodynamically significant PDA
- Sepsis
- Left ventricular dysfunction
- Adrenal suppression

Initial management

- Assessment of fluid balance - urine output, tissue turgor, venous return, serum sodium concentration (in the first few days).
- Hypotension is more frequent in extremely premature infants when insensible fluid losses are high, or third space losses are high ie: RDS, surgical neonate, NEC.
- First line of management is a bolus of normal saline (0.9%) 10ml/kg over 30 – 60 minutes, depending on clinical situation.
- Depending on the response clinically and the cause of the hypotension, a second bolus may be given.
- Second line of management are inotropes
• Dopamine
  - Commence at 5 to 10 mcg/kg/min, titrate to response.
  - Remember with low infusion rates that the dead space of the lines (usually UVC) means sufficient dopamine for a clinical response may take 30 – 60 minutes to reach the infant’s circulation.
  - Effective in septic shock where the peripheries are vasodilated
• Dobutamine
  - Helpful in situations where the cardiac output is low due to poor cardiac function and the baby needs inotropic support as well as blood pressure support ie: extreme prematurity, PPHN

Investigations
• FBC
• NEON
• Blood Cultures
• Echocardiogram to assess PDA and cardiac contractility

Management of Resistant Hypotension
• In babies with an adequate Hb (> 120g/L), a PDA closed or on treatment, who have persistent hypotension on high dose dopamine (15-20mcg/kg/min) +/- dobutamine, treatment with hydrocortisone may be considered by the neonatologist.
• In a RCT dopamine and hydrocortisone were equally effective in increasing MAP. What is not clear is which babies benefit the most from hydrocortisone.
• Hypotension is less common in neonates whose mother received a complete course of antenatal steroids so history of this is important.

Supraventricular Tachycardia

Presentation
• 30-40% of SVT presents in the first few weeks after birth.
• The majority of SVT in neonates is due to a re-entrant rhythm (extra electrical connection)
• Often asymptomatic and tolerated for many hours
• Tachypnea, poor feeding, pallor, sweating, irritability, lethargy

Non-Pharmacological Management
• Only use this in patient who are haemodynamically stable and not in shock
• Cardiorespiratory monitoring in place
• Explain the procedure to the parents
• Wrap arms in a towel and immerse the whole face (including mouth and nose – no need to occlude these as the baby will hold its breath) in an ice water slurry in a bowl/bucket for 5 seconds (there is an ice machine on Birthing Suite)
• Reverts SVT in 90% cases
• If unable to dip baby into the water (often the case in NICU) place an ice cold facecloth (ie: a facecloth dipped into a bowl of water and ice) to the entire face covering the eyes, mouth and nose for 10 seconds
• The clinical situation when the SVT occurs and the baby’s stability and tolerance to the SVT will determine when ice treatment is used and when medical staff need to be notified and will therefore be individualised.
• The medical team should be called immediately to assess the baby for the first episode of SVT. After this the medical team needs to clarify how long the period of SVT should be tolerated before ice treatment is to be used and when to call the medical team back for review

Pharmacological Management
• If ice has not been successful or contraindicated due to shock
• IV access in as large a vein as possible – antecubital fossa is preferred
• Place a 3-way tap
• Adenosine rapid bolus through one port then a rapid bolus of saline flush up to 5mL
• Adenosine temporarily blocks the AV node so asystole will be seen briefly before reverting to sinus rhythm
• Record the rhythm strip when giving adenosine
- Dose at 100mcg/kg then 200mcg/kg then 300mcg/kg and if still unsuccessful call the on-call Paediatric Cardiologist in Starship for further advise about adenosine doses (up to 500mcg/kg) or other drugs (amiodarone)

**Electrical Cardioversion**
- Treatment of choice if the baby is in circulatory shock and there has either been no response to iv adenosine or iv access has not been obtained
- This is rarely required and we have little experience with cardioversion
- A decision needs to be made at the time regarding stabilisation with intubation/ventilation and sedation with morphine and midazolam boluses (suggest 100mcg/kg dosing if airway is secured).
- If the baby is conscious they will need sedation and to do this safely the airway needs to be secured
- If the baby is unconscious then the time taken to intubate needs to be weighed up with stability of the baby
- Defibrillator is on Level 5 postnatal ward
  - Open the door on the right side of the machine
  - Press the ON button
  - Place the Paediatric electrodes over the apex of the heart and the second one either on the back or right upper chest
  - Connect the electrodes into the socket coming from the machine
  - Press the ANALYSE button
  - Press the SYNC button to make it a synchronous shock
  - Press the down arrow on the ENERGY SELECT Button (it defaults to 200J – adult shock)
  - The ENERGY SELECT button goes as low as 2J then in one joule increments to 10J
  - Aim for DC shock of 1J/kg and repeat at 2J/kg if unsuccessful
  - Press CHARGE button
  - Press RED button to deliver the shock

**Ongoing Management**
- Take a 12 lead ECG once back in sinus rhythm
- Half of the babies will have recurrence of SVT in infancy but 90% will have stopped by 1 year of age
- Consult Paediatric Cardiologist to discuss the need for ongoing medication and follow-up

**Right Sided Aortic Arch**
- Antenatal scans may diagnose a right sided aortic arch with the possibility of a vascular ring
- Right sided arches occur in 0.1% of the population and vascular rings are even less common
- A right sided aortic arch exits the heart and curves to the left but behind the trachea and oesophagus
- The branches of the aorta can run anteriorly to the heart or there can be a double aortic arch and therefore there can be blood vessels behind and in front of the trachea and oesophagus forming a vascular ring
- Vascular rings may be classified as complete or incomplete
- After birth the baby needs to be examined as part of their normal 24 hour newborn check
- If they are asymptomatic then the parents need to be informed of the potential symptoms of a vascular ring and given the parent information sheet (G:drive/Neonatal Handbook/Other Guidelines – print in colour, or copies in the parent information cupboard in NICU) and a short discharge letter needs to be sent to the GP
- An echo does not need to be arranged postnatally
- Further investigations would only be required of the baby became symptomatic from a vascular ring and echo is not the investigation of choice
- If a baby is born at a primary birthing unit then arrange for them to be seen in the CWH Wed clinic so that we can examine the baby and provide the necessary information

**Symptoms of a Vascular Ring**
- Complete vascular rings are more likely to be symptomatic than incomplete
- Symptomatic children will usually present before a year of age and rarely in the neonatal period
  - Respiratory: stridor, wheeze, cough, recurrent chest infections
  - Oesophageal: difficulty swallowing food gets stuck, choking, vomiting, regurgitation of food worsening of respiratory symptoms when eating
Screening for Long QT Syndrome

- This is for well newborns where a parent has been diagnosed with long QT syndrome
- Long QT syndrome is an uncommon hereditary heart condition (1/2000) where there is abnormal electrical activity of the heart with the risk of sudden death
- The commonest form has an autosomal dominant inheritance pattern
- When a baby is born with a confirmed or suspected family history of long QT there is concern around whether the baby has inherited the condition as well
- ECG’s are used to screen for long QT syndrome but they are not always diagnostic
- The best way to rule out long QT in the baby is with genetic testing if the specific mutation has been identified in a previous family member
- Only about 70% of families with LQTS have an identified gene defect.
- If a family have a known gene defect then they may already be known to CIDG (Cardiac Inherited Disease Group) and/or Genetics service and plans may have been made for genetic testing in the baby after birth. However this is always done after or in parallel with cardiological assessment.

Plan after Birth

- Request an ECG at 1 week and 1 month of age, and measure the QTc(corrected QT interval) carefully
- In a newborn the QTc should be <.045 sec
- Arrange for urgent review if the QTc is significantly prolonged (>0.49 sec)
- Note that ECGs in the first 3-4 days often have a QTc which are difficult to interpret.
- Each square on the ECG is 40 milliseconds or 0.04 secs. Count the squares/time from the start of the Q wave to the end of the T wave. This time is the QT interval. It then needs to be corrected for the heart rate so calculate the time from one R wave to the next R wave in seconds and calculate the QTc using the formula QTc=QT/√R-R (all in seconds)

- Check with Genetics or CIDG about their input with the family
- Refer to Dr Alex Binfield after the 1 week ECG for ongoing outpatient review, and he will discuss with Jon Skinner or the CIDG coordinator to arrange follow up and management. Jon will usually arrange to see the baby during the first few months.
- The infant should avoid QT prolonging medications until the condition is excluded (refer to Starship Guidelines - https://www.starship.org.nz/for-health-professionals/starship-clinical-guidelines/)
- If the mother is the gene carrier, reinforce the need for her to take beta blockers over the first nine months after delivery, it is a time of increased risk for her
**SURGICAL CONDITIONS**

**Gastroschisis**

**Background**
- Gastroschisis is a congenital defect of the anterior abdominal wall immediately to the right of the umbilicus.
- Small and large bowel protrude through the defect, and occasionally other abdominal organs eg: stomach, gonads
- 10% have a related gastrointestinal abnormality such as bowel atresia
- Other complications include bowel dilatation, ischaemia, and perforation
- Mortality is up to 10%, often related to septicaemia or the complications of gut dysmotility and/or short gut syndrome and parenteral nutrition.
- Reduction of the gut is best performed within a few hours of birth as the infant is at risk of fluid, protein and heat loss from the exposed bowel, and progressive oedema and dilatation of the bowel makes it increasingly difficult to reduce.
- In about 20% of cases reduction is not safely possible at the first procedure and in these a temporary silo is formed.
- Delay in establishing full enteral feeding is common with a median duration of around three to four weeks. TPN is required for most of this time.
- The median duration of hospital stay is around 6 weeks.

**Repair**
- The gastroschisis will usually be repaired in theatre under general anaeesthetic
- Occasionally, the bowel can be reduced in the neonatal unit depending on the condition of the infant and the appearance of the bowel

**Clinical Management**
- Wrap the bowel in cling wrap, supporting it centrally on the baby’s abdomen and avoiding any kinks in the bowel. The wrap decreases evaporative heat loss and prevents the bowel mesentery kinking
- Insert a large bore nasogastric tube (usually 8-10Fr) and place it on free drainage and frequent regular suction. The purpose is to keep the stomach empty of air and fluid.
- Cardiorespiratory monitoring
- Keep the baby warm and monitor temperature.
- Give IV Normal saline bolus 20ml/kg before transfer to theatre
- Monitor blood glucose
- Start amoxycillin and gentamicin
- Maintenance fluids are commenced at 90ml/kg/day and changed to TPN postoperatively
- A urinary catheter is inserted either before or during surgery to measure urine output
- A long line is inserted by the surgical team at the time of the gastroschisis repair in most babies

Babies with gastroschisis can be unstable in the first 24-48 hours after surgery with splinting of the diaphragm, high analgesic requirements, large 3rd space losses of fluid, persistent metabolic acidosis and dehydration.
- Analgesia: During theatre the babies are often given fentanyl. A loading dose of morphine (100mcg/kg) is given prior to commencing a morphine infusion (20mcg/kg/hr or more as required) before the NICU transport team bring the baby back to the unit. Further morphine boluses of 50mcg/kg are given if there are signs of pain.
- Fluids: A Normal saline bolus of 10-20ml/kg is often needed postoperatively as well if they remain hypovolaemic. Boluses of albumin 4% can be used instead of saline if the albumin is low (ie: <25). Nasogastric aspirates of >10mls in 4 hours should be replaced ml for ml with Normal saline over each subsequent 4 hour period.
Omphalocoele

Background
- Omphalocoele (or exomphalos) is a hernia within the umbilical cord where the contents are covered by a membrane (in contrast to gastroschisis where loops of bowels are completely exposed).
- Omphalocoele minor only contains bowel whereas omphalocoele major contains bowel as well as other organs, including the liver.
- Omphalocoeles have a high rate of associated anomalies (50-70%)
- Chromosomal associations are seen in about 30% and include trisomy 13, 18 and 21.
- Beckwith-Weidemann syndrome is seen in up to 10% of cases.
- Cardiac and renal anomalies occur in 30-50% of cases.
- Thorough examination after birth is essential to detect associated anomalies

Repair
- Immediate management includes protection of the defect with cling wrap to lessen excessive heat, fluid and protein loss.
- The baby should be NBM, on iv fluids and antibiotics and have a large bore NG placed.
- The surgical team should be notified as soon as possible after delivery
- Meditherm Warming Mattress should be used in theatre to prevent hypothermia
- Surgical management depends on the size of eviscerated contents, presence of an intact sac and associated anomalies.
- Treatment involves reduction of the herniated viscera.
- Surgical reduction may lead to transient respiratory and cardiovascular compromise, as well as occasionally bowel ischaemia.
- A staged reduction is the best method for large defects.
- A prosthetic silo is fashioned to allow daily reduction of abdominal contents back in to the abdominal cavity over a 5-10 day period
- A longline should be placed in theatre in most cases

Congenital Diaphragmatic Hernia
- This is a potentially life threatening congenital malformation caused by a defect in the diaphragm allowing abdominal contents to herniated into the chest
- 85% of cases are on the left and 2% are bilateral
- Seen in about 1:3000 live births
- Survival rate is 60-80% if born alive
- About 30% have associated anomalies ie: trisomy 13, 18, 21, cardiac, CNS, GU and GI malformations and these babies have a higher mortality rate
- Survival is determined by the severity of the pulmonary hypoplasia and pulmonary hypertension
- Treatment goals are to achieve adequate oxygenation whilst minimise pulmonary volutrauma and overdistension of hypoplastic lungs while maintaining adequate acid-base balance and blood gases

Determinants of a Poorer Prognosis
- Early antenatal diagnosis
- Delivery <37 weeks or <1500g birthweight
- Polyhydramnios (AFI >25cm)
- Liver in the chest
- Hydrops
- Multiple associated anomalies
- Right sided or bilateral lesions may also have a worse prognosis
- Lung to head ratio (the ratio of the area of the contralateral lung to the fetal head circumference):  
  - this is a numeric estimate of the size of the lungs  
  - ratio of <1.0 predicts survival is unlikely  
  - ratio of >1.4 predicts a higher survival rate
- Severe pulmonary hypertension after birth
Delivery

- Antenatal steroids have not been proven to be of benefit beyond 34 weeks gestation but should be considered up to 39 weeks gestation if delivery is by elective caesarean section.
- Planned day time delivery at term is advisable
- Attendance at delivery by Registrar or CNS/ANP and Neonatal consultant
- Neuromuscular blockers such as pancuronium should not be used routinely as there is no data to prove that this improves survival rates
- Elective intubation without prior bag/mask ventilation
- Gastric tube to decompress the stomach and intermittent or continuous suction
- Target preductal saturations of 80-90%
- Aim to minimise PIP to <25cmH20
- Surfactant should not be given routinely in term babies

Admission Procedures / Investigations

- NBM
- Fluid restrict 40-65ml/kg/day
- TPN when electrolytes are stable
- Central double lumen IV access
- Arterial line (preductal/right radial is preferred but a UAC is easier to site initially)
- Transcutaneous CO2 monitoring, pre and postductal saturations
- FBC, group and hold, arterial gas, coagulation profile
- CXR/AXR
- Urethral catheter to monitor urine output
- Head US – look for IVH (contraindication to ECMO)
- Echo –to assess pulmonary hypertension, right ventricular dysfunction, cause of hypotension and to rule out cardiac anomalies
- Renal US (not urgent) – for associated anomalies
- Maintain normal glucose, Ca, Mg, haematocrit, pH, temp so as to minimise pulmonary vasoconstriction
- Maintaining a normal systemic BP for gestational age
- If hypotensive give a 0.9% saline bolus 10-20ml/kg and consider inotropes if no improvement
- Start inotropes if there is hypotension, ventricular dysfunction or poor cardiac output
- Dobutamine is a better first-line inotrope in these babies as it has less effect on the pulmonary vasculature than dopamine
- Milrinone may be beneficial as an inodilator but can also cause systemic hypotension so may need to be given in conjunction with a vasopressor (dopamine, adrenaline, noradrenaline)
- Note that high doses (>10 mcg/kg/min)dopamine can cause increased pulmonary vascular resistance and may be counterproductive
- Hydrocortisone can be used as a second-line inotropic agent
- Give morphine for sedation and pain relief
- Neuromuscular blockade is not to be used routinely but may be required in unstable babies and should be discussed with the SMO prior to administering. Pain scores should be assessed first to ensure pain managment is maximised.

Ventilation

- Improved survival has been shown with gentle ventilation:
  - spontaneous respiration
  - permissive hypercapnia (50-70mmHg)
  - minimal sedation
  - avoidance of paralysis
  - keeping the PIP <25cmH2O or MAP <16 on HFOV, PEEP 3-5cm H20
  - targeting adequate pre-ductal saturations (85-95%) and post-ductal >70%
- Conventional ventilation is used initially but consider HFOV if needing to use higher PIP with conventional ventilation
- Do not use recruitment manoeuvres and avoid overdistension of the lungs which is easy to do
- If you are unfamiliar with HFOV then ask the Consultant for guidance about any changes to ventilation
Pulmonary Hypertension Management

- Inhaled nitric oxide does not always help - should only be used with echo proven pulmonary hypertension (right to left shunting across PFO/ PDA and suprasystemic pulmonary vascular resistance) after optimising lung inflation and LV function
- If inhaled nitric oxide is started use 10-20ppm and if it does not produce a clinical improvement then it should be discontinued
- Use nitric oxide cautiously in the presence of LV dysfunction as the increased preload can cause pulmonary congestion and worsening oxygenation
- Sildenafil can also be used in the setting of severe pulmonary hypertension
- If the ductus shuts in the presence of suprasystemic pulmonary pressures - commencing Prostaglandin E 10ng/kg/min to reopen the duct can reduce right ventricular strain and improve cardiac output
- ECMO is used rarely and only for rescue treatment and would be undertaken only in Auckland

Determinants of Severity

\[
\text{Oxygenation Index} = \frac{\text{MAP} \times \text{FiO}_2 \times 100}{\text{PaO}_2}
\]
\[
\text{Modified Ventilation Index} = \frac{\text{PIP} \times \text{RR} \times \text{CO}_2}{100}
\]

- \(< 6\) 98% survival
- \(6-17.5\) 46% survival
- \(> 17.5\) 0% survival
- \(< 40\) 96% survival
- \(40-80\) 36% survival
- \(> 80\) 0% survival

- Correlation between higher pH and lower CO₂ and increase in mortality

Surgery

- This is undertaken only when the infant is stable.
- Parameters indicating stability – normal systemic BP, preductal sats >85%, oxygen requirement <50%, lactate <3, urine output >1ml/kg/hr
- In severe cases it is performed when there has been improvement in the pulmonary hypertension and more stable ventilation (usually around 2-3 days of age)
- The diaphragm is repaired with sutures or a synthetic patch

\textit{CDH EURO Consortium Consensus 2015: Neonatology 2016;110:66-74}

\textit{Pediatric Pulmonary Hypertension Guidelines – American Heart Association 2015}

Oesophageal Atresia

1. Oesophageal atresia with a distal fistula between the trachea and oesophagus occurs in 85%
2. Oesophageal atresia alone occurs in 7%
3. Tracheo-oesophageal fistula alone occurs in 4% - often late diagnosis
4. Other variants including proximal fistula
Associations
- 50% overall have associated anomalies
- Most common is VACTERL: Vertebral/Anorectal/Cardiac/Tracheo-Esophageal/Renal/Limb
- Also CHARGE: Coloboma/Heart/Atresia choanae/Retarded growth/Genital hypoplasia/Ear
- Trisomy 18, 21 (7% have a chromosomal abnormality)

Diagnosis
- <10% diagnosed before birth with polyhydramnios or absent stomach, rarely see the upper oesophageal pouch antenatally
- Other antenatal anomalies may increase suspicion for oesophageal atresia
- Copious oral secretions unable to be swallowed, excessively “mucousy” baby
- Desaturations and respiratory distress
- Delayed diagnosis with choking with feeds (diagnosis should be made prior to feeding)
- Confirmation of diagnosis if unable to pass a 10G nasogastric tube 10cm beyond the gums in a term baby
- CXR shows the nasogastric tube coiled in the upper pouch at T2-T4
- CXR with bowel gas indicates presence of a fistula
- Gaseous abdominal distension suggests preferential air entry through the fistula and is potentially dangerous

Investigations
- Echo for congenital heart disease and to describe the side of the aortic arch (right in 2.5%)
- Renal ultrasound if urine has not been passed, not required if antenatal scans are normal
- Vertebral or rib anomalies may be seen on initial XRays
- Karyotype/Microarray if baby is dysmorphic
- Spinal US if associated anorectal malformation or low spinal abnormality

Pre-op Management
- If born outside CWH the baby needs to be retrieved in a pressurised plane, low flying helicopter or by road to prevent trapped intra-abdominal gas distending causing splinting of the diaphragm
- 10F gastric tube into the upper pouch on continuous suction
- Regular oral suction to prevent aspiration
- NBM – iv fluids
- TPN later on depending on clinical situation and gestation
- Nurse head slightly up to reduce risk of aspiration of saliva
- Antibiotics – amoxicillin at anaesthesia induction (baby may already be on antibiotics for other indications)
- Ventilate if needed - if ETT tip can be placed beyond the distal tracheal fistula it will reduce the risk of gastric distension. Beware gas entering the fistula causing gastric dilatation
- Meditherm Warming Mattress to go to theatre with baby to prevent hypothermia

Gastric Perforation
- Risk of gastric perforation from air going from trachea through fistula to the stomach which cannot be vented
- Gastric perforation can occur in any baby – ventilated or not, although it is more likely to occur in a preterm infant with HMD on assisted ventilation
- Presents with sudden deterioration – abdominal distension and tension pneumoperitoneum and despite increased ventilation, metabolic acidosis and difficulties with oxygenation occur
- Urgent needle paracentesis of the abdomen to deflate then proceed to laparotomy immediately
- Surgical preference is to prevent this complication by early ligation of the fistula in those with significant lung disease (HMD) with the oesophageal anastomosis occurring at the same time or later on depending on the patient’s stability

Long Gap
- > 4 vertebral bodies as the gap between proximal and distal parts of the oesophagus
- This is usually not associated with a fistula
- Assess the length of the gap, exclude a proximal fistula and place a gastrostomy for feeding
- Repair of oesophagus at a later stage after a period of growth (usually 3 months)
Post-op Management

- Primary repair of the atresia and ligation of the fistula via a right thoracotomy
- Normally no trans-anastomotic NG tube (unless preterm or multiple anomalies)
- Normally no chest drain placed unless there are concerns about the anastomosis
- Start feeds when the baby has shown they can swallow saliva
- Sign on baby’s incubator – do not remove/replace NG, do not suction orally past 7cm
- Antireflux meds will be required in the majority of cases

Complications/Long Term Issues

- Anastomotic leak (5%) can present rarely with tension pneumothorax, but are usually small and heal spontaneously
- Recurrent chest infections from pulmonary aspiration
- Tracheomalacia which causes the typical “TOF cough” and tends to improve with time
- Laryngeal nerve palsy detected by a hoarse cry sometimes seen after repair of an H fistula
- Feeding difficulties – related to oesophageal dysmotility stenosis, GORD, tracheomalacia
- Gastro-oesophageal reflux – treat aggressively to decrease the impact on stricture formation
- Recurrence of fistula is very rare
- Strictures needing dilatation (in up to 20%)
- Scoliosis and chest wall deformities

Bilious Vomiting

- Bilious vomiting is a sign of intra-abdominal pathology and must be taken seriously.
- It could be the first sign of malrotation with volvulus even if the abdomen is not distended and this is a surgical emergency
- Bilious aspirates can occur in the preterm infant on nasogastric feeds with feed intolerance, but, this is completely different from the term baby with bilious vomiting
- Usually babies with bilious vomiting will be on the postnatal ward

Management

- Consider the diagnosis of malrotation with volvulus
- Take an accurate history of feeds and vomiting
- View the colour of the vomit if possible
- Examine the baby for abdominal distension, abdominal tenderness and groin lumps
- Check the blood sugar (if they are obstructed they may not be absorbing enough milk to maintain blood sugars)
- AXR - if abnormal admit the baby to NICU
- If no other definite cause for the bile stained vomiting contact the Paediatric Surgeons, make the baby NBM and start iv fluids and antibiotics (amoxicillin and gentamicin)
- The Surgeons may request upper GI contrast studies to confirm the diagnosis
- If there is a clinical picture of peritonitis surgery will be required after the contrast study.

Bowel Obstruction

A variety of conditions may present with similar signs and symptoms. Antenatal scans, history and careful examination should help in establishing a diagnosis. The higher the obstruction the earlier the vomiting with less marked abdominal distension. The lower the obstruction the later the vomiting and more marked the distension

Presentation

- Abdominal distension
- Tenderness or pain on examination of the abdomen
- Vomiting – often bilious
- Delayed passage of meconium or failure to pass meconium within 48-72 hours
- Maternal history of polyhydramnios may indicate a high obstruction
Differential Diagnosis

- **Duodenal atresia**
  - Commonly associated with Down syndrome
  - Double bubble on AXR (dilated stomach and duodenum)
- **Malrotation with volvulus**
  - An embryological anomaly leaves some small bowel attached by a narrow pedicle of mesentery with increased risk of volvulus leading to ischaemia and bowel necrosis
  - Symptoms can be intermittent as the bowel can twist and untwist
- **Small bowel atresia – ileal and jejunal** (often multiple atresias)
- **Colonic strictures after NEC**
- **Meconium ileus**
  - Antenatal scans may have shown echogenic bowel
  - Associated with Cystic Fibrosis – take a family history
  - Check immunoreactive trypsin result from Guthrie Card
  - May need to do further molecular testing for CF and refer to CF Team
- **Hirschsprung Disease**
  - Delayed passage of meconium with increasing abdominal distension and vomiting
- **Hernia**
  - Examine for groin lumps consistent with a strangulated hernia
- **Anorectal malformations**
  - Often as part of VACTERL association

Management

- Notify Paediatric Surgeons
- NBM, iv fluids or TPN
- Large bore gastric tube (8F) on free drainage as well as regular aspiration to decompress the bowel
- Ventilate if systemically unwell or if abdominal distension may cause diaphragmatic splinting and compromise spontaneous ventilation
- Broad spectrum antibiotics (amoxicillin and gentamicin) with consideration of the addition of metronidazole
- Replace extra fluid losses such as large aspirates or vomits with normal saline - usually replace if >10ml in 4 hours but the baby’s birthweight, subsequent weights and electrolytes all need to be taken into account
- Pain relief – morphine bolus 25-50mcg/kg and an infusion of 10-20mcg/kg/hr if needed
- If not ventilated a morphine bolus of 25mcg/kg and an infusion at 10mcg/kg/hr should not cause apnoea
- If a strangulated hernia is the cause then the hernia needs to be reduced and if this is acheived then surgery can then be arranged electively
- Rectal washouts as directed by surgeons if Hirschsprung disease is a possibility – 10mL/kg 2-3 x a day
- Discuss with surgeons about likely timeframe to establish feeds postoperatively, if delay likely, consider Longline placement in theatre
- If born outside CWH the baby needs to be retrieved in a pressurised plane, low flying helicopter or by road to prevent trapped intra-abdominal gas distending causing splinting of the diaphragm

Investigations

- FBC, electrolytes, blood culture
- Group and Hold (cross match indicated if surgery is planned)
- Blood gas – metabolic acidosis marker of systemic illness and ischaemic bowel
- Coagulation profile if systemically unwell and bowel necrosis a possibility
- CXR/AXR
- Upper GI Contrast: malrotation – duodenojejunal flexure is to the right of vertebrae and lower than normal
- Lower GI Contrast: small bowel atresia / strictures/ meconium ileus / Hirschsprung disease – to identify the transition zone
- Rectal suction biopsy for suspected Hirschsprung disease
  - Done in NICU by Paediatric Surgical Team
  - Sucrose for pain relief although is a relatively painless procedure
  - Histology results in 1-2 days showing absent ganglion cells in the submucosa and increased staining for acetylcholinesterase
Necrotising Enterocolitis

Seen in < 5% of babies < 1500g with a mortality rate of up to 30%.

Unknown aetiology with a multifactorial pathogenesis including gut immaturity, bowel ischaemia, altered bacterial colonisation of the gut and enteral feeding.

Risk Factors
- Prematurity, IUGR
- Abnormal umbilical artery dopplers – absent or reversed
- Enteral feeding
- Formula feeding
- Lack of antenatal steroids
- Polycythaemia
- Term babies with severe hypoxia or cyanotic congenital heart disease

Presentation
This can be an evolving pattern or a sudden fulminant onset
- Bilious aspirates
- Abdominal distension, tenderness, discoloration
- Blood in stool
- Apnoea
- Temperature instability
- Metabolic acidosis
- Hypotension
- Falling platelets, neutrophils and Hb

Bells Criteria
- Stage 1 (suspected)
  - Unwell, apnoeas, aspirates, abdo distension, blood in stool
  - AXR nonspecific changes
- Stage 2 (definite)
  - Acidotic, abdominal tenderness, falling platelets
  - Pneumatosis or portal venous gas on AXR
- Stage 3 (advanced)
  - Hypotensive, DIC, peritonitis
  - Perforation and free gas on AXR

Investigations
- FBC, NEON, CRP, Blood culture, Coagulation profile
- Blood gas for acidosis, lactate, blood sugar (a high sugar is sign of sepsis)
- AXR – supine and lateral
  - Dilated bowel loops with thickened wall
  - Pneumatosis (air in the wall of the bowel)
  - Free gas seen as air accumulating under the liver in left lateral position
  - Stacking of loops on top of each other
  - Fixed bowel loops
  - Gas in the portal venous system seen as “black” air overlying the liver

Management
- Call neonatal consultant if you feel that clinically the baby may have NEC
- Babies developing NEC can deteriorate very quickly and require intubation, inotropes and volume support so it is best to inform the consultant early
- NBM for 7-10 days and large bore nasogastric for bowel decompression
- Start TPN and lipid after d/w consultant
• Broad spectrum antibiotics for 7-10 days to cover Gram +ve ,Gram –ve and anaerobic organisms
  – Vancomycin, cefotaxime and metronidazole are an appropriate regime for neonates
  – Amoxycillin, gentamicin and metronidazole can be used in babies who have not been on antibiotics
• Pain relief with a morphine infusion
• Correct electrolyte imbalances
• Correct anaemia and low platelets according to protocol limits as discussed with the consultant
• Inform surgeons after d/w neonatal consultant (if definite AXR changes call them even if after hours)
• 40% require surgery either due to failed medical management, perforation or a fixed bowel loop
• Consider a contrast study 4-6 weeks later for stricture formation if there are symptoms of bowel obstruction

Prevention
• Breast milk – up to 10x decreased risk of NEC compared to formula
• No feeding regimes have been shown to be better than others to prevent NEC
• Fortification of feeds has not been proven to cause NEC
• Probiotics have been shown to decrease rates of NEC and the most appropriate choice of probiotics and dosing is still being researched

Urogenital

Undescended Testes
• Most undescended testes are evident at birth.
• Descent is unlikely to occur beyond 3 months post-term
• Newborn testes are not retractile
• If undescended testis suspected, refer to paediatric surgery/paediatric urology at 3 months post-term
• Inform the GP and tell the parents that an operation is likely to be necessary if the testis is confirmed to be undescended

Hypospadias
• Presents as a bent penis (chordee), with an incomplete foreskin (dorsal hood) and a urethral meatus in an abnormal position (proximal to where it should be)
• Check that the testes are descended and that the scrotum is not bifid (this may indicate a DSD (intersex) problem
• Refer to the paediatric surgeon/urologist
• Neonatal circumcision contraindicated
• If the abnormality is severe, obtain a renal USS.

Hydrocoele
• Hydrocoele need no treatment unless they persist
• Examine to be certain there is no inguinal hernia
• If hydrocoele alone, reassure parents that these almost always resolve spontaneously,
• Refer to paediatric surgeon if there is diagnostic uncertainty or if they persist beyond 2 years of life

Inguinal Hernias
• Usually appear as an intermittent lump at the external ring in the groin
• Always needs surgical correction
• Paediatric surgical referral before discharge.
Ovarian Cysts

Ovarian cysts diagnosed antenatally

- The vast majority of these are benign follicular cysts and if they contain solid elements consider the possibility of a cystic teratoma
- Cystic masses containing solid elements or symptomatic ovarian cysts should be surgically removed and histology performed
- Septated or debris filled cysts usually mean that the ovary is already dead from torsion and ischaemia and these frequently disappear on follow up ultrasound scans
- All suspected ovarian cysts should be referred to the Paediatric Surgeons
- An early postnatal scan should be arranged – discuss timing with the surgeons and arrange follow-up in the surgical clinic after the scan
**ORTHOPAEDICS**

Developmental Dysplasia of the Hips

**Risk factors for developmental dysplasia of the hips (DDH)**

- Breech or transverse lie in the 3rd trimester
- First degree relative with history of DDH
- Severe oligohydramnios of long duration
  - These infants should have a hip ultrasound at 6 weeks after their due date regardless of a normal hip examination (scans done before 6 weeks are hard to interpret due to immaturity and need to be repeated so details such as the EDD are crucial to get the correct timing of the scan)
- The form includes information for the parents to understand why the scan is being requested and the follow-up required if the scan is abnormal
- Give the other copies of the form for the GP and LMC to the Neonatal ward clerk to distribute
- Details such as EDD, phone number and GP name are essential on the US request form
- All results are reviewed by the Orthopaedic DDH coordinator so ensure all referrals have Team DDH CDHB as the Responsible Clinician
  - Borderline results involve arranging a repeat hip US in 6 weeks
  - Abnormal results are referred on to Orthopaedics for a clinic review within 4 weeks
  - If you are contacted by LMC’s with a baby needing hip dysplasia screening please advise them to contact the DDH team directly by email: ddh@cdhb.health.nz or ph: 021 951 261. The Neonatal team should only request hip US scans on CWH inpatients.

**Abnormal Hip Examination**

- Hips that are dislocated or dislocatable or there is a major risk factor such as neural tube defect or arthrogryposis
  - Discuss the clinical findings with the neonatologist on service who may also examine the baby
  - After discussion with the neonatologist refer to orthopaedics by completing the Urgent DDH Referral form (found in Neonatal Forms under the letter “R” on the Neonatal Intranet). Click “full screen” and then “Open in Word” to edit the form and then print (do not save a copy). Email a scanned copy of the form to the DDH Coordinator at ddh@cdhb.health.nz, and to the NICU secretary nicuadmin@cdhb.health.nz to upload on to HCS.
    - The Coordinator can also be contacted on 021951261 if you have any questions
    - There is no need to phone the on-call Orthopaedic Registrar and no need to order an ultrasound as orthopaedics will arrange that after assessing the baby.
  - The referral will be reviewed and booked within 3 days and an appointment made within 2 weeks
  - If you are contacted by LMC’s with a baby with an abnormal hip examination please advise them to contact the DDH team directly by email: ddh@cdhb.health.nz or ph: 021 951 261.

**Talipes**

**Positional talipes**

- The foot can be brought into normal anatomical alignment when manipulated
- No need for Orthopaedic review but notify the Neonatal Physio (pager 5119 or referral in pigeon hole) who will review the baby on the ward or contact the family by phone to discuss exercises to perform.

**Talipes calcaneovalgus**

- The foot is pressed against the front of the shin is of no significance and needs no treatment.

**Talipes equinovarus**

- The foot is inverted, supinated and adducted distal to the talus
- Many babies are born with a minor degree of positional talipes equinovarus in which full ankle movements are possible and in these cases no treatment is necessary.
- Fixed talipes can be detected on antenatal scans and an antenatal referral may have gone to the Orthopaedic Paediatric Physio who should have met with the family to discuss the treatment after birth.
• Babies on the postnatal ward with fixed talipes should be referred to the Orthopaedic team by completing a yellow consult sheet (Fax 80806). Please include on this the Mothers NHI as the team may have met the family antenatally. A senior Orthopaedic Nurse will either visit the family in hospital or arrange to see them as an outpatient within 2 weeks to discuss the treatment required. There is no need to call the on-call Registrar.

• Babies in NICU with fixed talipes should have a yellow consult sheet faxed to the Orthopaedic team when the baby approaches term (if born prematurely) as casting will not be done prior to this as it is not developmentally supportive.

• Treatment involves casting from about 2 weeks of age in term babies or around term corrected age in preterm babies. The casts are changed every week for about 6 weeks and then they proceed to a tenotomy and a further 4 weeks of casting before starting to wear John Mitchell boots and bars.

Erbs Palsy

• This is caused by injury to the brachial plexus often after a difficult delivery
• Typically the arm is held limply by the side, medially rotated and the hand in a “waiters tip” position
• Flexion at the elbow is diminished
• Look for associated clavicular and humeral fractures and Horner’s syndrome (ptosis, small pupil)
• Refer to Neonatal Physio (page 5119 or place a referral in her pigeon hole in NICU) who will review on the ward if available and at 2 weeks of age. If the palsy has not recovered then they will refer to the Child Development Service for ongoing follow-up and management

Other Orthopaedic Issues

• Occasionally other orthopaedic issues occur that need to be discussed with the Orthopaedic team, eg: fractured humerus or femur, dislocated joints, congenital skeletal abnormalities.
• If they are non-acute then a consult sheet to Orthopaedic Outpatients is all that is required. This will be reviewed by the Orthopaedic Paediatric Physio in the first instance who will liaise with the available Paediatric Orthopaedic Surgeon and a time will be arranged to review the baby

• If it is acute and within work hours then complete a yellow consult sheet and contact Jan Armstrong (Charge Nurse) on phone 80812 or mobile 0276890189 and she will liaise with the Paediatric Orthopaedic Surgeon available and arrange for the baby to be seen. Faxes can be sent to 80806 or email to orthopaedics@cdhb.health.nz

• If it is an acute issue that is out of hours then call the Acute Orthopaedic Registrar as the first point of contact on 0272222723
**ENT**

**Ear Deformities**

- Deformational ear anomalies occur in 1:100 live births.
- Early external splintage may reduce long term deformity and the need for later surgical correction.
- Neonates with simple deformational ear anomalies should be referred to the plastics registrar (contact by phone via the operator) for consideration of simple splintage and a time will be made to apply ear moulding in Plastics Outpatients.
- This splintage is best applied in the first week of life and can be applied without anaesthetic.
- Follow up will be by Mr Chris Porter in an elective clinic 2-3 weeks later.
- Splintage may be required for 3-6 weeks.
- The equipment will be available in plastics outpatients for application (solder wire + silicone/paediatric feeding tube + steri-strips).

**Cleft Lip and Palate**

**Cleft Palate and/or Lip**

- May need admission but management on the postnatal ward is encouraged.
- All babies to be discussed with the SMO on service who will also need to approve readiness for discharge.
- To stay in hospital until feeding is established or a discharge feeding plan is made. This will take at least 4 days.
- Encourage breast feeding or time at the breast, but breast milk feeding via a “pigeon” or “Haberman” specialised feeding system is often used.
- Referrals needed (also see flow sheet below):
  - Cleft Nurse – works part-time but can act on referrals promptly. May have met the family antenatally.
  - Plastic Surgeon – will visit family on maternity or NICU if requested, otherwise will see in clinic after discharge.
  - Speech language therapist - will assess and provide appropriate specialised bottles. All babies need to be seen by SLT prior to discharge.
  - Lactation consultant – may have met family antenatally, involve early.
- There is a parent information booklet (The Blue Book) available to be given to the family.
- A Child Disability Allowance form should be completed before discharge if the palate is involved and registration for the National Travel scheme and carer support if appropriate.

**Pierre Robin Sequence**

- Pierre Robin syndrome is characterised by micrognathia (small jaw), retrognathia (posterior displacement of the chin), glossoptosis (the tongue falls backwards) and u shaped cleft soft palate.
- The severity of the syndrome, which presents in the neonatal period with upper airway obstruction and feeding difficulties, varies widely.
- Upper airway obstruction presents at, or shortly after birth as a result of retrognathia and subsequent posterior position of the tongue. The airway can be managed in a number of ways from postural nursing with the infant prone, nasopharyngeal airway to tracheostomy.
- Admit to the NICU for saturation monitoring if there is any suspicion of airway compromise.
- Referral pattern as per cleft palate pathway but may also need ENT review.
- Consider investigation/follow-up for other features of (autosomal dominant) Stickler syndrome, especially if there is a positive family history of Robin sequence or eye abnormalities, usually with geneticist referral.
- Babies that benefit from nasopharyngeal airway will have had full assessment, including a sleep study, prior to NP airway insertion. The decision will be made by Consultants with consultation with parents.
- Consider need for cardio-respiratory monitoring at home.
- See Procedures section for instruction on how to insert a nasopharyngeal airway

**Cleft Lip and Palate Pathway**

**At Birth**
Initial Assessment of airway by Neonatal Team

Admit to Postnatal Ward and assess airway and feeding
Admit to NICU if babies has retrognathia and a cleft palate (Pierre Robin) or babies with colour changes or significant feeding difficulties.

Notify Neonatal SMO of all births. Bronwyn Dixon provides all outpatient follow-up

Neonatal Outreach
Will home visit all cases irrespective of the need for NICU admission or not

Neonatal Team
notify all the following services

- Plastic Surgeon
  Kirk Williams
  Mobile: 0211091386
  Phone: 80157
  Fax: 81587
  Referral made by Plastics or Cleft Coordinator if needed
  For strapping of the lip, or, obturator (plate)

- Orthodontics
  Fax: 80246

- Cleft Coordinator
  Michelle Shand
  Mobile: 021589651
  Phone: 81974
  Fax: 80246

- Speech Language Therapist
  Nicki Mason
  Pager: 8698
  Phone: 86165
  Fax: 80107
  For management of feeding and feeding equipment

- Lactation Consultant
  Pager: 5040
  Phone: 85521
  For management of feeding and feeding equipment
Management of Fetal Renal Tract Dilation: *Antenatal* v1.0 Feb 2017 *ANFRD Chart*

**First US Assessment**
- **16-28 weeks**
  - Visible dilation in the first trimester is always abnormal
  - AP RPD < 4mm: +/± central calyceal dilation (no peripheral dilation)
  - No additional findings
  - A1: +/± central calyceal dilation (no peripheral dilation)
  - No additional findings
- **>28 weeks**
  - AP RPD 4mm to <7mm: +/± central calyceal dilation (no peripheral dilation)
  - No additional findings
  - A1: +/± central calyceal dilation (no peripheral dilation)
  - No additional findings
  - A2: +/± central calyceal dilation (no peripheral dilation)
  - No additional findings
- **AP RPD ≥7mm**
  - A3: PLUS any one or more of:
    - Peripheral calyceal dilation
    - Abnormal parenchymal thickness
    - Abnormal parenchymal appearance
    - Dilated ureters
    - Abnormal bladder wall or ureteroceles
    - Unexplained oligohydramnios
  - Material and fetal medicine referral
  - Exit protocol: Fetal medicine referral

**Follow-up US Assessment**
- Reassess using same criteria as First US Assessment pathway
- If any changes, update care plan and consider specialist referral

**A1: LOW RISK**
- Neonatal review, and virtual clinic follow-up
- Maternal and neonatal GP registration
- Initial postnatal ultrasound: 6 weeks. No antibiotics.

**A2: INTERMEDIATE RISK**
- Maternal and neonatal GP registration
- Initial postnatal ultrasound: Day 7 if >15mm +/- bilateral and all at 6 weeks (1-3 months).

**A3: HIGH RISK**
- Maternal and neonatal GP registration
- Consider antenatal paediatric specialist services referral
- Minimum postnatal ultrasound: US at day 7 as determined by FAAC
- Additional US within 24-48 hours after birth if suspected bladder outlet obstruction, oligohydramnios, abnormal parenchyma or worrying clinical presentation such as poor urine output. Clinical assessment drives urgency. Consider catheter placement if US delayed or concern about bladder outflow obstruction.
Management of Fetal Renal Tract Dilation: Postnatal v1.0 Feb 2017

First US Assessment
Timing as per the Antenatal Pathway. ALL scans will be reviewed under the virtual clinic process. Letter to GP will be sent.
Scans performed before 7 days of age may falsely underestimate dilation and should be repeated after 7 days.

N
AP RPD < 10mm
+- central calyeal dilation (no peripheral dilation)
No additional findings

P1
AP RPD 10 to < 15mm
+- central calyeal dilation (no peripheral dilation)
No additional findings

P2
AP RPD ≥ 15mm
+- central calyeal dilation (no peripheral dilation)
No additional findings

P2
AP RPD < 15mm
With peripheral calyeal dilation and/or dilated ureters
No additional findings

P3
AP RPD ≥ 15mm
With peripheral calyeal dilation and/or dilated ureters

P3
Any AP RPD
PLUS Any one or more of:
Abnormal parenchymal thickness
Abnormal parenchymal appearance
Abnormal bladder wall or ureteroceles
Anomalous kidneys, cystic kidney disease, symptomatic child or urinary tract infections

All children with abnormal renal scans require registration with a GP

N
NORMAL
Normal scan after 1 month age:
EXIT PROTOCOL: No further follow-up

N
LOW RISK
Virtual Clinic review, and letter to GP.
Low risk US after 1 month age:
Needs repeat US and assessment at 6-12 months of age. Review at GU clinic

P1
INTERMEDIATE RISK
Virtual clinic review
D7 scans reviewed (see P3)

P2
HIGH RISK
Needs urgent specialist input
Abnormal D7 scans reviewed at NICU scan meeting with Paediatric Surgeons. Confirm 6 weeks tests ordered.

P3
EXIT PROTOCOL
Referral or other management as appropriate

US assessment at 12 months
Use same criteria as first US assessment

If normal at 12 months then EXIT PROTOCOL
No further follow-up

If remains P1 at 12 months old then EXIT PROTOCOL
No further follow-up

If P2 or P3 at 12 months then follow appropriate pathway.
Review at GU clinic will be arranged.

1. Follow up at GU clinic will be arranged.
2. Repeat US 1–3 months
At discretion of responsible clinician, consider:
Prophylactic antibiotics — Radiology prescribe if VUR identified
MCU if bilaterally dilated ureters or calyces
(Mag3 or DTPA after 3 months age if suspicion of obstruction)

1. Urgent referral to local Specialist Paediatric Service. Tests at 6 weeks reviewed by Virtual clinic and GU arranged at monthly GU clinic.
2. Repeat US/other tests as determined by specialist
At discretion of responsible clinician, consider:
Catheter placement if concern about bladder outlet obstruction
Prophylactic antibiotics (Continue if grade 3 or more VUR)
MCU for assessment of reflux or bladder outlet obstruction
Mag3 or DTPA after 3 months age if suspicion of obstruction
Other Renal Issues

The flowcharts above only relate to antenatal renal dilatation, however, there are other antenatal renal anomalies that need follow up:

- Single kidney and Unilateral Multicystic dysplastic kidney:
  - MCU and US at 6 weeks
  - Antibiotics for 2 nights prior to MCU
  - Virtual clinic FU
- Horseshoe kidney, Duplex kidney, Pelvic kidney - without dilatation or dilatation <7mm:
  - US at 6 weeks
  - No antibiotics
  - Virtual clinic FU
- Family History of High grade VUR in parent or siblings ie: first degree relatives:
  - Incidence of VUR is 20-40% if a sibling is affected and 40-60% in offspring of mothers with VUR.
  - Over the years the level of VUR at 6 weeks age when the postnatal renal US was normal is usually grade 3 or less and the need for other than US monitoring has not been required in the majority of cases.
  - US only if no antenatal dilatation and no antibiotics are needed, however after discussion, acknowledge that the family may request an MCU due to their previous experience
  - US and MCU if antenatal dilatation > 7mm, regardless of peripheral calyceal involvement and give antibiotics

Antibiotic prophylaxis

- If an MCU is done < 7 days age give a dose of iv gentamicin prior to the MCU
- Prophylactic oral antibiotics can start from day 5
- Firstline choice is cotrimoxazole (0.25ml/kg/dose at night) and to continue until the MCU result is known
- If the baby is jaundiced then use amoxicillin 50mg nocte for a maximum of 2 weeks and then change to cotrimoxazole otherwise resistant organisms are more likely
- Who should start prophylactic antibiotics by 5 days of age
  - Babies booked for an MCU should be recommended antibiotics.
  - As noted above a single kidney or unilateral MCDK needs antibiotics just for the 2 nights prior to the MCU
  - Renal pelvis dilatation ≥7mm with additional findings (A3 group) and MCU planned.
  - Renal pelvis dilatation ≥7mm without additional findings due to family history and MCU planned
  - Pattern on antenatal scans highly suggestive of VUR eg fluctuating renal pelvis and ureter seen whatever the renal pelvis dilatation measurement is
  - Family history of VUR with early UTI history in other family members and normal antenatal scans - some parents will prefer antibiotics from day 5 depending on their experience. Signs and symptoms of UTI are variable and subtle.
  - Parents are sometimes reluctant for their baby to be on antibiotics – they should be advised depending on the severity of the antenatal findings. If at the mild end of the spectrum commencing 2 nights before the MCU is adequate.

Documentation

- Generic letter to be completed: Health Connect South – Add New Document – New Referral Document – Neonatal Specialty – Document type to get the correct template (Renal or Renal Family History) which explains the process to parents, LMC, GP
- The copy for the Antenatal Renal Virtual Clinic (Paediatric Outpatients) to be given to the NICU ward clerk.
- The US and MCU are ordered electronically. The SMO for the investigations should say R SINCLAIR so that the results go to the appropriate person to act upon
- The requests must have the Maternal NHI written in free text for the Radiologists to be able to view the antenatal scans at the time of the postnatal investigations
Renal Failure

- A presumptive diagnosis of renal failure in the newborn can be made if the urine output is less than 1 ml/kg/hr, lasting for >24 hours and/or the serum creatinine is above 0.09 mmol/l and urea above 7 mmol/l.
- The cause may be prerenal, renal, or postrenal.
- Prerenal - commonest cause usually due to under perfusion of normal kidneys.
- Postrenal - remember to examine the baby to look for bladder distension (urinary retention is common in babies on opiate infusions) and to determine whether the kidneys are enlarged (think of renal vein thrombosis, or congenital malformations)
- Differentiating prerenal from intrinsic renal failure can be difficult but crucial. Renal ultrasound may be helpful. Send a urine sample for analysis for blood and casts (which could indicate an arterial or venous thrombosis), and for culture.
- To distinguish prerenal renal failure from intrinsic renal failure use the following equations as a guide:

Fractional excretion of sodium (FE Na) = \( \frac{\text{Urinary Na} \times \text{Serum Cr}}{\text{Serum Na} \times \text{Urinary Cr}} \times 100 \)

<table>
<thead>
<tr>
<th>Diagnostic Indices</th>
<th>Renal Failure</th>
<th>Prerenal Oliguria</th>
</tr>
</thead>
<tbody>
<tr>
<td>Urine Sodium mmol/L</td>
<td>30 - 90</td>
<td>10 - 50</td>
</tr>
<tr>
<td>UNa/SNa</td>
<td>0.45 +/- 0.22</td>
<td>0.23 +/- 0.14</td>
</tr>
<tr>
<td>UUrea/SUrea</td>
<td>5.78 +/- 2.89</td>
<td>29.64 +/- 17.9</td>
</tr>
<tr>
<td>UCr/SCr</td>
<td>9.67 +/- 3.57</td>
<td>29.24 +/- 15.6</td>
</tr>
<tr>
<td>RFI</td>
<td>11.62 +/- 9.61</td>
<td>1.29 +/- 0.82</td>
</tr>
<tr>
<td>FEBa</td>
<td>4.25 +/- 2.18</td>
<td>0.95 +/- 0.55</td>
</tr>
</tbody>
</table>

Keep in mind that all these equations are useless in babies on dopamine, (which causes the renal indices of intrinsic renal failure regardless of the underlying cause) and diuretics
POSTNATAL WARDS

Transfers from NICU to the Postnatal Ward

This is a guideline and there needs to be an element of flexibility around:

- the acuity of the Delivery Suite, NICU and Postnatal on a daily basis
- the individual clinical situation
- the best situation for the baby and family to avoid separation wherever possible

Communication

- ISBAR form to be completed by NICU staff and to document the expected management on the postnatal ward including the requirement for observations or length of antibiotic course
- NICU staff to contact Postnatal Ward Clinical Coordinator to discuss the potential transfer
- Baby’s NICU red notes folder to transfer to the postnatal ward with the baby and to be returned to NICU after discharge

General

- Maintaining temp 36.5-37.5 in a cot
- If a baby is <2.3kg they will be admitted to NICU at birth, however, if the baby is stable as per the criteria below then discuss on day 3 if the baby can transfer to the postnatal ward to be with the mother. Rare to transfer a baby back to postnatal ward if <2.2 regardless of performance
- Infants who are now well can complete their antibiotic course on the postnatal ward
- Observations will be required 4 hourly for 24 hours if the baby had been on CPAP or oxygen or is on antibiotics. The need for these to be continued past 24 hours to be discussed with the Neonatal team.

Respiratory

- Not requiring oxygen
- Respiratory rate <60/min
- If respiratory rate is 60-70/min but effortless and not impacting on feeding and needing no specific NICU treatment transfer should still be considered
- NEWS score of 1 for respiratory rate 60-70/min can be an accepted variation that needs to be documented in the ISBAR handover and maternity multi-care pathway to highlight that the respiratory rate has been recognised and will be reviewed daily.
- Babies who receive CPAP in delivery suite but this is stopped on or shortly after admission should return to their mothers as soon as possible
- Babies who have short term CPAP/oxygen for 1-2hours and then have transitioned well
  - 2-4 hours of sats monitoring in NICU off respiratory support
  - maintaining sats ≥95% in air (review histogram for the past 2-4 hours off resp support)
- Babies who required CPAP/oxygen for >2hours
  - at least 6 hours of sats monitoring after coming off respiratory support
  - maintaining sats ≥95% in air (review histogram for the past 2-4 hours off resp support)

Feeds and Blood Sugars

- Babies who have short term CPAP <2 hours should have one breastfeed prior to transfer but this may not always be able to occur in NICU depending on the mothers mobility postpartum
- If the baby was on iv fluids/NG feeds these need to have been halved or stopped for at least 6 hours prior to transfer and the baby to have fed twice with 2 pre-feed sugars >2.6mmol/L
- If top ups are required then a specific feeding plan should be documented prior to transfer
NICU Team Prioritisation of Neonatal Reviews on Postnatal Ward

- When covering the postnatal ward, print off a patient list in the morning
- The babies that need a check have NICU beside their name on Flowview
- Make contact with the maternity co-ordinator (pager 5128) and maternity discharge facilitator (pager 5034) on arrival in the ward to discuss the prioritisation of the babies
- The patient list can be printed off Flowview and once a baby has been seen this needs to be updated on Flowview
- There is a handover book that sits beside the patient board on Level 5 and is a place to document any babies that need further review or tests followed up
- It is imperative to use this as a way of communication to maintain continuity of care as there are a number of staff covering the postnatal ward during the week and weekends.
- Midwives or GP’s may contact the postnatal staff member for assistance with organising follow-up hip scan, renal scans and prophylactic antibiotics if the baby was not born at Christchurch Women’s. It is easier for us to arrange the tests and this ensures they will get appropriate follow-up if needed.
- Before referring babies to consultants in other specialities we prefer that you discuss the abnormality you have found with the neonatal paediatrician on call or on service. Referrals to clinics or for investigations should always take place in the context of a full discussion with the parent(s) and notification of the GP and/or LMC.
- Electronically sign off the results of all babies reviewed by the Neonatal Team on the postnatal wards on a daily basis and at discharge

Newborn Early Warning Score (NEWS)

- Introduced 2015 as a quality measure to ensure babies in at risk groups have appropriate observations and a clear process to escalate care in concerning clinical situations
- All observations to be documented on the Newborn Observation Chart (C280106)
- After birth the baby needs their risk category to be reviewed and documented. This will dictate when they require NEWS observations and if oxygen saturations and blood glucose monitoring are also required.
- Refer to document C280106

Newborn Checks

Initial Newborn Check

- 0-2 hours to check for cardiorespiratory transition and for significant congenital anomalies
- Mainly completed by midwives
- Is not part of the resuscitation assessment
- If you do complete the examination fill in the column on the green sheet (QMR0044).

Full Newborn Check (at 24 hours)

- Involves reviewing the maternal notes to check blood and scan results and taking a history from the mother to check for any concerns in pregnancy, family history of newborn problems (heart, hips, kidney diseases)
- Column on the green sheet (QMR0044) for documentation and the Well Child Book
- Registrars and CNS/ANP to measure oxygen saturations on all babies when doing the full newborn check
- Midwives will check oxygen saturations on selected babies as documented in the NEWS
  - 1 and 4 hrs: intrapartum opioid analgesia, severe fetal distress
  - 1 and 4 hrs and prior to transfer: sepsis risk, meconium exposure
  - 12-24 hrs age - <37 weeks, <9th% weight, >98th% weight or infants of diabetic mothers
- Saturations to be checked on either foot until stable and should be ≥95%. If they are not then recheck a preductal reading on the right hand. If this remains <95% then assess and investigate for a cardiorespiratory cause for lower saturations.
Checks that are the responsibility of the Midwife

- NVD, uncomplicated instrumental deliveries
- Caesarean sections (all categories) where the Newborn Observations are normal and there are no additional concerns
- Babies of mothers with diabetes – refer for Neonatal input if there are blood sugar issues as per the guideline
- Breech deliveries - Neonatal team to be consulted if there are concerns about unstable hips for a second opinion and to ensure the hip referral forms are completed but the Neonatal team do not need to complete the full examination and this can be done by the midwife prior
- Babies briefly in NICU for <4 hours who have normal Newborn Observations
- LMC’s are responsible for ensuring that the initial and 24 hour check are completed
- If a midwife is not confident with performing this examination they should seek support from their midwifery colleagues or Neonatal Service ward Reg/CNS who can do the assessment with them.
- They can also seek further training from the midwifery educator at a convenient time.

Checks that are the responsibility of the Neonatal Team

- Antenatal consultation with the Neonatal Team
- Preterm delivery <37weeks
- Congenital abnormality
- An LMC or core midwife can request a review by the Neonatal Team at any time if they have any concerns such as respiratory distress, abnormal exam findings, unstable hips, murmur, antenatal anomalies that need follow-up
- Babies admitted to NICU for >4 hours.
- Babies admitted to NICU for <4 hours should be reviewed by the NICU Team but the full baby check may be able to be done by the LMC if there are no ongoing concerns.
- Babies born to mothers with complex mental health issues where it has been identified antenatally by the Mothers and Babies team that Neonatal review would be beneficial

Transfers

The Neonatal Team is often asked to check that a baby is well enough for transfer either to home or a primary birthing facility. For this to occur the following needs to be clarified:

The initial check has been completed and documented by the LMC or midwife

- The baby has had a normal temperature (36.5 – 37.5) recorded between 1-4 hours of age
- The baby has fed well on one occasion as this is a good sign of wellness
- The baby has been reviewed to ensure that the cardiorespiratory status is stable and the baby has transitioned normally
- Remember that babies 37 weeks and 9-25% may need longer before transfer.
- Prior to transfer to a primary unit before 24 hours of age a baby with risk factors must have a repeat NEWS of 0

Babies who CAN Transfer LESS THAN 6hrs age if NEWS = 0

- Vaginal delivery with no risk factors
- Non-complex instrumental delivery (see definition on Newborn Observation Chart)
- Intrapartum analgesia or GA
- Maternal GBS/PROM/Sepsis risk and antibiotics given > 4hrs before birth

Babies who CAN Transfer FROM 6 hrs age if NEWS = 0

- Maternal GBS/PROM/Sepsis risk and no antibiotics or antibiotics given < 4hrs before birth (need for ongoing 4 hourly observations until 24 hours of age)
- Thick meconium, or thin meconium with Apgars at 5 minutes < 9 (need for ongoing 4 hourly observations until 24 hours of age)
- Weight > 98th% with no maternal diabetes require 3 normal blood sugars before transfer
- Maternal GDM diet controlled require 3 normal blood sugars before transfer
Babies who CAN Transfer FROM 24 hrs age if NEWS = 0

- Intrapartum fetal compromise (to stay one day even if the lactate normalises at 3-4 hours). If there is a request to transfer or discharge prior to 24 hours then the whole clinical picture needs to be reviewed with the Neonatal team.
- Type 1 Diabetics or poorly control Maternal GDM must also have had 3 normal blood sugars
- Clinical chorioamnionitis
- Maternal GA (baby may be ready prior to 24 hours so this indication is maternally driven)
- High risk instrumental category – time > 20 minutes, more than 3 pulls, 2 or more cup detachments, Apgar<7 at 5 minutes.

Babies who CAN Transfer from Day 3 if NEWS = 0

- <9th % for weight and ≥ 37 weeks gestation

Babies who CAN Transfer from Day 4 if NEWS = 0

- Premature babies <37 weeks gestation

Infants < 37 weeks or Weight < 9th Centile

- Approximately 40% of babies born at 35 weeks and 70% of babies born at 36 weeks gestation remain on the postnatal ward (CWH audit 2013) and do not require admission to the neonatal unit.
- These preterm or low birth weight (LBW) babies are at higher risk of issues with temperature control, jaundice, establishing feeding, maintaining blood sugars and gaining weight.
- Parents should be informed of the unique characteristics of their preterm or LBW baby. For example, these babies may not wake spontaneously, may not feed effectively and may lack stamina to take adequate feeds
- Consequently closer scrutiny of breastfeeding and protection of lactation by hand expressing and/or electric breast pumping is required to ensure lactation keeps pace with baby’s caloric intake.

These babies require:

- Daily review, whilst inpatient, by the Neonatal Team.
- Neonatal team will perform the 24 hour baby check and document on Newborn Record (QMR0044)
- Standard NEWS observations at 1, 4, 12, 24 hours as well as oxygen saturations once within 12-24hrs and blood sugar monitoring 3 hourly initially
- A blood sugar check prefeed 3-4 hours after birth (combine with lactate if required) and repeat sugar prefeed until there are 3 consecutive levels ≥2.6mmol/L
- Referral and review by the Lactation Consultant team to formulate a feeding plan which will include cue based feeding with no longer than 3 hours between feeds with top-ups of expressed breastmilk (EBM) as available or donor breast milk (pasteurised or unpasteurised if applicable) or infant formula.
- Monitoring input and output that are consistent with postpartum age with clear documentation on Infant Feeding Record (Ref.2400431)
- Weight on day 3 is preferable
- Clearance by the Neonatal Team prior to discharge/transfer
- Recommend that these babies all stay at CWH until after their weight on day 3.
- Ensure a feeding plan is in place
- On day 3 consideration can be made to the mother and baby’s readiness for discharge or transfer after reviewing the whole clinical situation with the following options available:
  1. Stay at CWH for 4 days – mandatory if <37 weeks at birth
  2. Require ongoing oversight but this could occur at a Birthing Unit from day 3
  3. Be ready to be discharged home (least preferred option) but would need a weight prior to discharge on day 3 to ensure that this is a safe decision
- It is recommended that Vitamin D is supplemented (from birth) if <37 weeks or <2500g until 12 months age
- Iron to start from 4 weeks of age if they are breastfed and <37 weeks or <2500g birth weight. This is recommended to continue until 12 months age
- Babies needing Vitamin D and Iron should get a prescription before discharge from the Neonatal Team.
- A discharge letter will be written after final review
Feeding Red Flags

If any consecutive breastfeeds are A-D do a full set of newborn observations and consider recording feeding concerns in NOC/NEWS

Red flags that would trigger a score for feeding concerns include a baby who:

A baby who:

- Is lethargic and too sleepy to feed
- Not had a first feed in the first hour, followed by no feed in the first 4 hours.
- Has an initial first feed, then not waking for a feed within the next 6 hours.
- Regular feeding for 12-24 hours is then not interested for 6 hours. When a baby stops feeding after previously doing well, this can be a sign of early onset sepsis (within the first 48 hours), severe hypoglycaemia, and in rare circumstances intra-abdominal problems
- Has hypothermia, which may also increase the risk of hypoglycaemia
- Babies with risk factors identified, who are not feeding well, have more potential to show early signs of infection or develop hypoglycaemia

If there are feeding concerns, then a blood glucose check may be warranted. Consider the impact of maternal factors – eg; the reason for induction was poor growth at 37/38 weeks. Plus factors that delay lactation.

Some babies who run into difficulty have asymmetric growth and their birth weight plots at 9-25th centile, so are not initially in a NEWS risk group.

It is useful additional information to plot the baby’s birthweight on a customised antenatal GROW chart in the mother’s notes if available.

Management of Babies with Excessive Weight Loss

7-10% Weight Loss

- Observe a full breastfeed
- Ensure effective positioning and attachment
- Observe for effective suckling pattern, observe for milk transfer and use breast compressions throughout the feed
- Ensure minimum of 8 feeds in 24 hours
- Skin contact to encourage breastfeeding
- Observe for change in frequency / amount of urine and stools
- If top-ups are felt to be needed consider half tops ups (15-30mls)
- Reweigh after 24 hours
  - If weight increasing, continue to monitor closely and provide support
  - If no weight gain or further loss refer to the next section below

10-12.5% Weight Loss

- Follow the plan above
- Refer to Lactation consultant and Neonatal Team
- Express breastmilk after each feed and offer to baby
- Feed according to feeding cues with no longer than 3 hours between feeds
- Offer full top-ups (30-60mls) after each feed
- Consider breast milk substitute if inadequate EBM
- Reweigh after 24 hours
  - If weight increasing, continue to monitor closely and wean top ups as able
  - If no weight gain or further loss will need NICU admission

>12.5% Weight loss

- Refer to NICU for admission and management

Discharge Letter Criteria

- Discharge letters are required for the following babies on the postnatal ward:
  - Admission to NICU for > 4 hours prior to transfer to the postnatal ward
  - Received antibiotics
  - Requirement for Vitamin D and/or Iron after discharge (ie: <37 weeks or <2500g)
  - Babies with congenital abnormalities eg Downs syndrome, Cleft lip and palate
    - Check with consultant re need for outreach / discharge facilitation, physio assessment, early intervention after discharge, social work input, Child disability allowance
  - Referrals to other specialties have been made ie: ENT, Paediatric Surgeons, Plastics, Orthopaedics
  - Outpatient investigations have been made (excluding routine hip and renal scans)
  - If any clinic follow-up appointments are necessary
- Copies should go to the GP, LMC, Parents and other specialties involved in the care of the infant - this should be arranged by the postnatal ward admin staff
- If a baby needs follow-up to be arranged then bring a copy of the discharge letter to the NICU Ward Clerks who can facilitate the follow-up appointment. They are used to this process as opposed to the postnatal ward staff
Antenatal Ultrasound Abnormalities

- **Borderline cerebral ventriculomegaly** (ventricles 10-15 mm)
  - Head circumference and careful physical exam
  - Ventricle/s > 10mm – postnatal head US if this is the advice from the Fetal Anomaly Committee
  - Head US as an inpatient has the benefits of rapid resolution of any parental concerns as the majority of these scans will be normal.
  - If the head US is done as an outpatient then arrange for the parents to receive the results – either by phone contact or review in CWH clinic after the scan

- **Choroid plexus separation**
  - Physical exam including head circumference
  - Head US only if ventriculomegaly or structural abnormality (especially of corpus callosum) on later antenatal scan

- **Choroid Plexus cyst(s)** - Physical exam, if no abnormality, no investigation required

- **Isolated partial or complete agenesis of Corpus Callosum +/- absent Cavum Septum Pellucidum:**
  - Antenatal: Fetal MRI AND microarray
  - Postnatal:
    - Monitor blood glucose until 48 hours old, if < 2.6 mmol/L send hypoglycaemia panel immediately
    - Pituitary workup at 48 hours of age: T4, TSH, FSH, LH (inpatient)
    - Ophthalmology review (outpatient)
    - MRI if not obtained antenatally, microarray if not obtained antenatally

- **Isolated absent Cavum Septum Pellucidum after 20 wks GA:**
  - Antenatal: Fetal MRI, consider genetics only if additional anomalies found
  - Postnatal:
    - Monitor blood glucose for 48 hours, if < 2.6 mmol/L send hypoglycaemia panel immediately
    - Pituitary workup: At 48 hours of age, send T4, TSH, FSH, LH (inpatient)
    - Ophthalmology review (outpatient)
    - MRI if not obtained antenatally

- **Fetal cardiac echogenic focus** – physical exam, if no abnormality, no investigation
  Abdominal calcifications
  - Physical exam, if no abnormality, may need no investigation.
  - Consider TORCH screen if not done antenatally.
  - Make sure Guthrie card is done after 48 hours protein feeds

- **Antenatal diagnosis of aortic arch hypoplasia**
  - These cases have a potential arch abnormality detected on antenatal scans
  - A referral will have gone to Cardiology Akld with the scans being reviewed and a plan made that it is appropriate to deliver in Chch
  - Any fetus with a significant and likely duct dependent arch narrowing will have a plan made to deliver in Akld. However, the arch and PDA are dynamic structures so it is not always possible in borderlines/more mild cases to determine antenatally which cases will end up having a clinically significant arch narrowing that needs cardiology input soon after birth or prostaglandin to maintain systemic blood supply and which will have normal anatomy after birth.

Management
- Paediatric attendance at delivery is not required for this indication alone
- Keep baby with their mother on the postnatal ward if well and expect admission for a minimum of 3 days as monitoring and scans are required
- Daily Neonatal Team review and Neonatal Team to do the 24 hour baby check and to include pre and post ductal saturations at this time
- Monitor with newborn observations at 1,4,12 hours and then 12 hourly (including saturations) until the heart ultrasound is performed
- Lactate measurement 12 hourly until the heart ultrasound is done
- Heart ultrasound to be performed by NICU consultant day 2-3 or earlier if unwell
- The baby needs to remain an inpatient until the PDA closes and the aortic arch has been confirmed on scan as patent
- These babies may need a repeat scan at 6-8 weeks so discuss with Dr Alex Binfield (Paediatrician) regarding the requirement of this prior to the baby being discharged
PROCEDURES

If in any doubt, consult more detailed references and/or a more senior colleague. The consultants are proficient in these techniques and expect to undertake appropriate demonstration and supervision.

Procedure Note

For all ‘major’ procedures (arterial lines, central venous lines, intercostal drains, LP) we expect that irrespective of the success of the procedure there will be documentation of the procedure which should include:

- Time and date
- Name of the procedure, Indication
- Documentation of consent, (if needed)
- Antiseptic preparation and sterile drape
- Analgesic or anaesthetic (if any) technique
- Complications (e.g. blood loss), and
- Method used to judge success of the procedure (e.g. radiograph illustrating tip in correct position)
- Comment on position (whether satisfactory or not) and whether line was manipulated to a better position

For longlines, UVC, UAC this is to be completed electronically

- Open the patient on HCS.
- Add New Document/ Click Treatment/Rehabilitation Record/Click on current admission/CDHB CVAD Insertion Form opens
- Complete the form as well as possible but note this is a DHB wide form so not all parts are relevant
- Enter in the free text boxes the line length, tip position after XRay and any alteration to the line so when reviewed it is obvious what has happened to the line after insertion
- A separate form is needed for each line ie: 2 are needed if UVC and UAC are inserted
- The date and reason why the line is removed needs to be entered in when this occurs and that can be entered by the Nurse or Registrar/CNS/NNP
- Finalise the form when completed after the line is removed.
- Update the Level 3 Patient Summary under Vascular Access with the line type, tip position, if this is appropriate and what management plan has been discussed if the line is malpositioned.
- The line type and FINAL tip position is to documented on the central line sticker and placed on the Problem List

Sterile Technique and Skin Preparation

- Use meticulous sterile technique for all procedures.
- Cap, mask, sterile gown and gloves are needed for central vascular cannulation (i.e. umbilical lines and central venous lines).
- Gloves are strongly recommended for all other procedures where there is a risk of significant bodily fluid exposure, e.g. blood drawing and IV insertion
- Careful site preparation is an important step in preventing nosocomial infection. However:
  - alcohol can cause serious delipidation and damage to premature skin do not use alcohol only swabs
  - iodine can be absorbed across neonatal/premature skin in amounts sufficient to cause hypothyroidism so should not be used on the skin
- Never splash preparation solutions around carelessly
- Prepare the smallest area compatible with good hygiene.
- Use drapes intelligently to avoid having to prepare a large area of skin.
- < 28 weeks gestation, < 7 days old - use low strength chlorhexidine (0.1%, blue solution) especially for UAC, UVC and longline insertion to prevent burning to skin from stronger preparations
- < 28 weeks gestation and ≥ 7 days old – use chlorhexidine 2% swabs (alcohol free)
- ≥ 28 weeks gestation at any time - use chlorhexidine 2% swabs (alcohol free)
- Ensure the skin is allowed to dry before commencing the procedure
- Swabs containing alcohol 70% swabs are only to be used to clean iv ports and connections and not on the skin apart from when accessing a ventricular reservoir (see below)
Pain Relief and Sucrose

- Babies feel pain so before a painful procedure always consider analgesic options.
- Controlling a babies discomfort will improve your chances of success in the chosen procedure
- Don’t forget the value in using an assistant to ensure the baby’s comfort which may include swaddling or the use of a pacifier
- If a baby is ventilated this may include an increment of morphine.
- Sucrose is a well studied drug that provides pain relief in babies
- Sucrose is effective for most painful procedures and should be used for the following:
  - Heel prick, im injection, venepuncture, iv line insertion, arterial line insertion, arterial stab, LP, NG insertion, tape or suture removal, bladder puncture or catheterisation, longline insertion or removal, chest drain insertion or removal, dressing change, ventricular tap, ETT suction, echo, eye exam
- See the sucrose drug protocol for instructions on dosing
- Sucrose should be given in the mouth 2 minutes prior to the procedure to be effective
- Contraindications to using sucrose include: postconceptual age <26 weeks, parental refusal, medical paralysis, unsafe swallow, oesophageal atresia, tracheo-oesophageal fistula and metabolic conditions

Capillary Samples

- Remember to use the correct site, i.e. on the lateral aspect of the heel.
- Prewarming the heel may help both collection and validity of results.
- Clean the skin with Chlorhexidine swab.
- Use special heel-prick lancets.
- Blood gases measured from arterialised capillary blood are useful for pH, pCO2 and bicarbonate.

Venous Blood Samples

- "Broken Needle" technique often the easiest
- Use a 21-23 gauge needle specifically designed with plastic attachment or with the hub broken off to drip blood into the tubes
- Or collect (using careful sterile technique) from the hub at the time of insertion of IV.

Peripheral Venous Cannulation

- Wash hands and always wear gloves
- The cannulas most frequently used are 24 guage (yellow) and come in to different lengths
- After 2-3 unsuccessful attempts stop and ask someone with more experience to try (this could be the nurse or CNS/ANP or a more senior Registrar).
- After 4 attempts by 2 people the SMO needs to be informed to decide if they need to try, if a UVC is needed or if the cannula is not required.
- Place some gauze under the hub to protect the skin from injury
- Secure well with steri-strips, Tegaderm™ and a back-board secured with brown tape
- Place a green sticker in the notes with date and time of insertion

Arterial Puncture

- Radial stab - assess presence of the ulnar artery by palpation or by confirming perfusion of the hand when radial artery occluded.
- Fibre optic light is often helpful in the small baby.
- Use 24 gauge needle (either butterfly or straight needle), heparinised by drawing a small amount up and then thoroughly expelling it.
- Consider using 0.1 ml of 0.5% xylocaine without adrenaline subcutaneously for analgesia.
- Insert through the skin at 45 degrees, watching for a flashback.
- Press firmly on the site for two minutes after withdrawal.
Peripheral Arterial Cannulation

Indications
- Measurement of blood gases
- Blood pressure monitoring
- Frequent blood sampling

Contraindications
- Absent or blocked ulnar (if using radial) or radial (if using ulnar) artery
- Previously cannulated ulnar or radial artery
- Pre-existing circulatory insufficiency of the limb
- Local skin infection
- Malformation of the limb

Equipment
- Clean trolley
- Sterile gloves
- Chlorhexidine
- 24G IV cannula
- Heparinised saline syringe
- Extension set and SmartSite™
- Long splint
- Brown tape and Tegaderm™ dressing
- Transilluminator (Cold Light)

Technique
- Clean skin with chlorhexidine
- Suitable arteries to use are the radial or posterior tibial.
- Transilluminate to ensure patency of both radial and ulnar artery.
- Avoid excessive hyperextension of wrist
- Insert 24G cannula/needle slowly at 30 degrees until a flashback is seen
- Advance cannula into the artery
- Connect to a luer lock connector / extension that has a smart site connected with syringe attached,
- Flush with heparinised saline and secure with steri-strips and Tegaderm™
- The nurses will then connect this to the VAMP system and transducer to infusion pump.
- Use 1 U/ml heparin in 0.45% or 0.9% saline run at 0.5-1mL/hr to keep patent.

Complications
- Limb ischaemia, including gangrene of limb
- Blanching of hand or foot
- Skin ulcers
- Emboli
- Local or systemic infection
- Infiltration
Central Line Associated Bloodstream Infection Bundle

- A CLABSI bundle was created to reduce central line infections rates
- The grouping of evidence based interventions which are known to individually improve care, results in significantly greater improvement when they are applied together
- In the NICU it refers to the insertion and care of UVC, UAC and percutaneous longlines

Bundle Elements

1. Five moments of hand hygiene - refer to page 5
2. Standardised sterile insertion pack – still requires additional items (see below)
3. Insertion practice
   a. Use screens and signs – “sterile procedure in progress”
   b. Use hat, face mask, sterile gown provided in the pack
   c. Add sterile gloves, catheters, skin cleansing solutions and instruments
   d. Nurse to remain with the baby throughout the procedure
   e. Clean hands with surgical hand wash
   f. Keep baby warm throughout procedure
   g. Complete instrument inventory when returning items to the hospital aides for cleaning and resterilising as they are not disposable
4. Documentation
   a. Health Connect South electronic form
   b. Insertion sticker for Problem List
   c. Level 3 Patient Summary Sheet
5. Sterile access and line change practices – refer to Central Venous Access Device OPS PPN43
6. Quality improvements
   a. Education to new medical and nursing staff
   b. Audit of insertion documentation
   c. IV link nursing activities

Link to an education session on appropriate placement and imaging of central lines in NICU by Dr Phillipa Depree (Paediatric Radiologist, March 2020)


Umbilical Artery Catheterisation

Indication
- For secure arterial access in an unwell or ventilated baby for blood pressure monitoring and blood sampling

Contraindications
- Evidence of local vascular compromise in lower limbs or buttocks
- Peritonitis
- Necrotising enterocolitis
- Omphalitis
- Omphalocele

Length of Use
- UAC’s are removed when clinically not required and all would be assessed at 7 days to see if there was an ongoing need for the line depending on the patient’s clinical condition
Equipment
- Clean trolley
- Sterile gloves
- Chlorhexidine
- Tape measure
- Sterile central line pack which includes a gown, hat and mask
- Sterile instrument pack
- Cotton tape, suture material
- Heparinised saline (1u/ml) – 0.45% if <1000g and 0.9% for bigger babies
- Single lumen umbilical catheter: < 1000g 2.5 FG or 3.5 FG
- > 1000g 3.5 FG or 5.0FG if for an exchange transfusion

Preparation
- Measure the distance from the umbilicus to the shoulder tip and 1-2 cm in a small baby and 3-5 cm in a large baby. This is equal to the distance required to insert the catheter to a position above the diaphragm.
- Remember to add the length of the umbilical cord from site of insertion
- Alternatively, use the formula or chart below: Internal length for UAC (in cm) = 2.5 x (E.Wt. in kg) + 9.7

<table>
<thead>
<tr>
<th>Birth Weight (kg)</th>
<th>Internal Catheter Length (cm)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>UAC</td>
</tr>
<tr>
<td>0.5</td>
<td>10.95</td>
</tr>
<tr>
<td>1</td>
<td>12.2</td>
</tr>
<tr>
<td>1.5</td>
<td>13.45</td>
</tr>
<tr>
<td>2</td>
<td>14.7</td>
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<tr>
<td>2.5</td>
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<tr>
<td>3</td>
<td>17.2</td>
</tr>
<tr>
<td>3.5</td>
<td>18.45</td>
</tr>
<tr>
<td>4</td>
<td>19.7</td>
</tr>
</tbody>
</table>

- Prepare your equipment as much as possible prior – it is not the nurses job to do this for you
- A screen should be placed around the cot space prior to starting
- One nurse is required to stay with the inserter throughout the procedure
- Open up a central line pack on the procedure trolley
- Put on the cap and mask
- Scrub your arms and hands, putting on a sterile gown and gloves
- A 3.5 gauge single lumen catheter is standard but a 5G may be needed for an exchange transfusion or a 2.5 g in a baby <1000g.
- Flush the catheter and SmartSites™ with heparinised saline 1U/mL
- Have a helper open up the separate pack with the tools on to the trolley
- See above under “Skin Preparation” as to which swabs to use to prepare the cord and surrounding skin
- Drape the baby
- Tie the cord tie around the base of the umbilical cord and loosely knot
- Cut the umbilical cord 1-2cm from the base and tighten the cord tie if it bleeds
Insertion

- Identify the 2 arteries and 1 vein – the vein is patulous and often lies superiorly, the arteries are thick walled and usually smaller. At the abdominal wall level the vein is at 12 o'clock and the arteries at 4 and 8 o'clock.
- There are two techniques
  - either cut the cord aiming to bisect the vessel, or
  - clamp the end of the stump with a forceps and fold the cord stump over a forceps and cut with blade a lateral artedotomy, cutting the artery half way through the wall leaving the rest of the cord attached for the meantime (this is the method we recommend if you are relatively inexperienced because it is often easier and it leaves the other vessels untouched if you are unsuccessful).
- Dilate the artery with fine iris forceps.
- Slow, gentle dilation results in smooth muscle relaxation, whereas, rapid stretch damages the endothelium and promotes spasm. Time spent in ensuring dilation before catheter insertion increases the chances of success.
- Pass the catheter to the previously measured distance, plus the length of the cord from insertion site to base.
- If the catheter doesn't pass with gentle pressure, never force it.
- As you try to get the catheter to pass from the umbilical cord to the intraabdominal umbilical artery (which runs down the anterior abdominal wall, it is often helpful to apply gentle traction on the cord in a cephalad direction (towards the head), so that the catheter doesn't have to make a right angle turn at this site.
- Take samples of the UAC as needed and flush the line and ensure an infusion is started prior to the XRay to prevent the line clotting.

Securing the Catheter

- Secure the catheter with a suture through the edge of the Wharton's jelly, tied with reef knot, then wrapped carefully round the base of the catheter and tied securely again.
- Enclose the catheter and suture in a tape flag.
- Alternatively, attach a small tape "flag" to the base of the catheter and pass a suture through the edge of the Wharton's jelly, knot it and then pass it though the flag and knot again.
- Avoid elaborate architectural marvels with sutures and tape, including "argyle stocking" sutures. They are wonderful hiding places for microbes and rarely improve the security of the catheter.
- In bigger babies, whose skin will stand it and who can be expected to grab and tug on their catheters, consider the use of a simple set of 'football posts' made of zinc oxide tape and attached to the skin on either side of the umbilicus. Use another piece of tape to secure a loop of catheter between the posts.

Appropriate Tip Position

- Satisfactory position is between T7-T9 (preferred), or in a 'low" position at the level of the L3 or L4 vertebra.
- Note areas to avoid are coeliac axis (T12), SMA (T12-L1), renal arteries (T1), bifurcation (L4-5).
- Remove if there is mottling of leg(s) or buttock(s), although discuss with consultant, it may be worth trying some volume first.
**Line Imaging**
- Check position of the catheter with Chest and Abdo XRay - AP and Lateral
- If a UAC needs to be withdrawn to a more appropriate level then a Lateral alone is sufficient as this is also the best XRay to take to review a UVC after manipulation.
- Never insert the line in further after the sterile field has been removed, only withdraw if needed

**Documentation**
- Document the line insertion in Health Connect South (see page 137), Level 3 Summary Sheet and on the sticker for the Problem List
- Chart 0.45% (usually for <1500g babies) or 0.9% saline with 1unit/mL heparin
- Run the infusion at 0.5mL/hr

**Risks of UAC**
- Sepsis is a risk for any central line and needs to be managed with aseptic insertion technique and good quality nursing care after insertion
- Impaired lower limb perfusion may necessitate line removal
- Thrombosis, vasospasm, embolus, haemorrhage
- Malposition and vessel perforation

**Umbilical Vein Catheterisation**

**Indication**
- Central access for parenteral nutrition or infusions in preterm infants ≤30 weeks or <1500g
- Central access for parenteral nutrition for term infants with surgical conditions
- Central access for infusions in term babies with HIE, severe meconium aspiration or PPHN
- Exchange transfusion
- Infusion of hypertonic solutions eg: more than 10% dextrose for hypoglycaemia
• Urgent access at birth for resuscitation using volume expansion or adrenaline
• Unable to site a peripheral cannulae and access needed for antibiotics or fluids

**Contraindications**
• Peritonitis
• Necrotising enterocolitis
• Omphalitis
• Omphalocele

**Length of Use**
• If a UVC is well positioned at the IVC/RA junction then it will be reviewed on day 7 if still in-situ. The majority will then be changed for a longline if central access is still required. If a baby is likely to no longer require central access within a few days time then the UVC may remain in place until no longer needed.
• Note that 50% of UVC’s migrate in or out in the first week and so we need to be keep vigilant about the line placement despite it being initially well positioned. Review the line on any subsequent XRay irrespective of the reason for the XRay. Repeat an XRay if there are concerns that the internal length of the line has altered.
• A UVC that is in a low position after insertion or after manipulation is not well placed and should be removed as soon as possible and within with 48 hours and replaced with a longline or peripheral line if at all possible.
• Remember that TPN can be given peripherally so the risks of using a malpositioned UVC may outweigh the risks of TPN via a peripheral line.

**Equipment**
• Clean trolley
• Sterile gloves
• Chlorhexidine
• Tape measure
• Sterile central line pack which includes a gown, hat and mask
• Sterile instrument pack
• Cotton tape, suture material
• Heparinised saline (1u/ml) – 0.45% if <1000g and 0.9% for bigger babies
• Double lumen umbilical catheter:  
  - < 1000g  3.5 FG
  - > 1000g  3.5 or 5.0 FG

**Preparation**
• Distance can be estimated using standard tables or formulas

  **Internal catheter length for UVC (in cm) = 1.5 x (B.Wt. in kg) + 5.5, or**

<table>
<thead>
<tr>
<th>Birth Weight (kg)</th>
<th>Internal Catheter</th>
<th>Length (cm)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>UAC</td>
<td>UVC</td>
</tr>
<tr>
<td>0.5</td>
<td>10.95</td>
<td>6.35</td>
</tr>
<tr>
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<td>10.1</td>
</tr>
<tr>
<td>3.5</td>
<td>18.45</td>
<td>10.85</td>
</tr>
<tr>
<td>4</td>
<td>19.7</td>
<td>11.6</td>
</tr>
</tbody>
</table>

Remember to add the length of the umbilical cord from site of insertion
• Prepare your equipment as much as possible prior – it is not the nurses job to do this for you
• A screen should be placed around the cot space prior to starting
• One nurse is required to stay with the inserter throughout the procedure
• Open up a central line pack on the procedure trolley
• Put on the cap and mask
• Scrub your arms and hands, putting on a sterile gown and gloves
• Use a 3.5 or 5 French double lumen catheter depending on the size of the baby
• Flush the catheter and SmartSites™ with heparinised saline 0.1U/mL
• See above under “Skin Preparation” as to which swabs to use to prepare the cord and surrounding skin
• Have a helper open up the separate pack with the tools on to the trolley
• Drape the baby
• Tie the cord tie around the base of the umbilical cord and loosely knot
• Cut the umbilical cord 1-2cm from the base and tighten the cord tie if it bleeds

Insertion
• Identify the vein – it is patulous, has a thin wall and is often the vessel that bleeds when the cord is cut
• If the cord is cut at the base then the umbilical vein is found at 12 o’clock
• Hold the Whartons jelly with forceps and gently dilate the vein with the iris forceps
• Place the catheter in the vein opening and gently advance it which should only need slight pressure
• The umbilical vein turns cephalad inside the baby, so gentle caudad traction on the cord may help the catheter turn the corner.
• Catheters that either won’t pass to the distances required or do not aspirate blood are likely to have passed into a branch of the portal vein, rather than traversing the ductus venosus into the IVC.
• If this has happened withdraw the line to a position where blood can be aspirated and the lines flushes easily. Never leave a line in that does not aspirate blood.
• Ensure that an infusion is started through the line prior to the XRay to prevent the line clotting

Securing the Catheter
• Secure as for UAC above

Anatomy
Appropriate Tip Position

- **A UVC in the ideal position** is the tip at the IVC/RA junction which usually correlates to the level of the diaphragm. Only about 50% of insertions will be appropriately positioned on the first attempt. The report will say the UVC tip is appropriately positioned at the expected location of the IVC/RA junction.

**UVC (and UAC) well placed at T8-9 on AP and at the level of the diaphragm on lateral film**

**On AP XRay the line may head straight up or to the patients right but the tip should never end up directed to the patients right but should come back into the midline**

**On lateral XRay the line takes a postero-superior approach along the axis of the stomach**

**Falsely reassuring AP film with UVC appearing to be at the diaphragm. Lateral confirms the low position of the UVC in presumably in the umbilical vein. Highlighting the value of a lateral film**

**Anterior convexity then posterior convexity indicates line has passed through the ductus venosus and tip ends up anterior to the NG tube**

**The angle of the UVC should follow the red line and not the blue line which is too anterior**
• **A UVC that is high** and above the diaphragm is the only situation where you can be certain that the UVC has passed through the ductus venosus
  - Measure the distance from the tip to the level of the diaphragm (often easiest on the lateral) and withdraw the line to this distance
  - If the line is then at the IVC/RA junction then it is safe to leave in and use for 7 days before review assuming that there is no evidence or concern that the line may have migrated
  - If X Rays are taken for any other reason then the tip position should be re-evaluated as up to 50% of lines migrate (in or out) in the first week

![UVC too high in the right atrium – pull back to the level of the diaphragm](image)

• **A UVC that is low** and is below the diaphragm may have the tip in the umbilical vein, portal vein or hepatic vein and are not ideal and need to be recognised and discussed with the SMO.
  - The UVC in this position can be used in the short term however should be preferably removed within 48 hours and a longline inserted or peripheral access used
  - If a longline is unable to be inserted and central access is required then an X Ray every 48 hrs whilst the catheter remains in-situ can monitor for any migration or further malposition. Alternatively an US may help rationalise the tip position and risks associated with ongoing use.
  - Radiology will call NICU if the line is low and the report will say – the UVC tip is projected over the expected location of the umbilical vein/ductus venosus

![UVC is low presumably in the umbilical vein – remove within 48 hours](image)

• **A UVC that is deviating** to the right or left will be in a portal vein, hepatic vein or umbilical vein branch
  - This line must be either removed immediately (preferred option) or repositioned
  - If the line is felt to be required in the short-term then measurement and withdrawal of the catheter into the midline should be done with a repeat X ray
  - The UVC in this position can be used in the short term however should be preferably removed within 48 hours and a longline inserted or peripheral access used
  - If a longline is unable to be inserted and central access is required then an X Ray every 48 hrs whilst the catheter remains in-situ can monitor for any migration or further malposition. Alternatively an US may help rationalise the tip position and risks associated with ongoing use.
  - Radiology will call NICU if the line deviates and the report will say – the UVC tip is projected over the right/left liver, not in the expected location of the umbilical vein or ductus venosus and is therefore malpositioned.
Line Imaging

- Chest and Abdo XRay
  - All UVC’s need an AP and Lateral at time of insertion
  - If the line is withdrawn from a high position above the diaphragm then only a lateral film is required
  - If the line is withdrawn from a low position such as a portal or hepatic vein (ie: has deviated to the right or left) and will remain in for immediate use then both an AP and lateral will be required to ensure that the line is straight as the lateral alone will not be sufficient
- Always discuss the line position with the consultant if it is not in an acceptable position
- Very minimal patient rotation will make it appear that the catheter has moved from target anatomical landmarks
- If there is a radiolucency present over the liver then consider if extravasation may have occurred and investigate with an abdominal ultrasound
- An ultrasound may also be able to help identify the tip placement in the IVC/ductus venosus or inappropriate placement in the heart or liver vessels. This is not standard practice but may be required to assess if a line is safe to remain in-situ and will be dictated by the SMO.
- An US can be performed by the Neonatologist and the images saved and reported on by the SMO if they have the appropriate skill and/or if the scan is needed acutely overnight.
- Alternatively an Ultrasonographer or Radiologist may be asked to perform the US during the day with the Radiologist reporting on the tip position.

Documentation

- Document the line insertion in Health Connect South (see page 137), Level 3 Summary sheet and on the sticker for the Problem List.
- Continuous infusions through a UVC require 0.1U/mL heparin to maintain patency. A spare lumen needs to be flushed with 1mL of 0.1U/mL heparin 6 hourly

Risks of UVC

- Sepsis is a risk for any central line and needs to be managed with aseptic insertion technique and good quality nursing care after insertion
- Thrombosis is a risk but is uncommon. Avoid infusing platelets or clotting factors through a UVC unless there is no other option
• Haemorrhage
• Extravasation in the Liver
  - This occurs when the tip is in a smaller portal/hepatic/umbilical vein branch and is in a deviated/low and/or anterior position on XRay and this is why these positions are not recommended for longer term use ie: more than 48 hours
  - Extravasation usually develops in the liver parenchyma with a fluid filled cyst which can track into the peritoneal cavity as ascites
  - Babies present with abdominal distension and can go on to develop coagulopathy, metabolic acidosis and need reventilation and fluid and blood product support
  - If there is any suspicion of a UVC extravasating then immediate management is to stop all infusions through the line, ultrasound the abdomen and remove the line
• Cardiac Tamponade
  - If the UVC tip is left in the heart the tip can work its way into the myocardium resulting in fluid infusing into the pericardium leading to tamponade - so a UVC should not be left in the heart
  - This is rare and more often seen with longlines but needs to be considered in a baby with acute cardiorespiratory collapse with no clear cause
  - Immediate management is to stop all infusions through the line, ultrasound the heart and remove the line. Pericardiocentesis may be required under ultrasound guidance

Percutaneous Central Venous Lines “Longline”

Indication
• Central access for parenteral nutrition or infusions in preterm infants <32 weeks
• Central access for parenteral nutrition for term infants with surgical conditions
• Infusion of hypertonic solutions eg: more than 10% dextrose for hypoglycaemia
• Prolonged course or antibiotics or antivirals
• NOTE: Longlines can be inserted 7 days a week if there is clinical need - a weekend should not be seen as a barrier to insertion given current staffing levels

Equipment
• Sterile trolley, screen
• Sterile gloves
• Long line - There are 3 types of longlines
  - Silastic 24G catheter inserted via a butterfly in a prepared, sterile package
  - Argon 28G can be used in babies <1000gm or in those with difficult veins, however, the larger size catheter is preferable in bigger babies as it blocks less often and tolerates bigger volumes
  - Double lumen longlines are available and should be considered in babies needing multiple medications. Both lumens are 28G and require a continuous infusion running to keep them patent.
• Central line pack with hat, mask, drapes, scissors, gauze, SmartSites™, syringes
• Sterile iris forceps and straight forceps for holding drapes together (a full instrument pack is not needed)
• Chlorhexidine
• Heparinised saline
• Steri-strips and Tegaderm™ to dress
• Measuring tape to measure distance from entry site to desired tip location

Aiming for optimal tip position

Note measuring correctly is key!
• For insertions in the right arm, measure from the insertion site along the predicted course of the vein, from the elbow crease to the anterior axillary line/armpit, across to the right sternal boarder and down to the level of the nipple line or third intercostal space (As pictured below).
• **For insertions in the left arm**, measure from the insertion site along the predicted course of the vein, from the elbow crease to the anterior axillary line/armpit, across to the right sternal boarder and down to the level of the nipple line or third intercostal space.

• It is likely that your measurement will be 1-2cm longer than the right.

• **For insertions into the leg**, measure from the insertion site along the course of the vein to the level of the umbilicus.

• An audit of final lengths gives the following guide for upper limb catheters into the antecubital fossa.

<table>
<thead>
<tr>
<th>Weight Range</th>
<th>Length Range</th>
</tr>
</thead>
<tbody>
<tr>
<td>500-800g</td>
<td>7-8 cm</td>
</tr>
<tr>
<td>800-1200g</td>
<td>8-9 cm</td>
</tr>
<tr>
<td>1200-1500g</td>
<td>9-10 cm</td>
</tr>
<tr>
<td>1500-2000g</td>
<td>11-12 cm</td>
</tr>
</tbody>
</table>

**Preparation**

• Prepare your equipment as much as possible prior – it is not the nurses job to do this for you

• A screen should be placed around the cot space prior to starting

• One nurse is required to stay with the inserter throughout the procedure

• Open up a central line pack on the procedure trolley

• Put on the cap and mask

• Scrub your arms and hands, putting on a sterile gown and gloves

• There are 3 types of longlines
  - Silastic 24G catheter inserted via a butterfly in a prepared, sterile package
  - Premicath 28G can be used in babies <1000gm or in those with difficult veins, however, the larger size catheter is preferable in bigger babies as it blocks less often and tolerates bigger volumes
  - Double lumen longlines are available and should be considered in babies needing multiple medications. Both lumens require a continuous infusion running to keep them patent.

• Check that the silastic line will pass all the way through the butterfly, by feeding it from blunt end to fine end, then pulling it all the way through. **NEVER** pull it backwards, as you can cut it with the butterfly tip without intending to.

• Fit the metal section into the blue adaptor and screw tightly, the metal should be fully enclosed

• Flush the 24G catheter or double lumen(s) and Short Needle free minibore 14cm extension set (BC2042). with heparinised saline (0.1U/mL) and prefill the butterfly with saline or a yellow cannula if using this with the Argon 28G

• **The 28 G Argon PICC lines** have a fine wire introducer so cannot be flushed prior to insertion.

• Have a helper open up the separate pack with the tools on to the trolley

• If the baby is intubated and ventilated, a dose of systemic analgesic can be appropriate.

• In bigger babies who are ventilated, a dose of muscle relaxant such as pancuronium can also be useful

• See above under “Skin Preparation” as to which swabs to use to prepare the cord and surrounding skin

• Using a gauze square to hold the hand or foot, while you wash from shoulder to wrist or groin to ankle. Continue to hold the hand or foot up in the air while you wrap a sterile drape round the hand or foot and place another one under the limb.
Insertion

- A variety of sites can be useful, but antecubital veins or the long saphenous vein are reliable sites.
- A sterile gauze folded on diagonal can be helpful as a tourniquet if applied gently.
- Make a venipuncture.
- Immediately release the tourniquet, if you are using one.
- Using fine non-toothed forceps, thread the silastic catheter into the vein for the pre-measured distance plus the length of the butterfly (just under 5 cm).
- The hardest part is getting the catheter to insert from the tip of the needle into the vein at about 4cm.
- Patience and perseverance are the keys to success.
- Place your finger on the vein just above the butterfly/cannula to hold the line, then gently withdraw the butterfly/cannula.
- The 28 G Argon PICC lines insert using a 24G angiocath or the peelable butterfly included in the pack.
  - Once in your measured position, pull back the angiocath/butterfly and then remove the guidewire and flush with heparinised saline (0.1U/mL) using Short Needle free minibore 14cm extension set (BC2042).
- If using the larger catheter then feed it back and off the end of the catheter, unscrew the blue adaptor, remove the butterfly then reattach the blue adaptor to the catheter.
- If using a Argon 28G with a yellow cannula then it cannot be removed but pull to the end of the line.
- Gently aspirate, then flush (may not be able to aspirate a 28G line due to its small calibre).
- Ensure an infusion is started prior to the X-ray so that the line does not clot.

Securing the Catheter

- Ensure that bleeding from the site has stopped (this can take several minutes of local pressure with a sterile gauze square).
- Place a Steri-strip close to the insertion site as an anchor.
- Curl the catheter into several loops and secure with steri-strips.
- Place a small piece of gauze under the blue adaptor or cannula to prevent pressure sores.
- The insertion site and the blue adaptor or remaining part of the cannula need to be dressed with Tegaderm™ but ensure it is not circumferential.
- Make sure there are no kinks or places where the line will block off if the baby moves the limb (ie: at he wrist or foot) before placing the Tegaderm™ over the catheter as it is hard to remove once it is put on.
- Provided it has been possible to draw back and flush the line, start a continuous infusion of saline via a T34 pump, while awaiting a radiograph to confirm position of the line. This is to prevent the line from clotting.

Line Imaging

Position verification upper limb

- In all upper limb 28g Argon PICC’S no contrast is required.
- Remove the NG tube for the initial X-ray.
- Take an AP film without contrast.
- For 24G lines an AP film only should be sufficient without contrast.
- If there is doubt on the plain film Water soluble contrast (Omnipaque – stocked in NICU) should be used.
  - Take an AP film after the contrast is inserted. The lumens of most neonatal PICCs accommodate a volume of 0.3 ml or less. Replace the SmartSite™ / bung with a fresh flush.

Position verification lower limb

- For both sized catheters (24 and 28G) a lower limb long line should have an AP and lateral film taken without contrast in the first instance.
- If bowel gas obscures the line a repeat film with contrast will be requested after discussion between the radiologist and SMO.
- All long lines positions are to be re-checked by X-ray following any manipulation.
- Radiographers can place the radiology plate in the incubator tray.
- All line placement to be confirmed with Pediatric Radiology consultant (may also involve registrar).
- Consider rechecking line internal length on X-ray to confirm tip position.
Appropriate Tip Position

Upper Limb Longline
- The ideal location for the catheter tip is parallel to the vessel wall in the superior vena cava, just proximal to the right atrial junction.
- The suggested landmark on the chest X-ray is at the level of the carina or within one vertebral body below carina.
- This location is described as being 1 cm outside the heart in a premature infant and 2 cm outside the heart in a full-term infant.
- Insertion from the right is likely to lie more vertically than from the left due to the angle of the wall.
- A catheter tip with this orientation and in this location decreases the risk of vessel-wall irritation and allows the infusion to enter the bloodstream at the point of highest blood flow, maximizing the diffusion of infiltrates.
- When the line position is shorter – they can be used back to the axilla, but close monitoring for signs of phlebitis is required. A shorter duration of use is expected.

Lower Limb Longline
- The appropriate tip position is in the iliac vein or IVC
- In practice this means that the tip should preferably be above the groin and below the diaphragm

Note:
- Measuring distances on the images are inaccurate due to magnification and other factors, and this needs to be kept in mind when assessing distances for withdrawing lines etc.
- When upper and lower limb long lines are shorter than optimal, they can be used for TPN but use the peripheral formulation and closely monitor for signs of phlebitis. A shorter duration of use is recommend that is < 4 days.
- Shorter than optimal is assessed as follows:
  - Upper limb lines: Tip lateral to the medial end of the clavicle (i.e.: within subclavian or axillary veins)
  - Lower limb lines: Tip inferior to the position of the femoral head (i.e.: within femoral vein)
Radiology reporting
- Radiologist reports will state where the line tip is thought to be.
- Any inappropriate positioning will be reported and phoned to the NICU requester (default is pager 5025), or neonatal SMO.
- After hours line insertion will be reviewed by the inserter with the radiology registrar in the first instance. NICU SMO must be advised so they can review the films. A Paediatric Radiologist SMO is available at all times to review as required (On-call Paed Rad SMO can be called after-hours via switch).
- Given there is still some uncertainty about the ideal position, the radiologists are not expected to advise RMOs/Nurse specialists whether a line is ok to use, this needs to be approved by the NICU SMO.

Documentation
- Document the line insertion in Health Connect South (see page 137), Level 3 Summary sheet and on the sticker for the Problem List. It is important to put in the final measurement if they have been manipulated for auditing in the future.

Risk of a Longline
- Sepsis is a risk for any central line and needs to be managed with aseptic insertion technique and good quality nursing care after insertion
  - Longlines are kept in for as long as they are needed are not routinely replaced.
  - If there are concerns that a longline may be a source of an infection then it should be discussed with the SMO whether it should be removed.
- Extravasation
  - This occurs most often when the longline has been unable to be inserted as far in as possible and the tip is either in the groin or the shoulder. Consider extravasation if the limb is becoming swollen or the pressure on the pump rise significantly. Remove the line.
- Cardiac Tamponade
  - Longlines with the tip in the heart can burrow in to the myocardium and eventually infuse fluid into the pericardium and so a longline should not be left in the heart
  - This is rare but needs to be considered in a baby with acute cardiorespiratory collapse with no clear cause
  - Immediate management is to stop all infusions through the line, ultrasound the heart and remove the line. Pericardiocentesis may be required under ultrasound guidance

References
PICC Placement in the Neonate

Intraosseous Lines

Indications
- Emergency access to circulation when other routes have failed
- Administration of resuscitation fluids, blood products, and medications

Equipment
- 15 G intraosseous needle set
- Intraosseous drill

Procedure
- Clean and drape skin
- Attach needle to IO drill
- Insert needle at 90 degrees to the skin into anterior medial aspect of tibia, 1-2cm below tuberosity (see picture below)
- After reaching bone, squeeze trigger and drill through the cortex
- Remove stylet, place stabilizer over catheter hub, connect extension tubing
- Aspirate to confirm position and flush with normal saline
- Administer fluid or medications as indicated
- Remove as soon as alternative access is obtained
Complications
- Fracture
- Damage to growth plate
- Infection
- Extravasation
- Compartment syndrome

Chest Needle Aspiration
- For emergency aspiration of a suspected pneumothorax possible diagnosed after transillumination
- While setting up for formal chest drain insertion, drain with a 22G butterfly connected via a three way tap to 10 ml syringe.
- Put on gloves.
- Clean the skin with an appropriate antiseptic, then insert needle in the midclavicular line in the second intercostal space, perpendicular to the skin.
- Remember that the neurovascular bundle runs just beneath the inferior border of the ribs.
- Avoid the nipple area.
- You will usually feel a change in resistance as the needle penetrates the pleura.
- Draw back on the syringe, then turn the stopcock and depress the syringe plunger to expel the air.
- Repeat the process as often as necessary, until chest drain is inserted, or until it becomes clear that the pneumothorax is not reaccumulating.
- If you are not experienced in the technique of chest drain insertions, drainage with a butterfly will always suffice until a more experienced colleague can get there to assist you.
- If the baby improves significantly with needle drainage, and you are uncertain as to whether a chest drain is necessary, the end of the butterfly can be put into a small bottle of sterile water to create a temporary underwater drain (that will relieve a tension pneumothorax if it reaccumulates) until the baby's situation can be fully assessed over 10-20 minutes.
- This is also the technique used for emergency drainage of a pneumopericardium, (if there is cardiac tamponade), except that the needle is cautiously inserted just to the left of the xiphisternum, with the tip pointing towards the tip of the left scapula. Draw back frequently as you insert the needle, and have someone keep a watch on the ECG trace on the monitor. The same technique can also be used for emergency drainage of a large tension pneumomediastinum.
- After removing the needle dress with a Tegaderm™

Chest Drain Insertion

Indications
- Pneumothorax in a baby with significant respiratory distress or is ventilated
- Tension pneumothorax
- Pleural effusion

Equipment
- Clean trolley
- Sterile gloves, gown, hat, mask
- Chlorhexidine
- Chest drain (small, medium or large baby)
- Sterile central line pack
- 3 way tap
- 1% Xylocaine and 1ml syringe
Insertion of Chest Drain by Incision – Blake Drain or Atrium Drain

- Position the baby with the affected side up and the arm fully adducted.
- Open a cutdown tray and use sterile gown and gloves, hat and mask.
- Use midaxillary/anterior axillary line.
- Prepare the area with antiseptic - see above under “Skin Preparation” as to which swabs to use.
- Drape the area.
- Remember analgesia i.e. use lignocaine 1%, for all babies and additionally morphine if required and respiratory depression is not a concern.
- Make a small incision (2mm) in skin with a scalpel blade, parallel to the ribs, in 4th to 5th intercostal space.
- Keep well away from breast tissue.
- Use blunt dissection with artery forceps until the pleura is entered.
- DO NOT use the trocar, because of likelihood of perforating the lung or other vital structure (e.g. ventricle).
- It can be a good idea to leave the artery forceps in place to avoid losing the hole through the chest wall and to guide insertion of the drain.
- Alternatively, the fine curved artery forceps can be closed over the end of the drain or inserted into the side port near the end and used to direct the tube.
- Insert drain without trocar into pleural space.
- It is important to aim anteriorly as that where the air will be.
- If the drain is for a pleural effusion then the drain should be aimed posteriorly.
- Ensure that the side hole(s) are inside the pleural space, but be careful not to insert the tube too far as you can damage mediastinal structures (including the phrenic nerve).
- The Blake drain needs to be trimmed before insertion as they are quite long. There is a transition zone that is easy to identify where the drain changes in size and this must be inside the pleural space otherwise the drain will leak.
- Look for humidity in the tube to confirm that it is in the pleural space.
- Suture with single suture through one or both ends of the skin incision, then knotted and tied neatly round the chest tube.
- Do not use purse string sutures as they tend to cause puckered scars.
- Dress with Tegaderm™.
- Avoid heavy dressings that can restrict chest wall movement and obscure surveillance of the site.
- Request AP and lateral chest radiographs.
- Use 5 cm water pressure suction initially.

Insertion of Chest Drain by Seldinger Technique – Cook Pigtail Drain

- Position the baby with the affected side up and the arm fully adducted.
- Open a cutdown tray and use cap, mask, gown and gloves.
- Use midaxillary line if possible.
- Prepare the area with antiseptic - see above under “Skin Preparation” as to which swabs to use.
- Drape the area.
- Remember analgesia i.e. use lignocaine 1% for all babies and additionally morphine if required and respiratory depression is not a concern.
- Attach a 3 way tap and 10ml syringe to the needle and insert the needle and syringe into the 4th to 5th intercostal space mid or anterior axillary line and advance through the chest wall aspirating until air is aspirated and you are in the pleural space.
- It is important to aim anteriorly as that where the air will be.
- If the drain is for a pleural effusion then the drain should be aimed posteriorly.
- Remove the syringe and 3 way tap and attach the guide wire introducer.
• Insert the guide wire through the introducer into the pleural space.

• Hold the guide wire and remove the needle, leaving the guide wire in place.

• Advance the dilator over the guide wire to enlarge the track through the chest wall then remove the dilator.

• Slide the chest drain over the guide wire, advance it through the dilated track until all side holes are in the chest.

• Hold the drain and remove the guide wire.
• Attach the supplied connector to the chest drain and attach to the underwater seal drain tubing.
• The drain does not require any suturing but should be held in place with Tegaderm™.
• Avoid heavy dressings that can restrict chest wall movement and obscure surveillance of the site
• Request AP and lateral chest radiographs. Use 5 cm water pressure suction initially.

**Continuous bubbling** in the underwater seal can indicate that:
• There is a very rapid air leak.
• Try reducing the ventilator pressures (especially PEEP) as blood gases allow, and/or switching to high frequency ventilation.
• Lung was perforated, (other clues to lung perforation are return of blood from the chest tube, suggestive location of tube tip and increased density around the tip of the tube on radiograph).
• Side hole is outside chest wall, (check radiograph)
• There is a leak in the system (check all the connections).

**Failure of the tube to bubble** intermittently can mean that:
• The pneumothorax has resolved (check by transillumination and/or radiograph)
• The tube tip isn't in the pleura (e.g. in the chest wall or mediastinum check radiograph)
• The tube is blocked (it may respond to being gently aspirated or flushed with 1-2 ml of air - do the above manoeuvres first)
The air is loculated and the tube tip isn’t reaching it. This should also be evident from the chest films. The first thing to do is to try repositioning the baby so that the air is near the tube tip, but sometimes reinsertion of the drain is necessary.

**Bladder Ultrasound**

- Unplug the machine and coil up the cords so you don’t run over them
- Remove the brake by pushing the pedal at the bottom to the middle position
- Maneuver to the bedside and plug in
- Turn on the machine by pressing the On/Off button on the left of the machine
- Use the Roller ball to Study Description arrow, click SET, choose Neonatal Abdo
- Move the Roller ball to “Emergency ID” on the right of screen and click SET
- Move the roller ball to START and click SET
- You do not need to enter any patient details as these scans will not be kept
- On the touch screen touch Neonatal General 11MC4 (ignore the numbers)
- On the next screen tap the LINEAR box i18LX5
- Take the large linear probe (straight edges) labelled i18LX5
- Use a single gel packet and put gel on the probe
- Place probe horizontally across the abdomen above the pubic bone in skin crease
- The top of the picture on the machine is where the probe is looking from
- A bladder will be a black round space at the top of the picture
- When finished turn machine off with the On/Off button at the left hand side
- Then move roller ball to SHUTDOWN and click SET
- Wipe the baby’s skin dry
- Clean the probe with the wipes in the basket below.
- Put the machine back in its parking slot, put brakes on and plug it in at the wall

**Bladder Aspiration**

- Use the ultrasound to confirm the presence of urine before attempting an aspiration
- This is the most commonly used technique to obtain a sterile urine sample in a neonate where sepsis is suspected. Urine should be sent for microscopy, culture and sensitivities and group B strep antigen
- Urine should be sent for fungal elements where systemic fungal infection is suspected in extreme preterms
- Urine samples sent for CMV PCR, Gp B Strep Ag or biochemistry need not be sterile.

**Contraindications**

- Recent voiding or dehydration where chances of success are low
- Superficial skin infection over site
- Significant genitourinary abnormality
- Distended bowel or abdominal mass
Equipment
• Sterile gloves
• Sterile dressing pack
• Chlorhexidine
• Ultrasound to check there is urine in the bladder
• Small syringe
• 23g blue needle
• Specimen jar

Procedure
• Explain procedure and indication to parents
• Position infant supine with arms and legs gently restrained by assistant
• Prepare skin with chlorhexidine - see above under “Skin Preparation” as to which swabs to use
• Insert 23G (blue) needle 1-2cm perpendicular plane in midline just superior to pubic bone
• Aspirate
• If no urine obtained retry later when bladder is more distended.

Complications
• Bleeding
• Perforation of abdominal organ
• Infection

Urinary Catheterisation
The ultrasound machine can be used to assess the presence of urine in the bladder prior to catheterisation. See the instructions attached to the ultrasound machine on how to do this.

Indications
• Urinary retention if bladder expression unsuccessful. Consider this in heavily sedated infants.
• Low urine output – no urine in 6 hours, or < 0.5 ml/kg/hour.
• Urethral valves with urinary retention
• Surgical infants who are at high risk of high intra-abdominal pressure postoperatively, ie: gastroschisis

Contraindications
• Abnormal bladder or urethra (except hypospadius)
• Abnormal perineum or ambiguous genitalia

Complications
• Perforation of the urethra or bladder.
• Knotting of catheter, causing trauma on removal (the risk of this is minimised by avoiding use of feeding tubes as catheters and inserting to correct length).
• Possibly increased by using feeding tubes instead of umbilical catheters or Foley catheters

Equipment
• Term: >3kg babies 8Fr or 6Fr Foley catheter (3ml balloon)
• Preterm: <1kg 3.5Fr umbilical catheter, >1kg 3.5Fr or 5Fr umbilical catheter if 6Fr Foley too large
• Only use the Cook catheter which has no introducer
• Chlorhexidine to clean skin - see above under “Skin Preparation” as to which swabs to use
• 10 ml syringe, Urine collection system, Sterile gloves
• Dressing pack, KY jelly
Insertion distance (approximate)
- Term: Male 6cm, Female 5cm
- Preterm: <750g Male <5cm, <750g Female <2.5cm
- The balloon extends 2.5cm back from the tip on the 6Fr catheter and 3cm on the 8Fr catheter – you will get urine draining back before the catheter is sufficiently in the bladder
- If the balloon is to be inflated it is advised that the Cook catheters are inserted to the hub before the balloon is inflated then withdraw to the above length
- If the catheter meets resistance do not inflate the balloon
- As a general rule place a purpose built urinary catheter (Cook catheter)
- Use the smallest catheter possible
- If the 6Fr catheter is too large the next best option is to insert a UAC into the urethra
- Avoid using feeding tubes as these may increase the risk of complications.
- No introducer to be used
- Balloon should not be inflated unless under ultrasound guidance in daylight hours – this is because of prior cases of balloons being inflated within the posterior urethra
- If the baby repeatedly passes the catheter when they empty their bladder despite adequate taping the balloon will need to be inflated – discuss with Neonatal SMO before proceeding. Consideration should be taken about whether these infants still need the catheter

Insertion Technique (Females)
- Place infant supine, with the thighs abducted.
- Put on sterile gloves.
- Separate labia, clean area around the meatus with antiseptic solution using anterior-to-posterior strokes
- Drape the area using a plastic backed paper sheet with central “hole”
- Apply sterile KY lubricant to tip of the catheter.
- With non-dominant hand spread the labia and identify the urethra.
- Gently insert catheter until urine is visible in catheter tubing.
- If catheter is accidentally inserted into vagina, leave in place, insert new catheter anterior to the first
- Connect to closed urinary collection system. Secure the catheter by taping to infant’s leg.
- No catheter balloon to be inflated until after radiology has confirmed the catheter is 3cm beyond the posterior urethra (formal US scan)

Insertion Technique (Males)
- Place infant supine, with the thighs abducted.
- Put on sterile gloves.
- Clean the penis with antiseptic solution starting at meatus and moving down the shaft of the penis.
- Drape the area using a plastic backed paper sheet with central “hole”.
- Apply sterile KY lubricant to catheter tip.
- Stabilise the penis with non-dominant hand, perpendicular to the body.
- Gently insert the catheter into the meatus until urine is seen in the catheter.
- Slight resistance may be felt as the catheter passes through the external sphincter. Hold the catheter in place with minimal pressure – generally spasm will relax after several minutes allowing easy passage.
  NEVER FORCE THE CATHETER – if they are going to be held up it will be in the posterior urethra
- Connect to closed urinary collection system.
- To prevent dislodgement, tape catheter securely to lower abdomen, rather than the leg to help decrease stricture formation caused by pressure on the posterior urethra.
- No catheter balloon to be inflated until after radiology has confirmed the catheter is 3cm beyond the posterior urethra (formal US scan)
Lumbar Puncture

- This is an important procedure and essential to diagnose meningitis in a neonate which usually has few if any localising clinical signs in a neonate.

Contraindications
- Lumbosacral abnormalities
- Skin infection at site
- Respiratory or cardiovascular instability (start antibiotics and delay until stable)
- Coagulopathy (including thrombocytopenia)
- Signs of raised intracranial pressure (decreased level of consciousness, bulging fontanelle, papillary abnormality or other focal neurology) or known non-communicating hydrocephalus or structural abnormality (discuss with consultant regarding need for imaging)
- Actively convulsing

Equipment
- Sterile trolley
- Gloves, gown, hat, mask
- Chlorhexidine
- EMLA patch if LP is not urgent in a term baby
- Sterile dressing pack
- 25 gauge spinal needle for small babies, 22G for term infants
- Sterile LP sample tubes

Procedure
- Wear a sterile gown and sterile gloves, hat and mask
- Remember the inferior end of the spinal cord lies opposite the body of the third lumbar vertebra at birth and with growth it recedes to be at L1-2 in the adult.
- Explain procedure and indication to parents
- Ensure the infant is stable enough to tolerate procedure
- In term infants apply an EMLA patch 45-60 minutes prior to procedure unless this delay is unacceptable.
- If the infant is ventilated consider giving an increment of morphine prior to procedure.
- Local anaesthetic is not routinely used.
- Prepare using an LP kit and scrub for procedure
- Have the assistant position the infant in a lateral decubitus position with back perpendicular to the side of the bed. Be aware that neck flexion may cause apnoea in this age group.
- Prepare skin with chlorhexidine - see above under “Skin Preparation” as to which swabs to use
- Drape the area with sterile drapes
- Palpate the L3-4 interspace (at the imaginary line joining the iliac crests) and use this space or one below
- Insert the LP needle slowly withdrawing stylet when a slight “pop” is felt.
- If resistance is felt stop, withdraw and reposition
- Collect 10 drops in to each sterile pottle and send for microscopy, culture, Group B Strep Antigen, sensitivities, biochemistry and if indicated HSV PCR. Remember to number the samples and indicate to lab if baby has been on antibiotics.

Complications
- Hypoxaemia, apnoea, bradycardia related to positioning
- Headache: difficult to be objective in newborn
- Infection: strict aseptic technique needed
- Nerve penetration: ensure midline puncture and no higher than L3-4
- Traumatic tap: more likely if advance needle too far: an insensitive sign of intraventricular or subarachnoid blood
- Intra-spinal epidermoid tumour: formed if epithelial tissue is introduced into the spinal canal by a needle without a stylet
Ventricular Reservoir Tap

**Indications**

Clinical symptoms of increased intracranial pressure.
- Apnoea, bradycardia, hypertension
- Poor feeding
- Lethargy
- Hypotonia
- Ultrasound evidence of progressive ventriculomegaly.
- Rapid increase in head circumference (>1.5cm a week)
- Tense fontanelle

**Contraindications**

- Low circulating blood volume
- Cellulite or abrasion over reservoir site
- Sunken fontanelle

**Aims of Treatment**

- To decrease progressive ventriculomegaly.
- To allow head growth at a rate of < 1cm per week.

**Equipment**

- 2% chlorhexidine + 70% alcohol swab to clean skin
- 25 gauge butterfly needle or a brown 26 gauge needle
- 5ml syringe
- Standard sterile pack, mask, gown, gloves
- Additional sterile drapes as needed for maintenance of a sterile field

**Precautions**

- Maintain strict asepsis – wear sterile gloves and a sterile gown, hat and mask
- Monitor and correct serum electrolytes every other day if more than 10ml removed daily.
- Be prepared to provide rapid fluid replacement should the infant not tolerate large volumes being removed. Replace fluid removed with intravenous normal saline.
- If skin breakdown occurs, select an insertion site away from broken area.
- Do not place IVs on same side of scalp.

**Technique**

- Consider the use of sucrose for analgesia although at operation the area is denervated
- Place the infant with head in neutral position in anticipation of a 20 to 25 minute procedure.
- Cut any long hair that interferes with the surgical area but do not shave operative area.
- Wear sterile gloves.
- Clean skin with chlorhexidine/alcohol over the reservoir and surrounding skin with a diameter of 4cm.
- Allow to completely dry for 2 minutes.
- Position the sterile drape to maintain a sterile field.
- Attach the syringe loosely to the butterfly needle or brown needle (as you will need to disconnect it throughout the procedure)
- Insert needle through the skin just into the reservoir bladder.
- Select an insertion site different from the one most recently used.
- Angle needle not more than 30 degrees from vertical (see picture above)
- The base of the reservoir is metal so it cannot be punctured.
- Withdraw the required amount of CSF (fluid must be withdrawn no faster than 1ml/kg/minute)
• Once the syringe is full detach it from the needle leaving the needle in place and discard the fluid and reconnect the needle to the syringe.
• Limit the total volume of CSF drained at each tapping to no more than 30ml or 15ml/kg (whichever is less).
• The initial puncture should not exceed 10ml in volume and can be increased on sequential taps at a rate of not more than 5ml/day.
• It is sometimes advisable to tap twice a day at 15mls each time to avoid events.
• Sample CSF for culture, cell count, glucose and protein at least every three days.
• If fluid is blood-stained (from old haemorrhage), biochemical analysis may not be helpful.
• Culture dark fluid at least every three days.
• Remove needle and hold firm pressure for 2 minutes or until CSF leakage from skin stops.
• Repeat drainage at intervals dictated by clinical response +/- ultrasound markers. Repeat once a day but as often as twice daily. Aim to increase daily volume sufficient to prevent progressive ventriculomegaly.
• The volume taken off each day should result in an initial concavity of the fontanelle, with some overlapping of the cranial sutures.
• If the sutures are still overlapping and the fontanelle concave the following day, the interval between aspirations should be lengthened appropriately.
• Follow the response with daily head measurements and cranial ultrasound scans as required.

Complications
• Local skin breakdown
• Hypovolaemia, hypoproteinaemia, hyponatraemia (check electrolytes every 2-3 days)
• Wound or reservoir infection
• Ventriculitis
• CSF leak from puncture site
• Obstruction of ventricular catheter
• May precipitate further haemorrhage if large amounts of CSF removed

Nasopharyngeal Airway Insertion

Equipment
• Pulse Oximeter
• Appropriate sized soft (Ivory) Endotracheal tube, cut to enable fixation.
  – Commonly 2.5mm, 3.0mm, 3.5mm Portex Ivory tubes
• Comfeel base tape x3, Tegaderm, cut to shape.
• Sucrose

Technique
• Ensure that baby has been assessed and will benefit from nasopharyngeal tube placement.
• Attach oxygen saturation monitor if not already present.
• Measure length of NP airway required (distance from lateral nostril to tragus of ipsilateral ear or see chart below).
• Cut ETT to length and longwise to facilitate taping (see pictures).
• A fresh tube (ivory tracheal tube, Portex, UK) is prepared by cutting at the measured length plus 5 cm (fig 1). The tube is then cut down the midline of the underside of the tube to the measured length - this is the inside curve of the tube. Two cuts are made to each side of the midline to create two thin strips 3 mm wide, which are used for anchoring the tube to the child’s cheeks. A further cut is made creating two additional strips on the top. One of these top strips shortened to the measured length plus 2 cm, is used for anchoring the tube to the dorsum of the nose. The remaining top strip is cut off at the measured length. The wedge that is cut off depends on whether the nasopharyngeal tube is used for insertion into the right or left nasopharynx (fig 2). (Masters et al, ADC 1999, 80 (2): 186-7)
• Coat tube with lubricant jelly
• Insert tube into nare along base of nose, you will feel resistance at the back of the nose. Rotate the tube and gently advance around the back of the nasopharynx. Advance forward to the point previously measured. Hold in place.
• Apply Comfeel to above infant’s upper lip on each side and on dorsum of nose. Secure strips from tube to skin with Tegaderm™
• Ensure tube is in the nare comfortably. Care must be taken with edge of cut tube on nare.
• Suctioning should be performed initially after each feed but may be able to be less often
• Change nasopharyngeal airway weekly or sooner if frequently obstructing.
• Consider gentle nasal irrigation with normal saline drops and suctioning if baby appears snuffly.

http://adc.bmj.com/content/80/2/186.full.pdf
(both images Masters et al, ADC 1999, 80 (2) 186-7)

http://www.gosh.nhs.uk/health-professionals/clinical-guidelines/nasopharyngeal-airway-npa#Appendices
IMMUNISATION

Immunisation Schedule and Timing

Table 1: National Immunisation Schedule, commencing 1 July 2017

<table>
<thead>
<tr>
<th>Antigen(s)</th>
<th>DTap-IPV-Eta/ HepB/Hib</th>
<th>PCV10</th>
<th>RV1</th>
<th>MMR</th>
<th>Hib</th>
<th>VV</th>
<th>DTap-IPV</th>
<th>Tdap</th>
<th>HPV9</th>
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<tbody>
<tr>
<td>Brand</td>
<td>Infanrix-hexa</td>
<td>Synflorix</td>
<td>Rotarix</td>
<td>Priorix</td>
<td>Hiberix</td>
<td>Varilrix</td>
<td>Infanrix-IPV</td>
<td>Boostrix</td>
<td>Gardasil9</td>
<td>ADT Booster</td>
<td>Influvac</td>
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<td>Mylan</td>
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<td>annually</td>
</tr>
</tbody>
</table>

6 weeks
- Immunisations should be given to babies as close as possible to 6 weeks postnatally, regardless of birth gestation. See below for exclusions to this.

3 months
- Immunisations to be given at 3 months age (13 weeks, day 91), irrespective of when the initial immunisations were given as long as there has been a minimum of 4 weeks since the 6 week immunisations.

5 months
- Immunisations to be given at 5 months age (22 weeks, day 151). There is a target of this being given not earlier or later than 4 days around 5 months of age.

Practicalities
- There may be reasons to give an immunisation outside the fixed times
- Delayed - until a month after steroids, waiting for a baby to be well enough, waiting for the parents consent, if immunisation will hold up discharge then they may be delayed until a week after discharge to be done at the GP practice
- Early – to ensure discharge is not delayed with a family or a clinical situation where it is better to immunise early and be monitored than risk immunisation not happening in the community or being potentially unsafe

Immunisations
- The discharge letter and plunket book must state what has been given and when
- Immunisations should be given to babies as close as possible to 6 weeks postnatally, regardless of GA
- Immunisation should be deferred if the baby is very small and sick or unstable or is on corticosteroids.
- Discuss with the consultant if in any doubt.
- Some consultants prefer to vaccinate babies when they are still on CPAP to have that respiratory support around the time of potentially provoking apnoeas
• The presence of a progressive brain lesion is the only contraindication to vaccination (pertussis) likely to be of relevance in the NICU.
• IVH and most other intracranial lesions are not contraindications to pertussis vaccination, but if any doubt discuss with a consultant and document specific explanation to parents, for medicolegal reasons.
• From 1 July 2017 the pneumococcal vaccine for infants has reverted back to Synflorix for most babies but with Prevenar 13 for high risk babies
  - < 28 weeks gestation
  - Chronic lung disease
  - Cardiac disease with cyanosis or failure
  - Down Syndrome
  - Renal failure or Nephrotic syndrome
  - Intracranial shunts
  - Primary immune deficiency / HIV
  - Asplenia (anatomical or functional)
• Previous immunoglobulin infusions are not a contraindication to having their 6 week, 3 month or 5 month vaccines (including Rotavirus)
• The Nurse, Registrar or CNS/NNP should discuss immunisations with the family when they are due and the baby is medically stable to have them
• Verbal consent must be obtained and this needs to be documented by any staff member on the multicare pathway. Only then should the immunisations be prescribed
• The immunisations should be given in the presence of the parents if that is their preference
• It is not uncommon for babies to have mild temperature instability, increased apnoea, or irritability.
• It can be difficult to be sure whether these changes are due to the vaccine, or due to sepsis and it may be necessary to do a sepsis work-up, but keep in mind that immunisation can cause temporary elevation of the CRP and ‘left shift’ of the white cells.
• Apnoea monitoring for 48 hours after immunisation is appropriate if the baby is not on any monitoring

Rotavirus
• From 1 July 2014 Rotavirus vaccination was added to the schedule
• From 1 July 2017 the vaccination changed from Rotateq to Rotarix
• Rotarix is a live attenuated vaccine given orally at recommended ages of 6 weeks and 3 months
• The first dose must be given by 15 weeks (Day 104) chronological age and the second must be completed by 25 weeks (Day 174)
• Doses can be given no closer than 4 weeks apart
• Protection probably lasts 3 yrs and provides cover for infants when they are vulnerable to dehydration.
• The vaccine has been shown in Australia to reduce hospitalisation for young children with Rotavirus gastroenteritis by 70%.
• Reasons not to administer the vaccine at 6 weeks chronological age would be if the baby was unable to take the 1.5mL volume orally or if the medical team felt the baby was too unwell/unstable to be vaccinated.
• If the vaccine is not given at 6 weeks of age then administer either at discharge or with the second immunisations (whichever is earliest)
• Standard universal precautions apply when looking after the NICU baby post-vaccination ie: handwashing and using gloves when changing nappies.
• The risk of this causing transmission to nearby un-immunised hospitalised babies is extremely low and the benefits of vaccination outweigh this risk (Ref: Immunisation Handbook 2014, Peds 2014:133: e1555-1560)
• There is a possibility of an increased risk of the rare condition of intussusception after the first dose of the vaccine although not all large data collections have this finding.
• The vaccine has been given in the USA since 2006 with no reported increase in intussusception rates
Live Vaccines

Blood Products
- Live vaccines such as Varicella and MMR vaccine need to be delayed after certain blood products or immunoglobulins have been given (see Immunisation Handbook). Does not apply to Rotavirus vaccine.
  - Hep B Ig – withhold live vaccines for 3 months
  - Varicella ZIG – withhold live vaccines for 5 months
  - Packed RBC – withhold live vaccines for 5 months
  - Whole blood (exchange transfusion) – withhold live vaccines for 6 months
  - Platelets – withhold live vaccines for 7 months
  - IV Immunoglobulin (Intragam) – withhold live vaccines for 10 months

Steroids
- If a baby has received >14 days of high dose steroids such as a prolonged course of dexamethasone or prednisone ≥ 2mg/kg daily or on alternate days (Immunisation Handbook)
  - Delay live vaccines for 4 weeks (MMR, Varicella)
  - Rotavirus immunisation should be deferred for 4 weeks if able but ensure it is given by Day 104 or the baby then becomes ineligible for this vaccine
- If a baby has received ≤14 days of high dose steroids such as a course of dexamethasone or prednisone ≥ 2mg/kg daily or on alternate days
  - Vaccinations can be given as soon as the steroid course is completed however if possible delay live vaccines 2 weeks (MMR, Varicella, Rotavirus as above). (Immunisation Handbook)

In consultation with Tony Walls (Infectious Diseases) our plan in Christchurch is to:
- Delay the 6 week im immunisations for 4 wks after the end of the 10 day dexamethasone course
- Delay Rotavirus immunisation for 4 wks after the end of the 10 day dexamethasone course but give earlier if needed at Day 104 to ensure they do not become ineligible
- Delay the 6 week im immunisations if on prednisone ≥ 2mg/kg until closer to discharge. The reason is because the immune response to the vaccine will be dampened and if this is left to closer to term corrected the baby may either be off steroids by then or the immune response may be better. This is infrequent and can be individualised for the patient’s clinical situation. It is best to get one immunisation in prior to discharge even if still on prednisone
- No need to delay im and oral immunisations if on hydrocortisone for physiological replacement

Congenital Heart Disease
- Live vaccines need to be delayed for 8 months after cardiac bypass.
- This does not apply to the Rotavirus vaccine

Maternal Hepatitis B Carrier (HBsAg positive)
- The risk of mother passing the virus to her baby during delivery is high, and if not infected at birth, the baby remains at risk of hepatitis B infection from mother during the first five years.
- Although the baby’s infection may be mild or even asymptomatic, chronic hepatitis occurs in up to 90% of infants who acquire the virus at birth.
- Thus, infection early in life results in a much higher risk of chronic hepatitis than in adulthood.
- Chronic hepatitis puts the baby, later in life, at high risk of transmitting the virus to others such as sexual partners and offspring, and of death from chronic liver disease or hepatocellular carcinoma.
- Recognition and counselling of mothers who are hepatitis B carriers should begin early in antenatal care.
- All pregnant women should be screened.
- Pacific Islanders, Africans, Asians and New Zealand Maori are high risk groups for chronic Hepatitis B carriage (this is presumed to be mostly due to perinatal acquisition).
- Carriage is also more common in people who have occupational or social exposure to human blood.

Management
- Wearing disposable gloves (to protect yourself) when handling the baby at birth and until they are bathed
- Early bathing of the baby to remove maternal blood and body fluids in warm water only
- Before any im injections the skin is to be cleaned with an aqueous chlorhexidine (alcohol-free) swab
As soon as possible after birth, the infant should receive *Hepatitis B Immunoglobulin* (HBIG) 100 IU IM and *Hepatitis B vaccine* 0.5mL IM at separate sites.

Vaccine and HBIG are likely to be fully effective when given up to 12 hours after birth, and will protect some infants even when given after that, but there is no advantage in delay.

If the father or a household contact has Hepatitis B it is appropriate for the baby to receive the Hep B vaccination at birth but immunoglobulin is only indicated to prevent transmission from the mother during birth.

The baby will need subsequent hepatitis B immunisations as per the National Immunisation Schedule.

Infants of HBsAg positive mothers should be tested for HBsAg and antibodies to HBsAg one and three months after completion of the vaccine series. This will identify those few infants who have become chronically infected despite immunisation and will aid in their long term medical management. It will also identify infants who lack antibody and who should receive further doses of vaccine.

If mother’s HBsAg status is unknown at the time of delivery, maternal blood should be sent for testing.

However, prophylaxis needs to begin immediately to be effective, so if she belongs to a **high risk group**, you should follow the protocol above for infants with know positive mothers without waiting for the results.

If the mother proves to be negative the usual hepatitis B vaccine can be given at 6 weeks, 3 months and 5 months, and the serology testing after completion of the vaccine schedule can be omitted.

We advise immunisation against Hepatitis B for all health care workers who are at risk of exposure to blood or bodily fluids. We also advise obtaining serologic proof that immunity has developed.

### BCG Vaccine

**High rates of TB exist in New Zealand among population groups from Asia, Africa and the Pacific, particularly in recent immigrants.**

The role of vaccination is to protect individuals at high risk of exposure.

BCG was introduced for neonates in 1976 and is effective in preventing extra pulmonary disease.

The LMC needs to assess the risk of TB in pregnancy and plan for vaccination after birth.

In babies admitted to the Neonatal Nursery the Neonatal Staff also have the responsibility of considering eligibility for BCG vaccine.

Preterm delivery or being of low birth weight are not contraindications to vaccination but babies should wait until they are >34 weeks before being vaccinated in order to ensure an adequate response.

If eligible can be referred to the Public Health Nursing service by completing the [referral form](#).

Families should be referred to the Vaccination Clinic at CWH by phoning the Public Health nurses at Burwood Hospital on 99777 (internal) or 383 6863 (external) and faxing a request over 383 6878.


### Neonatal BCG Eligibility Criteria

- Living with a person with current TB or a past history of TB
- Living with one or more people who within the last 5 years lived for a period of ≥ 6 months in countries with a rate of TB ≥ 40 per 100,000
- During their first 5 years they will live for ≥ 3 months in a country with a rate ≥ 40 per 100,000 and are likely to be exposed to those with TB

**Areas with rates of TB ≥ 40 per 100,000:** Africa, South America, Russia, India, China (not Hong Kong), South East Asia (not Singapore), Pacific (not Cook Islands, Fiji, Niue, Samoa, Tokelau, Tonga)
OPHTHALMOLOGY

Retinopathy of Prematurity

- With advances in monitoring and oxygen delivery systems, immaturity is a greater risk factor than oxygen
- All babies with birthweight < 1250g or < 30 weeks should have an ophthalmology referral for screening for retinopathy of prematurity (ROP)
- Babies ≥ 30 weeks or ≥ 1250g may need screening for ROP (SMO discretion) if they have had
  - Hydrops
  - Grade 3 or 4 IVH or post haemorrhagic hydrocephalus
  - Severe sepsis
  - Nitric oxide for PPHN
  - Twin to twin transfusion syndrome
  - Prolonged period of high inspired oxygen

ROP – Patient Identification

- When a baby is admitted and fulfils the criteria for screening they should be entered into the Eye Book in reception and the date for the first eye screen calculated
- From 4 weeks of age check again that the baby is in the Eye Book

ROP Screening – when to start and when to stop

- If < 26 weeks – first ROP screen is at 30-31 weeks corrected gestation
- If ≥ 26 weeks – first ROP screen is 4 weeks after birth
- Timing of further reviews will vary but should not be longer than 2 weeks apart and sooner if there are concerns on the eye examination
- Screening must continue to a minimum of 36 weeks corrected if 28/29 weeks
- Screening must continue until 38/39 weeks if born <28 weeks
- If the regular ophthalmologist is on leave an alternative check will be arranged by the ophthalmology department to ensure no deferments occur.
- If treatment for ROP is required it should happen with 48-72 hrs of diagnosis

Eye Examination Process

- Give parents information prior to the first eye examination so they can be aware of the process and why it is required
- Outreach nurses coordinate the eye clinic and are another check to ensure eligible babies are not missed
- Eye drop stickers are available in the drug cupboards to help prescribing – these need to be put in the notes and signed for prior to being given
- Eye drops are 0.25% cyclopentolate + 2.5% phenylephrine drops – one drop per eye 30-60 min pre-exam
- Sucrose should also be given prior to the examination

Follow-Up after ROP Screening

- ROP screening sheets to be filed under Screening in the clinical notes and uploaded to HCS at discharge
- If a transfer to another centre is planned before 36 weeks corrected, before routine checks are completed or before 38/39 weeks in those < 28 weeks, contact the CHCH ophthalmologist and the accepting unit to identify that an ophthalmologist is contacted and available in that area for ongoing checks or transfer will need to be delayed.
- Timaru have an Ophthalmologist Khalid Mohammed who can perform ROP checks but the default should be that babies < 28 weeks complete ROP screening at CWH prior to transfer as they are higher risk
- Babies who required screening for ROP should be seen at 6 months of age by the ophthalmologist as they have a higher risk of other eye issues. A referral process has been developed post ROP screening so that this referral is generated once ROP screening has been completed to be seen in 6-12mths in outpatients.
- Outpatient follow-up requests to be sent to Mark Elder for Chch and Ashburton patients, Logan Robinson for West Coast patients and Khalid Mohammed for South Canterbury patients.
- Babies who require laser treatment for ROP should be seen at 3 months by the ophthalmologist
Risk Factors for Eye Disease (apart from ROP)

- Examination of the eyes is an essential component of each check during infancy, and certainly each discharge examination. The examination should include a check that baby:
  - can fix and follow, has a full range of eye movements, has normal pupils and red reflexes (exam requires an ophthalmoscope).

Ophthalmology Consults for Conditions other than ROP

- Complete a consult sheet and fax to 81479 where the referral will be triaged by Dr Antony Beddgood.
- Indicate on the referral our sense of urgency and whether the baby needs to be seen in NICU or if they can travel to the Eye Department (where they can get a more thorough assessment)
- These consults will usually be seen at a separate time to the ROP rounds

Urgent Referrals

- Absent Red Reflex – may be due to a cataract or retinoblastoma. Get a second opinion from an experienced colleague before referring especially in babies with darker skin as the reflex will be different
- Red eye – due to infection or cellulitis (not routine conjunctivitis that is responding to treatment, or sticky eyes that are usually due to blocked tear ducts that resolve over the first year of life)
- Trauma to the eye – not isolated conjunctival haemorrhage
- Symptomatic, unwell baby with CMV or Toxoplasmosis – potential to consider acute treatment

Less Urgent Referrals

- Eye Malformations – colobomas, proptosis
- Abnormal eye movements – nystagmus, not fixing and following
- Work up for septo-optic dysplasia
- Close relative with a history of retinoblastoma or congenital eye abnormalities that may affect vision. Note that 90% of cases of retinoblastoma are sporadic and will not have a family history
- Suspected or confirmed CMV or Toxoplasmosis in an asymptomatic well baby
- Inborn errors of metabolism – risk of cataracts or other ocular signs
In 2009 a national newborn hearing screening programme commenced in Canterbury with automated ABR (aABR) or automated otoacoustic emissions. From July 2015 this has changed so that all babies are screened with aABR and the risk factors for those requiring ongoing hearing surveillance have been modified. For babies in NICU the risk factor sheet should be completed by the nurses, CNS/ANP or Registrars prior to screening (at around 36 weeks corrected gestation or 48 hrs prior to discharge). The risk factor form is then available for the hearing screeners who will make the necessary referral to audiology if required. The timing of the hearing surveillance through audiology will differ depending on the risk factor.

### Risk Factors for Hearing Loss Requiring Surveillance
- Craniofacial anomalies – pinna, ear canal, cleft palate (not an isolated cleft lip)
- Syndromal diagnosis with known association with hearing loss (such as Trisomy 21, CHARGE, Stickler syndrome, Goldenhar, Pierre Robin, Wardenburg, Pendred)
- Proven congenital CMV, Rubella or Toxoplasmosis infection (not herpes or syphilis)
- Ventilated for > 5 days
- Nitric oxide requirement
- HIE Grade 2 or 3 or received cooling
- Ototoxic medications with levels outside the therapeutic range
- Severe jaundice at or above exchange transfusion level – will be rescreened first
- Bacterial meningitis or meningoencephalitis
- Grade 4 IVH with post-haemorrhagic hydrocephalus
- Head trauma

### Specific Situations
- Any baby who does not pass the aABR will be offered a repeat screen and only then referred to audiology for a diagnostic ABR
- Babies with Trisomy 21 will be referred for a diagnostic aABR with audiology even if they pass their screening test
- Babies who have severe jaundice will be screened or rescreened (if already done prior to the jaundice) first and will not be automatically referred to audiology for hearing surveillance
- Babies who develop meningitis after passing the screening aABR will need to be referred to audiology by the Paediatrician
- A family history of hearing loss is no longer a risk factor – these children will all have their hearing screened at the B4 school check

### Reasons for an ENT Referral
- Structural ear and facial anomalies
- Remediable middle ear disease may contribute to the hearing problem.
CARE OF THE INFANT OF THE DRUG USING MOTHER

- The majority of these infants will have mothers who are on the Methadone in Pregnancy Programme (MIPS).
- We have observed that approximately 50% of babies of women on methadone will require treatment for withdrawal.
- The majority of infants requiring treatment will exhibit significant symptoms within the first three days after birth.
- Never give intramuscular naloxone to the infant of a narcotic dependent mother, as it will cause dramatic onset of severe withdrawal symptoms including seizures.
- If baby of a narcotic dependent mother is depressed at birth and maternal narcotics are suspected to be the cause, it is much safer to support respiration.
- Maternal serology for Hepatitis B will have been checked during pregnancy and the results should be available at the time of delivery.
- If mother is HBsAg positive the appropriate protocol for management of the infant is a very high priority (see earlier section).
- If mother's hepatitis B carrier status is unknown, she is in a high risk group and the HBIG/hepatitis B vaccine protocol should also be begun.
- Experience from the MIPS programme indicates that for most of the mothers on the programme are HBsAg negative, but (as in many programmes elsewhere) nearly all have antibodies to hepatitis C, indicating a high likelihood of hepatitis C carriage and risk of intrapartum transmission to the baby.
- Although there is no vaccine or antiserum available to help reduce this risk, we advise bathing of the baby as soon as possible after delivery, and careful adherence to universal precautions.
- Infants at risk of withdrawal should be admitted to the Level 1 Nursery.
- Our current policy is to encourage breastfeeding.
- Maternal hepatitis C infection is not a contraindicated to breastfeeding, although HIV infection is.

Assessment of the Infant at Risk

- Finnegan Scoring should be commenced for all infants at risk.
- Score 4 hourly from birth - review of behaviour after a feed is optimal timing.
- More frequent scoring may be needed if scores are high eg 2-3 hourly.
- When a baby with NAS has 3 consecutive Finnegan scores averaging 8 or more, the baby should be commenced on oral morphine as detailed below.
- Also consider infection, cardiac and/or biochemical abnormalities.

Treatment

- To treat opioid withdrawal we use oral morphine solution (1 mg/ml).
- Babies of mothers on multiple drugs may need phenobarbitone.

<table>
<thead>
<tr>
<th>Finegan Score - should be done 4 hourly</th>
<th>Morphine Solution (1.0mg/ml)</th>
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</thead>
<tbody>
<tr>
<td>Average &gt; or =8 for 3 scores</td>
<td>0.4 mg/kg/day in 4 divided doses</td>
</tr>
<tr>
<td>At least 2 scores &gt; or = 8 despite 0.4mg/kg/day</td>
<td>0.6mg/kg/day in 4 divided doses</td>
</tr>
<tr>
<td>At least 2 scores &gt; or = 8 despite 0.6mg/kg/day</td>
<td>0.8mg/kg/day in 4 divided doses</td>
</tr>
</tbody>
</table>

- If the dose required is greater than 0.8mg/kg/day then the baby must have a cardiorespiratory monitor.
- The dose should be given 6 hourly.
- Once scores have fallen consistently below the treatment level (< 8) the dose of morphine may be reduced every 48 hours. The usual increment that the dose is dropped by is 0.05mls (0.05mg) per dose however at times the decrease will need to be smaller. For example a baby on 0.5mL every 6 hours will be reduced to 0.45mL every 6 hours for the next 48 hours before the dose is reviewed again.
- If infants are assessed as suitable for early discharge and withdrawal at home, they can be discharged if the stabilisation dose is ≤ 0.5mg / dose or they have successfully had a dose reduction without escalation of withdrawal symptoms.
- Once the dose is at 0.05 mg/dose, the treatment can be changed to prn.
- Once the baby has needed no treatment for 48 hours, he/she can be discharged home and all care issues have been addressed.
- Follow up for babies of narcotic dependent mothers should be arranged in the Wednesday morning Clinic at Christchurch Women's Hospital Outpatients.
Babies of Mothers with Mental Illness

General considerations
- Mental illness combined with the stress of post-partum period may impair a mother’s ability to comprehend or retain information and make decisions.
- Communicate with compassion and document clearly. Consider written information and ideally have a support person present when conveying complex or distressing information about their baby.
- Involve the father in discussions about the care of his infant however be mindful that there may be complex relationship issues, as well as legal and confidentiality issues.
- Babies of mothers with mental illness are more likely to have exposure to smoking, alcohol and substance misuse, poverty, social adversity and family violence so there may be care and protection considerations.
- Mothers with mental illness may struggle to bond with their infant and care should support the mother-infant relationship and minimise periods of separation.
- Encourage breastfeeding if no contraindications but support formula feeding or mixed feeding if it is the mother’s informed decision or best way to protect maternal/infant wellbeing. Lack of sleep is a common precipitating factor for a relapse of severe psychiatric illness.

Birth care plans for women with complex needs
- Women living with complex mental illness will have a complex needs birth plan (usually completed by 34 weeks gestation). A copy is placed in a confidential folder in the locked Neonatal ACNM office and should be consulted if her infant needs neonatal assessment.
- A FloView notification and a psychosocial support plan sticker on the front sheet of a woman’s notes alerts practitioners that the woman has a complex needs birth plan.

Poor Neonatal Adaptation and Neonatal Abstinence/Withdrawal
- Poor Neonatal Adaptation is a collection of symptoms seen in up to a third of neonates exposed to psychotropic medications in-utero, mainly SSRIs. Whether the symptoms relate to side effects of the medication or a withdrawal phenomenon is not clear.
- Symptoms include tremors, irritability, lethargy, hyper/hypotonia, poor feeding, tachypnoea, temperature instability, nasal congestion, hypoglycaemia, vomiting and diarrhoea.
- In the majority of infants, symptoms are mild and self-resolve, usually by 72 hours.
- Never assume infant irritability/lethargy/poor feeding/jitteriness is solely due to maternal medications. They could be signs that an infant is seriously medically unwell and the infant must be assessed and investigated appropriately. “Effects of maternal medications” is a diagnosis of exclusion.
- Supportive care should include an explanation of the infant’s symptoms to the mother, support for feeding, swaddling/skin to skin cuddles and pacifier use for settling.
- Neonatal Abstinence/withdrawal phenomena are seen with benzodiazepines and opiate exposure.
- Benzodiazepines and opiates can begin 2-3 days after delivery so a longer observation time is necessary. Finnegan/NAS scoring is only validated for opiate withdrawal and is required for opiate withdrawal in term infants who are usually admitted to Neonatal Unit for observation and management.

Babies Exposed to Lithium
- In utero lithium is transferred freely to fetus across the placenta.
- Lithium has a narrow therapeutic index (therapeutic levels in adults 0.4-1.2mEq/L) and requires close monitoring to avoid toxicity (seen in adults at levels > 1.2mEq/L, no data on infants).
- Lithium can potentiate the effects of muscle relaxants.
- Lithium levels can fluctuate during pregnancy and at the time of birth.
- Babies exposed to lithium in-utero may be at increased risk of Ebstein’s anomaly or other cardiac defects (first trimester exposure) and should have had a detailed fetal anomaly scan.
- Some infants exposed to lithium may be hypotonic and require initial breathing support after birth and the neonatal team should be present at the birth.
- Some infants exposed to lithium may experience a prolonged ‘floppy baby syndrome’ associated with lethargy, poor sucking, tachypnoea, tachycardia, respiratory distress, cyanosis and hypotonia. The neonatal team should do the 24hr check including a pulse oximetry check.
- Babies whose mothers are on lithium at the time of delivery are at risk of lithium toxicity in the immediate postpartum if mother’s levels are high or if they become dehydrated. Extra monitoring and support with feeding may be needed to prevent dehydration and reduce risk of lithium toxicity.
A cord blood lithium level should be taken on all recently exposed infants and reviewed by neonatal team. There are no current recommendations to guide interpretation. Below is a table of suggested actions. However regardless of blood lithium level, any baby who is showing signs suggestive of lithium toxicity (lethargy, hypotonia, poor feeding) should be reviewed by neonatal team and will likely need admission.

<table>
<thead>
<tr>
<th>Lithium level &lt;0.6</th>
<th>Lithium level 0.6-1.2</th>
<th>Lithium level &gt;1.2</th>
</tr>
</thead>
<tbody>
<tr>
<td>- support demand feeding</td>
<td>- ensure adequate hydration</td>
<td></td>
</tr>
<tr>
<td>- do not allow baby to go &gt; 4 hours without a feed</td>
<td>- support feeding</td>
<td></td>
</tr>
<tr>
<td>- do not allow baby to go &gt; 3 hours without a feed</td>
<td>- monitor lithium levels</td>
<td></td>
</tr>
<tr>
<td>- monitor lithium levels</td>
<td>- check renal function</td>
<td></td>
</tr>
<tr>
<td>- consider admission to neonatal unit for cardiorespiratory monitoring</td>
<td>- support hydration</td>
<td></td>
</tr>
<tr>
<td>- monitor lithium levels</td>
<td>- check renal and thyroid function</td>
<td></td>
</tr>
</tbody>
</table>

As lithium can affect thyroid function it is important to ensure all exposed infants have their newborn screening test.

Breastfeeding is not recommended for women on lithium as lithium is excreted into breast milk in variable amounts (up to 42%) and there is a risk of neonatal toxicity, particularly if the baby is unwell or premature. There is also a paucity of data on long term outcomes however there is currently no data to suggest lithium affects neurodevelopment long term. It may be acceptable for babies to have a first feed of colostrum to aid establishment of the intestinal microbiome. Beyond that donor human milk or formula milk is advised.

If a mother decides to breastfeed on lithium the risks and uncertainties need to be carefully explained to the mother, and there needs to be careful clinical oversight by Mothers & Babies Team with infant blood monitoring (blood tests on day 2, 7, 14 and then 3 monthly for lithium level, renal and thyroid function). Additional testing should be done if baby becomes unwell or if maternal levels are high or if maternal dose is increased (allow 5 days before testing for steady state drug level to be reached). If baby stops breastfeeding or is predominantly formula milk fed then this monitoring is not needed. It is important to emphasise that this monitoring does not guarantee the infant will not suffer adverse effects.

KEY POINTS
1. Neonatal team to attend delivery of all babies of mothers on lithium
2. Cord blood to be taken for lithium level in all exposed infants (neonatal team to review result)
3. Close observation of baby after birth with feeding support to avoid dehydration
4. Neonatal team to do 24 hour check (including pulse oximetry check due to increased risk of cardiac defects).
5. Breastfeeding not recommended. If mother chooses to do so blood test monitoring of baby required.

Additional Resources:

New Zealand Formulary (www.nzf.org.nz)
Medsafe (www.medsafe.govt.nz)
Bumps (www.medicinesinpregnancy.org)
MotherToBaby (www.mothertobaby.org)

You can also contact the Mothers and Babies Service Liaison Paediatrician for advice and the Hospital Drug Information Service. There is also a Health Pathways Guideline.
### BABIES < 32 or < 1500GM BIRTH WEIGHT CHECKLIST

<table>
<thead>
<tr>
<th>AGE</th>
<th>ACTION</th>
</tr>
</thead>
</table>
| Day 1        | Attach baby name label to transfusion form Ref:9311  
Start vitamins IV with lipid or orally – Vitamin A and Vitamin D for all and Micelle E if <1250g or <30wk  
Enter baby in eye book if <1250 g or < 30 weeks |
| Day 2-3      | US Head  
Parents receive information regarding Probiotics  
Start Probiotics the day after birth |
| Day 7        | Start oral Vitamin A and Vitamin D for all and Micelle E (if <1250g or <30wks) when lipid stops  
US head routine D 7-10  
Parents receive information regarding HMF  
Start Folic acid if <1500g and will not receive HMF long term |
| Day 14       | US head if significant abnormalities on 1st scan(s) |
| Day 28       | 1st eye exam due if 26-29+6 weeks at birth  
Start Iron (check drug profile for exclusions)  
US head if significant abnormalities on earlier scan(s) |
| Day 42       | 1st eye exam due if <26 weeks at birth  
US head  
1st vaccine (discuss with consultant) not weight dependent  
Start Fe if not already done so |
| >36 weeks gestation | Full examination including hips and eyes (if no ROP check)  
Review nutrition with dietician  
Stop Probiotics at 36 weeks corrected  
Stop Vitamin A if off respiratory support  
Review medications for discharge  
Lung function assessment from 35<sup>th</sup>-36<sup>th</sup> weeks if born <28 weeks  
Late head US if < 28 weeks gestation - discuss timing with SMO  
Review if an echo is required before discharge |
**TUBES FOR LAB TESTS**

- Intranet/Divisions/Canterbury Health Lab/Testing information will provide you with the blood volume and tube required for all tests done at Canterbury Health Labs
- Green tube = Lithium heparinised
- Pink tube = EDTA
- Red tube = Plain tube

<table>
<thead>
<tr>
<th>BLOOD TEST</th>
<th>VOLUME</th>
<th>TUBE</th>
<th>COMMENTS</th>
<th>LAB</th>
<th>TESTED</th>
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</thead>
<tbody>
<tr>
<td>Amino acids</td>
<td>0.6ml</td>
<td>Green or Red</td>
<td>On ice</td>
<td>80118</td>
<td>Mon-Fri</td>
</tr>
<tr>
<td>Ammonia</td>
<td>0.6ml</td>
<td>Pink</td>
<td>On ice</td>
<td>80397</td>
<td>Daily</td>
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<tr>
<td>Caffeine level</td>
<td>0.5ml</td>
<td>Call the lab to send over a Paediatric size non-gel green tube</td>
<td>80322</td>
<td>Tues and Fri</td>
<td></td>
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<tr>
<td>Chromosomes</td>
<td>0.6ml</td>
<td>Green</td>
<td></td>
<td>80881</td>
<td>Mon-Fri</td>
</tr>
<tr>
<td>Chrom. microarray</td>
<td>1-2ml</td>
<td>Pink – 2 full tubes minimum</td>
<td>Do not send if clotted Write clinical details</td>
<td>80881</td>
<td>Mon-Fri</td>
</tr>
<tr>
<td>Cortisol</td>
<td>0.6ml</td>
<td>Pink, Green, Red</td>
<td></td>
<td>80888</td>
<td>Daily</td>
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<tr>
<td>CRP</td>
<td>0.6ml</td>
<td>Green</td>
<td></td>
<td>80397</td>
<td>Daily</td>
</tr>
<tr>
<td>Cystic Fibrosis screen</td>
<td>0.5ml</td>
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<td></td>
<td>80548</td>
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<tr>
<td>Full Blood Count</td>
<td>0.25ml</td>
<td>Pink</td>
<td></td>
<td>80373</td>
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<tr>
<td>Ferritin</td>
<td>0.6ml</td>
<td>Green or Red</td>
<td></td>
<td>80397</td>
<td>Daily</td>
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<tr>
<td>Gentamicin level</td>
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<td>Green</td>
<td></td>
<td>80397</td>
<td>Daily</td>
</tr>
<tr>
<td>Group and Coombs</td>
<td>0.25ml</td>
<td>Pink</td>
<td></td>
<td>80375</td>
<td>Daily</td>
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<tr>
<td>Group and Hold</td>
<td>0.25ml</td>
<td>Pink</td>
<td>Handwrite label</td>
<td>80310</td>
<td>Daily</td>
</tr>
<tr>
<td>Immunoglobulin</td>
<td>0.4ml</td>
<td>Green, Red, Pink</td>
<td></td>
<td>80334</td>
<td>3 x week</td>
</tr>
<tr>
<td>JAUN screen</td>
<td>0.6ml</td>
<td>Green</td>
<td></td>
<td>80397</td>
<td>Daily</td>
</tr>
<tr>
<td>Lactate:Pyruvate</td>
<td>1ml</td>
<td>Special tube</td>
<td>Call 89761 for tube</td>
<td>80118</td>
<td>Mon-Fri</td>
</tr>
<tr>
<td>NEON</td>
<td>0.6ml</td>
<td>Green</td>
<td></td>
<td>80397</td>
<td>Daily</td>
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<tr>
<td>Phenobarbitone</td>
<td>0.6ml</td>
<td>Green</td>
<td></td>
<td>80397</td>
<td>Daily</td>
</tr>
<tr>
<td>Phenytoin</td>
<td>0.6ml</td>
<td>Green</td>
<td></td>
<td>80397</td>
<td>Daily</td>
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<tr>
<td>TSH/T4 at Biochem</td>
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<td>Green or Red</td>
<td></td>
<td>80397</td>
<td>Daily</td>
</tr>
<tr>
<td>TSH/T4 at Endo lab</td>
<td>1.2ml</td>
<td>2 full Red</td>
<td>Venepuncture not heel prick</td>
<td>80848</td>
<td>Mon-Fri</td>
</tr>
<tr>
<td>Vancomycin</td>
<td>0.6ml</td>
<td>Green</td>
<td></td>
<td>80397</td>
<td>Daily</td>
</tr>
<tr>
<td>Vitamin A and E</td>
<td>0.5ml</td>
<td>Red, Green, Pink</td>
<td>Protect from light</td>
<td>80332</td>
<td>Fortnightly</td>
</tr>
<tr>
<td>Zinc</td>
<td>1ml</td>
<td>Green</td>
<td></td>
<td>80317</td>
<td>Mon-Fri</td>
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</table>
NICU Radiology Ordering and Reporting

- A Paediatric Radiologist SMO is available at all times to review films Monday – Friday 0800 to 1700, then they are on call. Contact them via the operators.
- NICU has a priority Xray (Chest and Abdomen) service from a radiographer Monday to Friday 0800-1630. Contact on Pager 5082 for urgent xrays after submitting an electronic order on HCS.
- After 1630 weekdays and on weekends/public holidays contact Pager 8937 after submitting the electronic order. The radiographer comes across from Chch Hospital.
- If the pager is not being answered call 80777 for Chch Radiology or 81854 for ED Radiology for assistance.
- The NICU SMO must be advised when xrays are taken so they can review the films. Ideally this should be electronically by Inteleviewer in hospital or remotely. The NICU SMO must have reviewed the films prior to requesting.
- After hours the Radiology registrar can be asked to review the film. They have varying experience and so direct referral to the Paediatric Radiologist SMO is appropriate and preferably by the NICU SMO if able.
- Out of hours and weekend radiology for other investigations such as US, contrast studies, MRI
  - Consider the highest level of urgency when requesting the test on HCS and follow-up with a phone call. Discuss the timing with the Paediatric Radiology SMO, especially if the Paediatric surgeon also needs to assess eg: upper GI contrast study.
- For further information on radiology reporting of UVC/UAC/Longlines please refer to the Procedures section of the Handbook

Electronic Radiology Ordering

Refer to documents on G:drive/NIC/Neonatal Handbook/Other Guidelines/Electronic Radiology for more detailed information

Principles

- The only time when a paper form can be used is when a baby needs an urgent XRay prior to being allocated an NHI or if the systems are down
- All results will come through electronically even if ordered on paper
- Reg/CNS/NNP to sign off radiology results only for inpatients they are directly caring for
- Inpatient ordering should be linked to the current inpatient encounter
- Outpatient ordering should be done without an encounter as it does not relate to the current admission
- When ordering inpatient tests change the responsible clinician to the NICU SMO on-service so that the report goes to them for viewing ± sign off
- No radiology should be ordered with the LMC as the responsible clinician
- Any outpatient results need to be left for the SMO to view, act on and sign off
- Renal investigations – Ruth Sinclair must always be the responsible clinician, add maternal NHI to the form for ease of reviewing antenatal scans
- Hip US – Matthew Wallenstein must always be the responsible clinician
- If you need to add any information free text in the “Other Booking Information” section

Approval For Radiology Requests and Sign Off

<table>
<thead>
<tr>
<th>NICU service</th>
<th>RMO</th>
<th>CNS-ANP or NNP</th>
<th>SMO</th>
</tr>
</thead>
<tbody>
<tr>
<td>PLAIN XRAYS</td>
<td></td>
<td></td>
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</tr>
<tr>
<td>Chest radiograph with lateral view</td>
<td>Request and Sign off</td>
<td>Request and Sign off</td>
<td>Request and Sign off</td>
</tr>
<tr>
<td>Abdominal xray with lateral or decubitus view</td>
<td>Request and Sign off</td>
<td>Request and Sign off</td>
<td>Request and Sign off</td>
</tr>
<tr>
<td>Skeletal survey</td>
<td>Request after consultation with SMO and Sign off</td>
<td>Request after consultation with SMO and Sign off</td>
<td>Request and Sign off</td>
</tr>
<tr>
<td>NICU service</td>
<td>RMO</td>
<td>CNS-ANP or NNP</td>
<td>SMO</td>
</tr>
<tr>
<td>------------------------------</td>
<td>----------------------------------------</td>
<td>----------------------------------------</td>
<td>-------------------------------</td>
</tr>
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<td>Skull XRay</td>
<td>Request after consultation with SMO and Sign off</td>
<td>Request after consultation with SMO and Sign off</td>
<td>Request and Sign off</td>
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<tr>
<td>ULTRASOUND</td>
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<td>Head Ultrasound</td>
<td>Request and Sign off</td>
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<td>Renal US Outpatient</td>
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<td>Request and Sign off</td>
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<tr>
<td>Hip US Outpatient</td>
<td>Request</td>
<td>Request</td>
<td>Request and Sign off</td>
</tr>
<tr>
<td>Abdominal US</td>
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<td>Request and Sign off</td>
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<tr>
<td>Renal US Inpatient</td>
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<td>Request after consultation with SMO</td>
<td>Request and Sign off</td>
</tr>
<tr>
<td>Spine US</td>
<td>Request after consultation with SMO and Sign off</td>
<td>Request after consultation with SMO and Sign off</td>
<td>Request and Sign off</td>
</tr>
<tr>
<td>US cysts/lumps/collection joints/thyroid</td>
<td>Request after consultation with SMO and Sign off</td>
<td>Request after consultation with SMO and Sign off</td>
<td>Request and Sign off</td>
</tr>
<tr>
<td>Heart</td>
<td>Request on paper radiology form</td>
<td>Request on paper radiology form</td>
<td>Request and Sign off</td>
</tr>
<tr>
<td>INTERVENTION TESTS</td>
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</tr>
<tr>
<td>Barium swallow, small bowel follow-through or barium enema</td>
<td>Request after consultation with SMO and Sign off</td>
<td>Request after consultation with SMO and Sign off</td>
<td>Request and Sign off</td>
</tr>
<tr>
<td>Micturating cystourethrogram</td>
<td>Request after consultation with SMO</td>
<td>Request after consultation with SMO</td>
<td>Request and Sign off</td>
</tr>
<tr>
<td>NEUROIMAGING</td>
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<tr>
<td>CT</td>
<td>Request after consultation with SMO</td>
<td>Request after consultation with SMO</td>
<td>Request and Sign off</td>
</tr>
<tr>
<td>MRI</td>
<td>Request after consultation with SMO</td>
<td>Request after consultation with SMO</td>
<td>Request and Sign off</td>
</tr>
<tr>
<td>NUCLEAR MEDICINE</td>
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<tr>
<td>Nuclear medicine tests</td>
<td>Request after consultation with SMO</td>
<td>Request after consultation with SMO</td>
<td>Request and Sign off</td>
</tr>
</tbody>
</table>
FOLLOW-UP CRITERIA

Christchurch Hospital Paediatric Outpatients Consultant Clinic

Babies followed up by a designated Neonatal Paediatrician at 6 weeks after discharge unless stated otherwise in the multidisciplinary care plan.

- All babies <32 weeks
- All babies <1500g at birth
- Some babies ≥ 32 weeks on home caffeine (refer to respiratory section)
- Seizures or abnormal neurology
- Grade 2 or 3 Hypoxic Ischaemic Encephalopathy
- Congenital abnormalities
- Syndromes
- Congenital infection
- Cardiac conditions – including a patent PDA at discharge
- Metabolic conditions
- Endocrine conditions
- Or at the lead SMO’s discretion

Christchurch Women’s Hospital Neonatal Clinic

Routine NICU Referrals

Timeframe usually 6 weeks after discharge but individualise as needed and try to avoid coinciding with 6 weeks of age when the GP will be seeing them as well. Will be seen by the SMO rostered for the clinic that month.

- 32-32+6 weeks at birth
- Birthweight 1500-1800g
- ≥ 33 weeks or > 1800g with additional concerns such as feeding and growth
- Caloric supplements
- Severe jaundice or haemolysis
- Birth trauma not fully recovered
- HIE Grade 1
- Significant neonatal infection
- Home oxygen – if term and expect to wean within 4 weeks and not fulfilling criteria for Chch Hospital Clinic
- Follow-up of investigation results (time appointment depending on results availability)

Community Referrals

- GP, LMC, Plunket can refer to the CWH Neonatal Clinic up to 6 weeks corrected age
- An appointment will be made usually within 2-4 weeks time.
- The expectation is that 1-2 visits are all that is required but if ongoing follow-up is needed then they will continue follow-up in POPD with the SMO seeing them in the CWH Neonatal Clinic
- Reasons for referral
  - ≥ 33 weeks or > 1800g with new concerns such as (not an inclusive list):
    - Feeding issues
    - Poor growth
    - Developmental concerns
NICU DISCHARGE LETTERS

- Discharge summaries are needed for every baby and are completed on Health Connect South
- There are 2 discharge letter formats – complex and non-complex
- An interim letter for babies who have lengthy stays on the unit should be typed by the level 3 or 2 registrar. These babies will be <28 weeks or have had a complicated course and discharge is not expected for wks
- Copies of letters should go to:
  - GP, LMC, Parents, Outreach Nurses, Plunket
  - Other specialties eg: Fetal Medicine, Obstetrician, Surgeon, Neurosurgeon, Genetics etc depending on their involvement and follow-up of the patient

ARRANGING OUTPATIENT FOLLOW-UP

It is the whole team's responsibility for ensuring that the appropriate outpatient appointments are requested. There are 2 ways for this to occur.

Appointments for Clinics held in Paediatric Outpatients or Ashburton clinic
eg: Neonatal, Endocrine, Paediatric Surgery, Cardiology, Neurology, Gastroenterology, Renal, Infectious diseases, Haematology, Respiratory

- If the clinic, clinician and timeframe is on the discharge letter the NICU Ward Clerk will arrange this with the appropriate booking clerk in POPD.
- If there is not a specific clinician i.e.: any Paediatric Surgeon will do then make that clear on the letter so the booking clerks know just to book it in to the next available clinic in the required timeframe

Appointments for Clinics held outside Paediatric Outpatients
eg: Orthopaedics, Plastics, ENT, Dermatology, Dental, Neurosurgery

- These are mainly surgical specialties.
- It is the responsibility of the Reg/CNS/NNP to ensure that a follow-up appointment has been requested.
- This may be done by faxing a consult sheet to the relevant specialty, phoning the relevant specialty admin team or talking to the medical/surgical team and confirming that they will be booking the follow-up
- Please document on the Multicare Plan that a referral was sent and when
- On HCS you can now check under the patients NHI for what clinics are waitlisted and booked which is extra reassurance that the referral has been received and acted upon
DEVELOPMENTAL FOLLOW-UP

- Babies < 32 weeks can be referred for Early Intervention or Assessment and Monitoring depending on their need. Note, that it is usual for all babies <30 weeks to have some developmental follow-up.
- Babies ≥ 32 weeks and deemed at high risk for developmental delay can be referred on an individual basis for Early Intervention only. For example:
  - Chromosomal abnormalities or syndromal diagnoses
  - Grade 2/3 HIE
  - Neurological abnormalities ie: severe IVH, MCA infarct, hydrocephalus, seizures
  - Complex surgical babies ie: diaphragmatic hernia, oesophageal atresia/fistula, gastroschisis.
- Babies 30 – 34 weeks not needing other developmental follow up will be offered Premie Playgroup
- A decision is made by the time of the MDT discharge planning meeting based on the clinical course, head ultrasound/MRI results and physiotherapy assessments. This is to be documented on the multi-care pathway.
- The chart below describes the various services that provide developmental follow-up in Canterbury.

Each service is set up differently but most services have the ability to have core staff in a role or contract out to other services as required for the following therapists:
- Physio, Conductor (for Conductive Education) OT, SLT, Social Worker, Family Support Worker, Kaiwhakapuawai, Psychology, Early Intervention, Education Support Worker

<table>
<thead>
<tr>
<th>Champion Centre</th>
<th>CCS Disability Action</th>
<th>Ministry of Education and Child Development Service</th>
<th>Conductive Education</th>
<th>Ashburton Child Development Service</th>
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<tbody>
<tr>
<td>Centre based</td>
<td>Home based and Early Childhood Centre visits</td>
<td>Home and centre based and Early Childhood Centre visits</td>
<td>Centre based</td>
<td>Home and centre based</td>
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<tr>
<td>Assessment and Monitoring</td>
<td>Wkly to 4mths corrected then Assessments: 4mth, 8mth, 1,2,3,4 years corrected</td>
<td>Assessments: 4mth, 12mth, 18mth, 2yr, 2.5yr, 3yr, 4yr, 4yr and 9 mths</td>
<td>N/A</td>
<td>N/A Individualised</td>
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<tr>
<td>Early Intervention</td>
<td>Weekly</td>
<td>1-2 weekly as needed</td>
<td>Individualised, PT/OT 2 weekly</td>
<td>2-3 sessions per week</td>
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</table>
**CHILD DISABILITY ALLOWANCE ELIGIBILITY**

The Child Disability Allowance is a non-taxable payment made to the main carer of a child with a serious disability in recognition of the extra care required.

The child must require constant care and attention for at least 12 months because of their disability over and above that of a child of a similar age.

Medical practitioners completing these forms are randomly audited to ensure practice meets these guidelines.

For consistency for our patients the list below explains what conditions make the baby eligible for a child disability allowance. There will be conditions that occur rarely that will not be on this list.

- Syndromes with life-long disability eg: Trisomy 21, chromosomal abnormalities, spina bifida
- Cleft palate, tracheostomy, nasopharyngeal airway
- Chronic renal failure
- Born at <26 weeks gestation as this vulnerable group have increased health needs
- Discharged on home oxygen and enteral feeds by nasogastric or gastrostomy
- Home TPN
- Discharged on home oxygen alone for chronic lung disease of prematurity or known pulmonary hypoplasia for example in association with congenital diaphragmatic hernia.
- Major brain anomalies eg: hydrocephalus, migrational disorders, significant bleeding or clots, Grade 2 or 3 HIE with abnormal MRI findings with the expectation of ongoing disability
- Complex heart disease or in heart failure eg: Tetralogy of Fallot, Transposition, left or right heart hypoplasia, need for surgery in the first year of life
- Chronic skin conditions eg: epidermolysis bullosa, ichthyosis
- Severe musculoskeletal conditions eg: Larsen syndrome, severe contractures limiting mobility

Babies that are not eligible for a child disability allowance include:

- Short-term nasogastric feeding at home
- Short-term oxygen after meconium aspiration
- Uncomplicated small membranous or muscular VSD’s
- High health needs are expected to improve before a year of age
- Referral to Early Intervention alone is insufficient
DRUG PRESCRIBING

- All medications in the unit should be prescribed in a consistent manner to avoid errors in dosing and administration.
- The following format is the preferred prescribing for all drugs and those where the dose is calculated from a formula mg/kg/day.
- If there is a dilution required different to the stock issued by pharmacy this should be documented.
  - Date and Time of Charting
  - Working weight documented
  - Drug NAME in CAPITALS
  - Indication for the drug
  - Concentration eg: (mg/ml)
  - Dose eg: (mg/kg/day)
  - Dose in mg
  - Frequency
  - Specify time period if it is an infusion eg: over 30 minutes
  - If the drug is infused through a pump with guardrail limits then the dose/kg/time period ie: mg/kg/hr, mcg/kg/hr, mcg/kg/min etc is also required (see drug protocols)
  - Prescribers NAME in CAPITALS

Discharge Prescriptions

- These can be generated in Health Connect South with the discharge letter and preferably electronically sent to the pharmacy or alternatively be printed off.
- Some drugs and formulas need Special authority numbers from the Health Benefits centre before a long term prescription can be continued. If in doubt ask the pharmacists.
- Most consultants have access to on-line approval of special authority numbers so there does not always need to be a delay in prescribing
- Dieticians now can obtain special authority numbers for nutritional supplements and arrange the prescriptions
- At the weekly sit down ward round on Thursday discharge planning issues should be undertaken – this realistically begins on moving to level 2. The discharge facilitator or dietician will indicate when this needs to be done.
- The most common discharge medications are Vitamin D and Ferrous sulphate (from 4 weeks postnatal age, if breast fed). Copies of parent information sheets can be printed off from the HCS discharge letter to be handed to the parents at discharge. Do a script for 3 months.
- Micelle E (continued in CF and if Vit E deficiency anaemia), folate and Zinc are usually stopped at discharge.
- If the baby will be on complicated or multiple medications ask the pharmacist to meet with the parents before discharge to review details and give the parents a medication yellow card.

Electronic Prescriptions

- These are the preferred method for prescriptions from July 2020.
- A paper copy does not need to be sent to the pharmacy with the exception of scripts for Class C controlled drugs (codeine and benzodiazepines).
- Morphine still needs to be prescribed in triplicate on paper

Preparation

1. Register with ERMS Online by going to https://erms.health.nz
2. Click Enter
3. Click Forgot Password
4. Enter your username which is your CDHB Username followed by .cdhb
5. Set a password
6. Keep a note of your password safely somewhere as it will be a complicated one
7. Do this now so you don’t have to do it later when you are trying to discharge someone.
8. In HCS - click on User Settings and enter your MCNZ number/Nurse Practitioner number if it is not entered and update preferences
Prescription

1. Arrange with the patient which pharmacy they want to collect their medications from
2. Choose the Canterbury pharmacy from the drop-down menu
3. If the pharmacy is not in Canterbury (covers Chch, Ashburton) or the West Coast then the options are to print the script and give it to the parents or email the script to the out of area pharmacy
4. Enter the medicines on the prescription (the Supply box must be ticked) and click Finish and then click Script
5. When the Print box appears click on the PDFWriter option (if this does not show up then the Service Desk need to add this to the computer. All VDI computers already have this)
6. Click Print and the script will be saved (not actually printed)
7. A Save As box will appear. Change the filename to the NHI and save the PDF on the computer desktop
8. Log on to ERMS online
9. Click Create Referral
10. Select Pharmacology and Pharmacy and then Pharmacy Referral Online
11. Enter the NHI and then click the details will then autopopulate except the phone number which is a mandatory field – either enter the number or enter 0 to bypass the issue
12. Choose the pharmacy from the drop-down box under Service Provider
13. Attach the prescription from the desktop and submit
14. Delete the prescription from the desktop
MULTIDISCIPLINARY CARE PLAN (MCP)

- Multidisciplinary Care Plan replaces the Multidisciplinary Care Pathways that was introduced to the Neonatal Service in March 1999. It is a tool to assist the entire neonatal team with the holistic management of infants and families within the Neonatal Service.
- All infants are commenced on the MCP on admission
- The MCP is designed to be continued throughout the infant's stay in the Neonatal Service, and all disciplines are encouraged to continue their contribution until discharge.
- The plan provides an overview of care received, a record of referrals, a summation of assessments and results, and is also a prompt for management of clinical care and discharge planning.
- It is divided into 5 sections: Admission, Parental Information/Teaching, Clinical Care, Referrals and Assessments, Discharge Planning.
- Clinic follow-up plans can and should be documented on the last page and can be referred to when completing the discharge letter to ensure the baby gets seen at the right clinic in the right timeframe
- The MCP is easy to complete, a date and signature against the relevant information when the task has been completed is all that is requested in most cases.
- Of particular note for medical clinicians are sections within clinical care. Setting of parameters such as oxygen saturations and blood pressure mean are within this section as is the checklist for Babies < 32 weeks or < 1500gms
- Documentation in both the clinical notes and on the MCP may still be required in some instances. For example, expansion on discussion in Multidisciplinary meetings or results of infant assessments. Documentation in the MCP allows people to locate more quickly within the clinical notes details of these.
- Try not to be offended if nursing staff remind you of your duty to complete the MCP. They have been asked to do this.
- Documentation is very important. “If it has not been signed – it has not been done” is the presumption